

THE CORE FACILITY FOR INTEGRATED MICROSCOPY



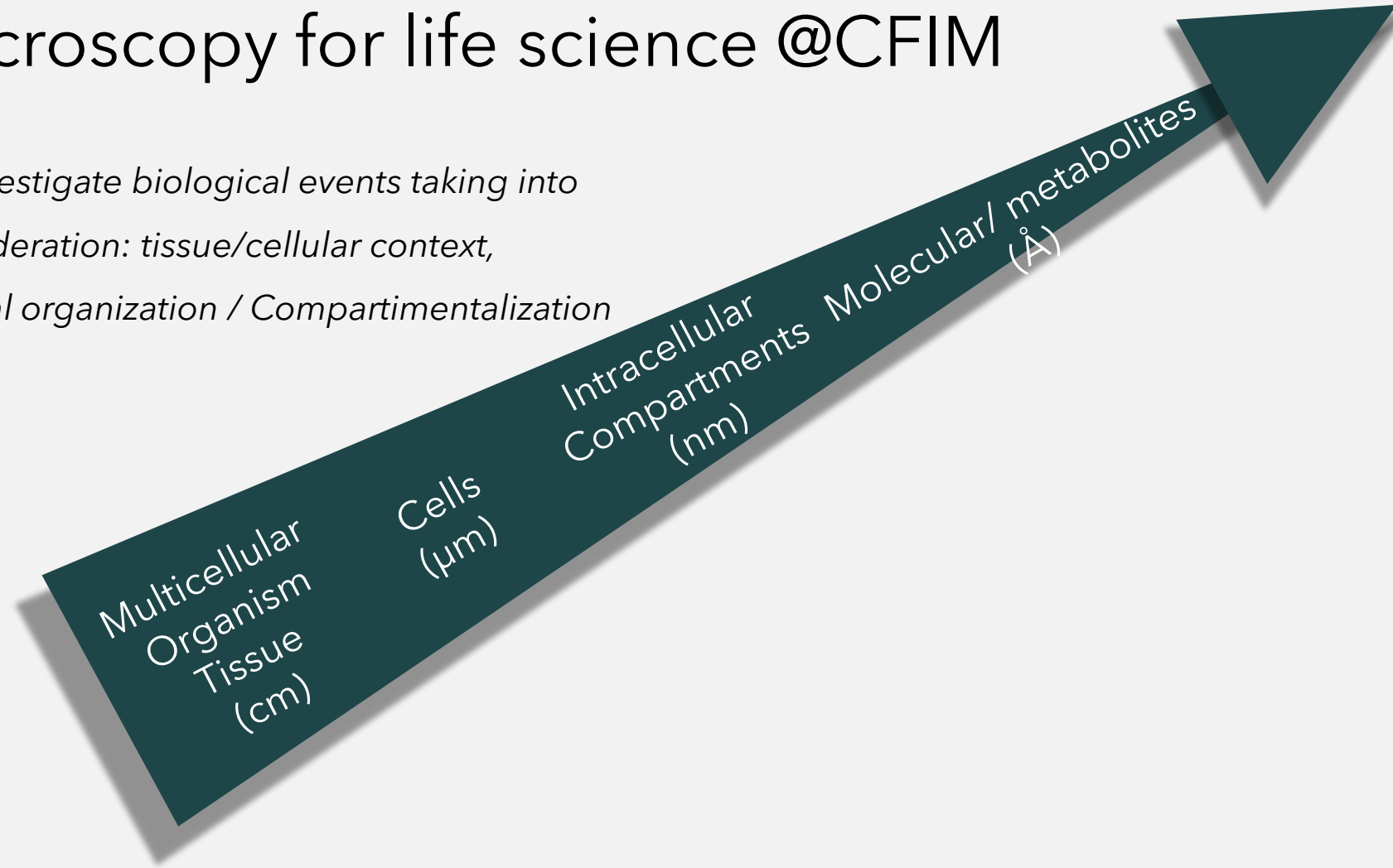
Faculty of Health and Medical Sciences, University of Copenhagen

THE CORE FACILITY FOR INTEGRATED MICROSCOPY

Microscopy for life science @CFIM



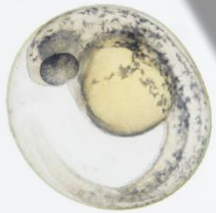
*To investigate biological events taking into
consideration: tissue/cellular context,
Spatial organization / Compartmentalization*



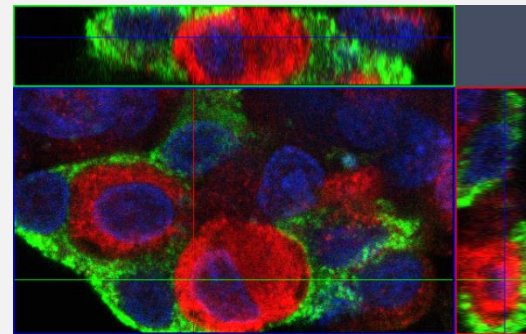
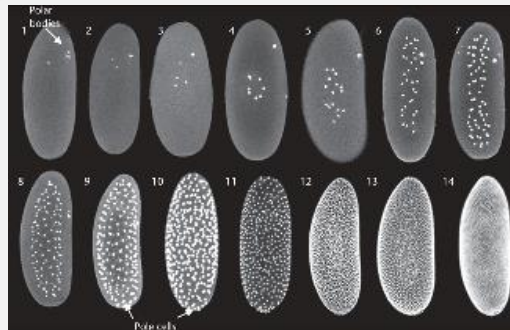
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Multicellular
Organism
Tissue
(cm)



Organism development / Cell migration /
Tissue organisation / Pathology

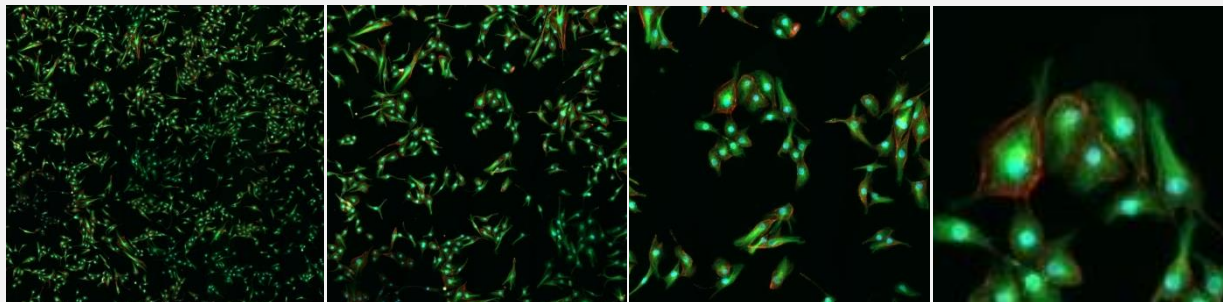
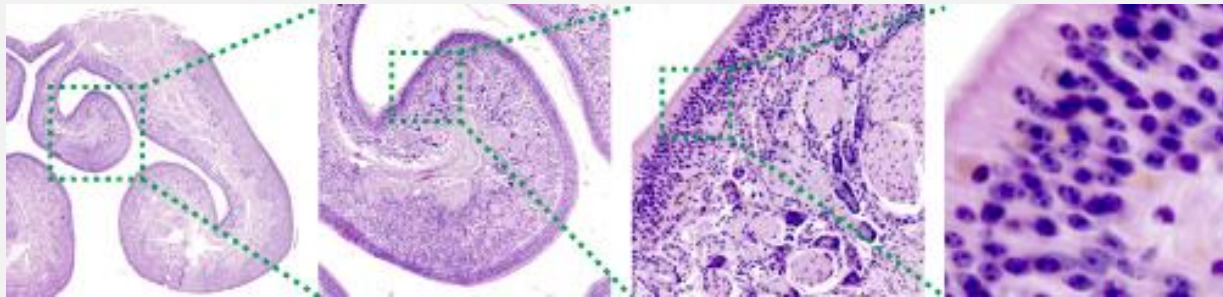


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Digital Pathology

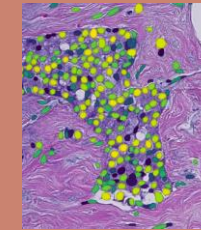


➔ Large amounts of image-based data

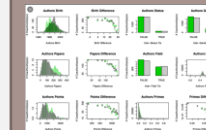


Machine- / Deep- learning
Based pixel classification

Automatic
segmentation
of objects

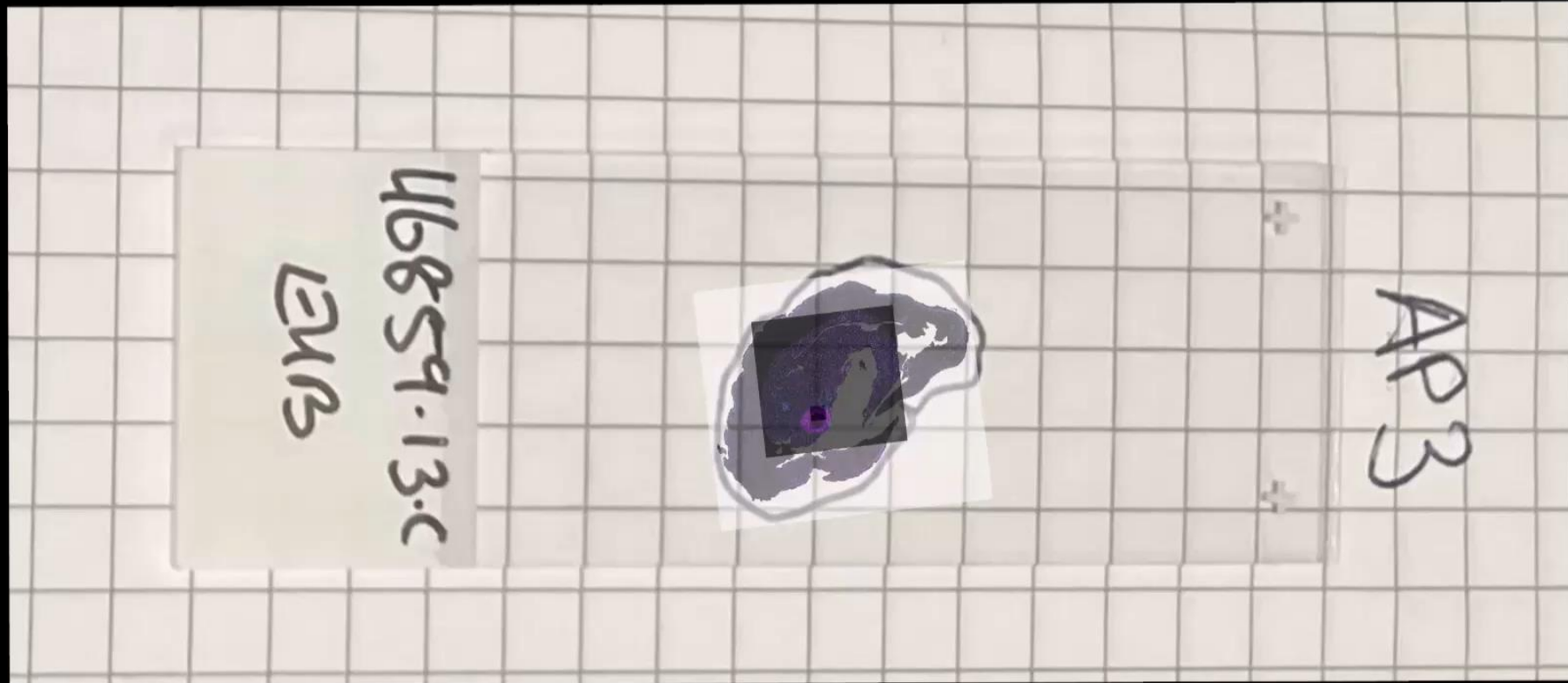


Automatic
segmentation
of objects



Unbiased
Quantitative data

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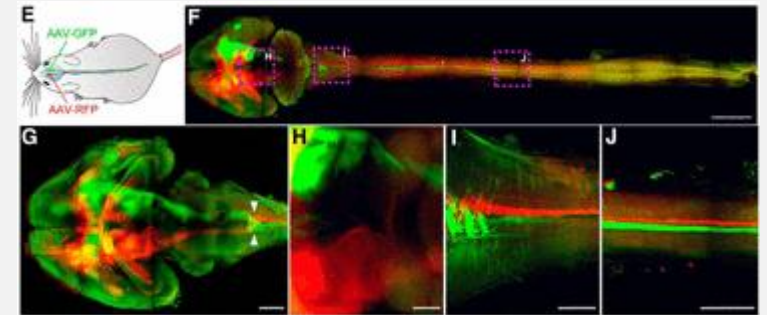
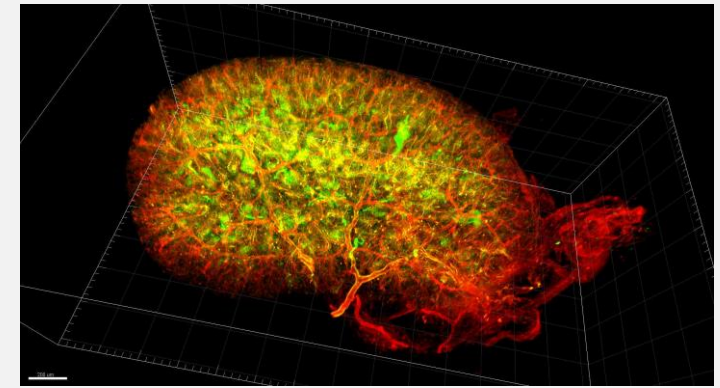
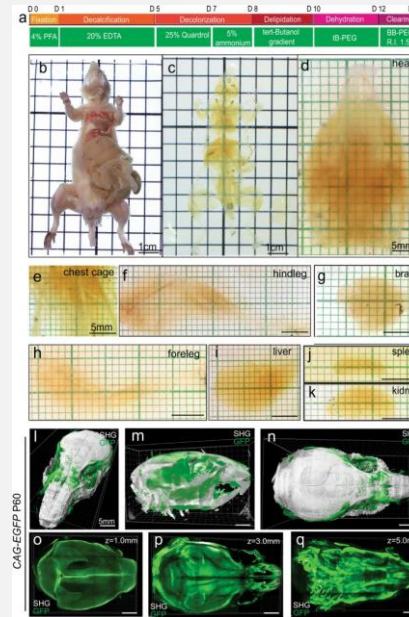
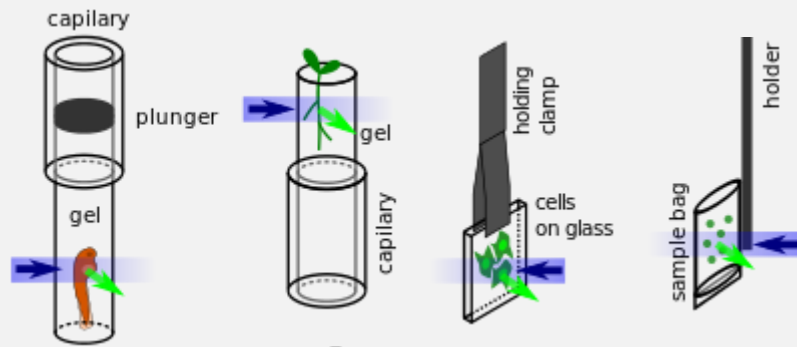
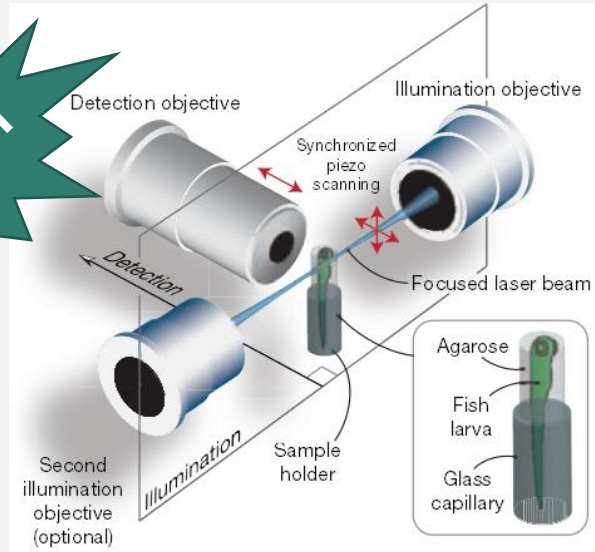


Courtesy of Sophie Amalie Blirup-Plum

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Selective Plane Illumination Microscopy Light sheet Microscopy (live and cleared)

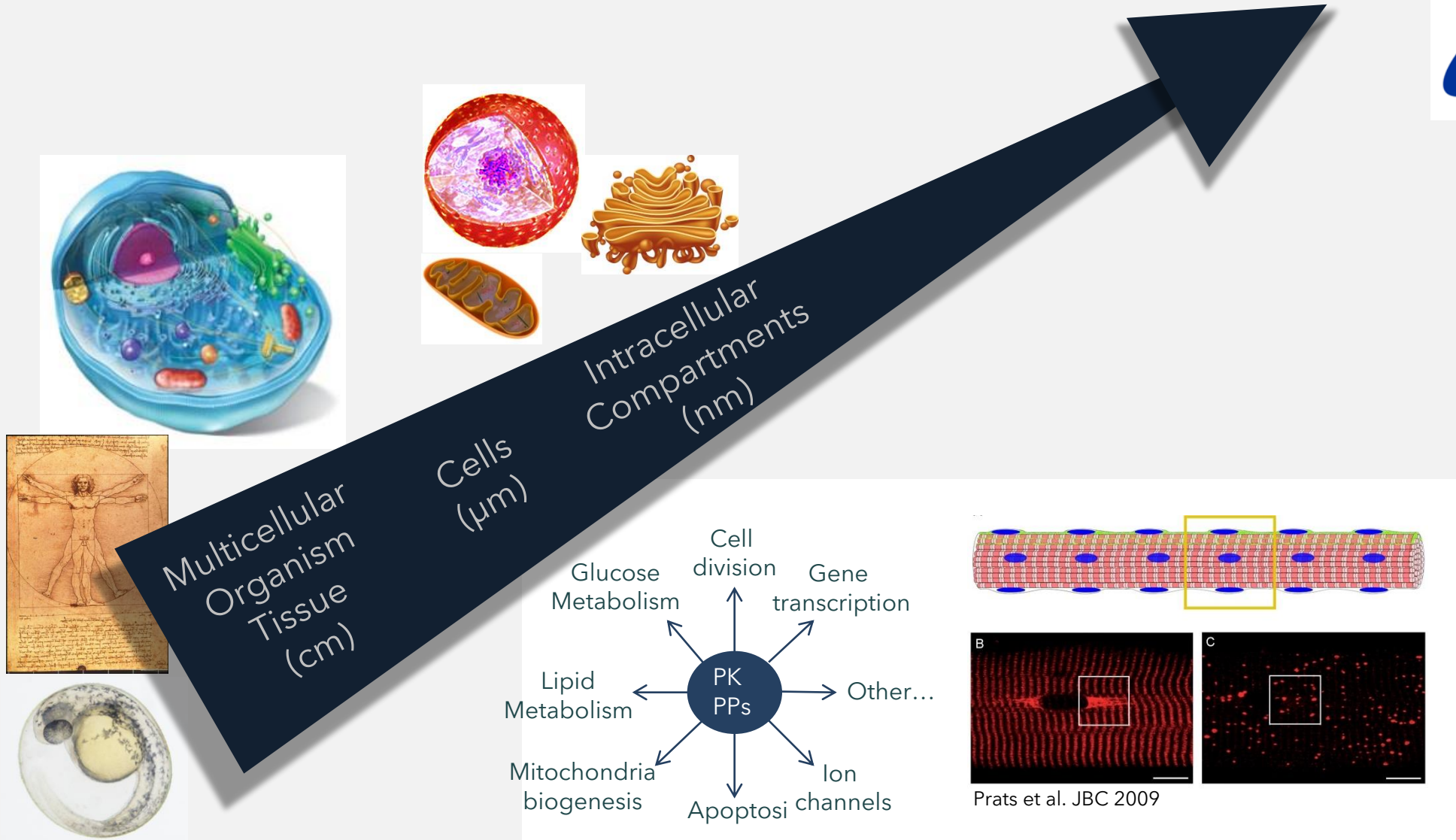
**Relatively
NEW
From 2021**



Mano et al 2018

**NNF infrastructure
Bio DEEP**

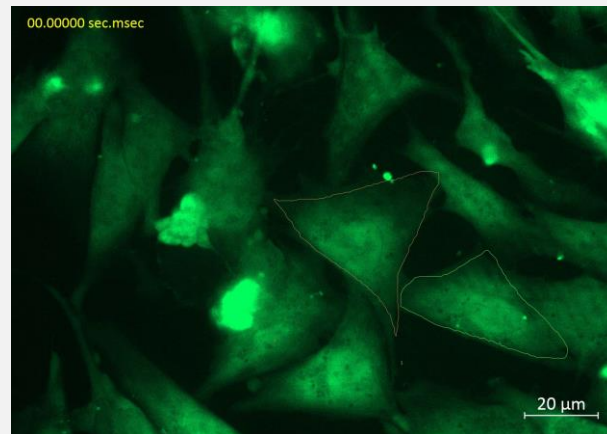
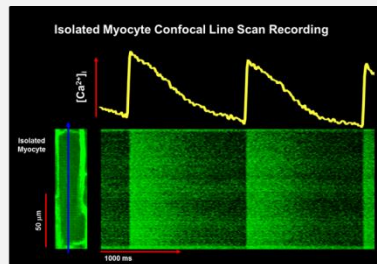
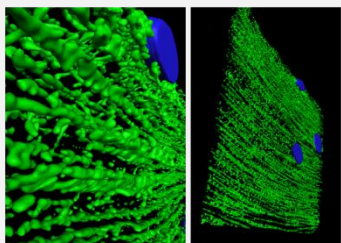
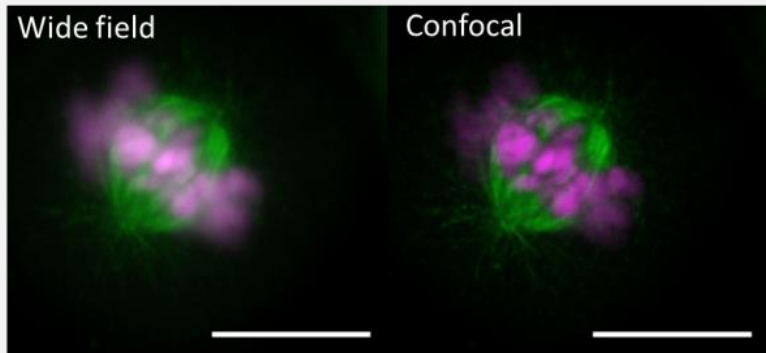
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THE CORE FACILITY FOR INTEGRATED MICROSCOPY

- Cell behaviour
- Dynamics of Intracellular processes
- 3D imaging of intracellular structures

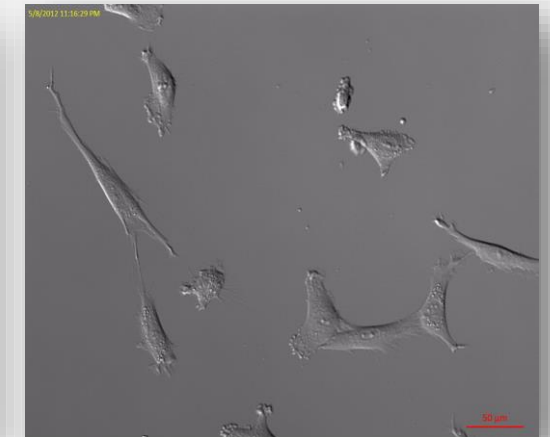
Widefield fluorescence Microscopy Laser Scanning Microscopy



Very fast

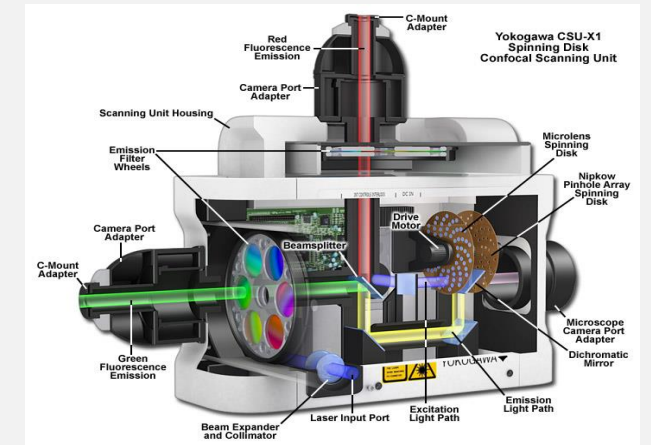


Very Slow

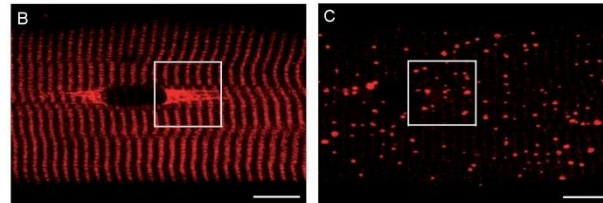
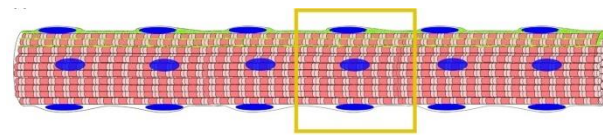
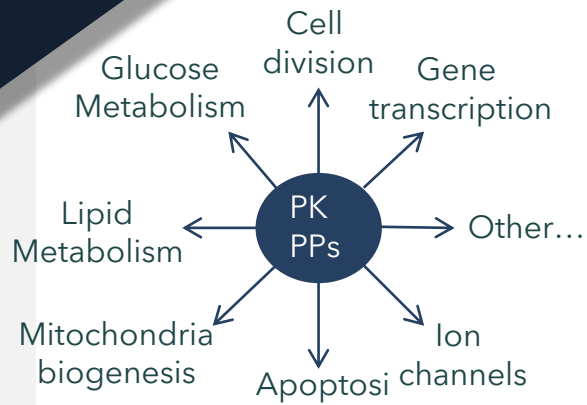
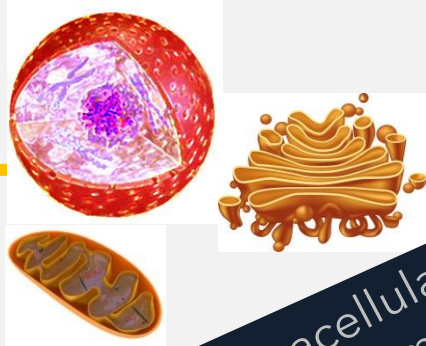
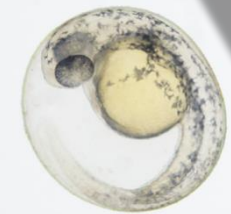
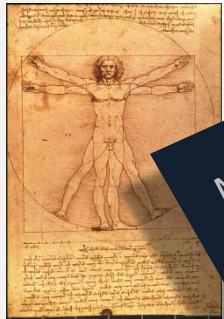
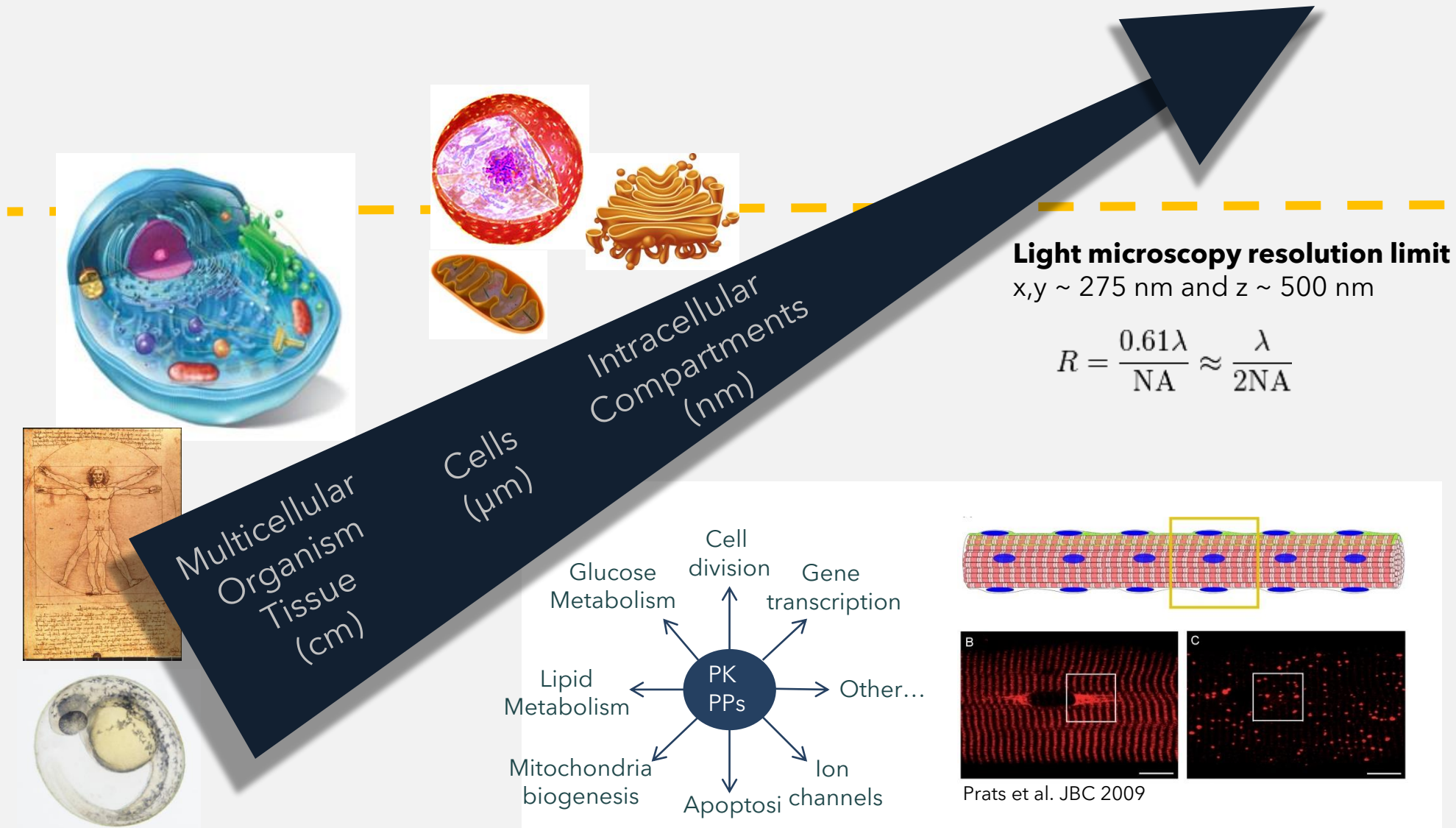


Cell division (~ 9hours - app. 0.1 Hz) by Vibe Nylander

Spinning disk



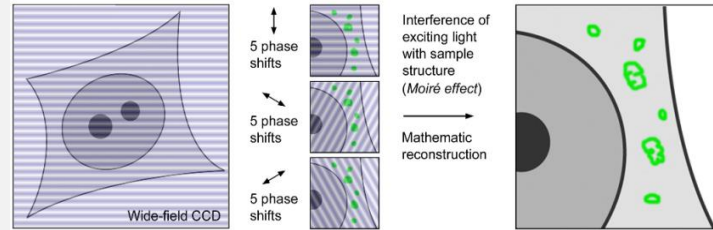
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Prats et al. JBC 2009


THE CORE FACILITY FOR INTEGRATED MICROSCOPY

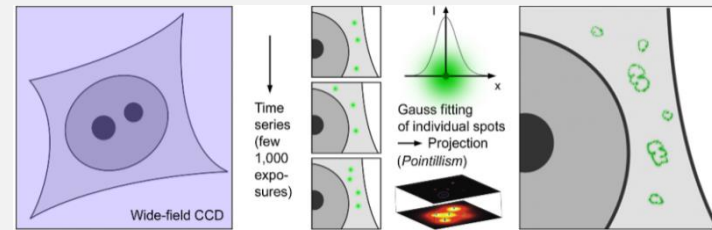
- Structured Illumination Microscopy



- Single Molecule Localization Techniques

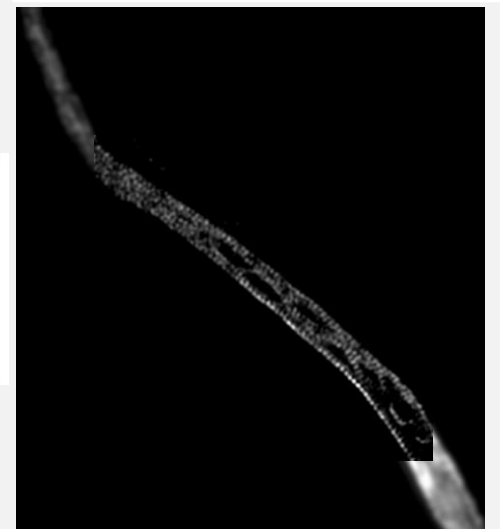
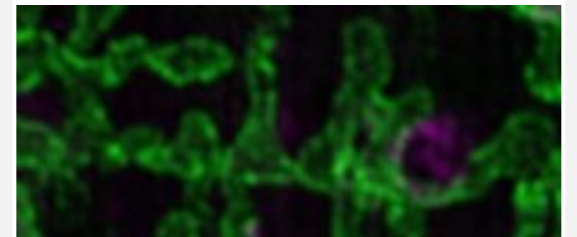
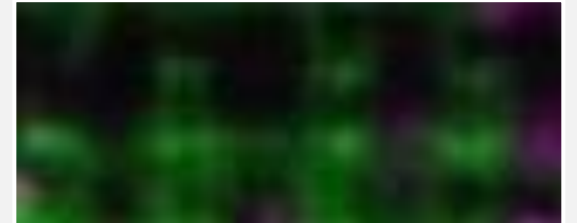
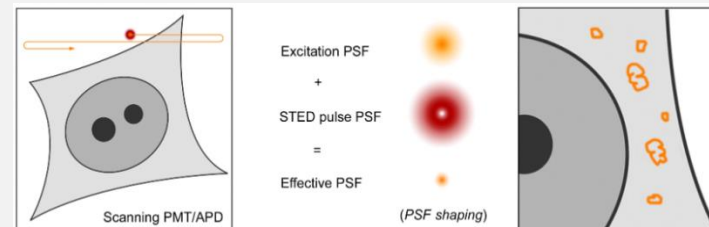
Eric Betzig 


Moerner 



- Stimulated Excitation Depletion - STED

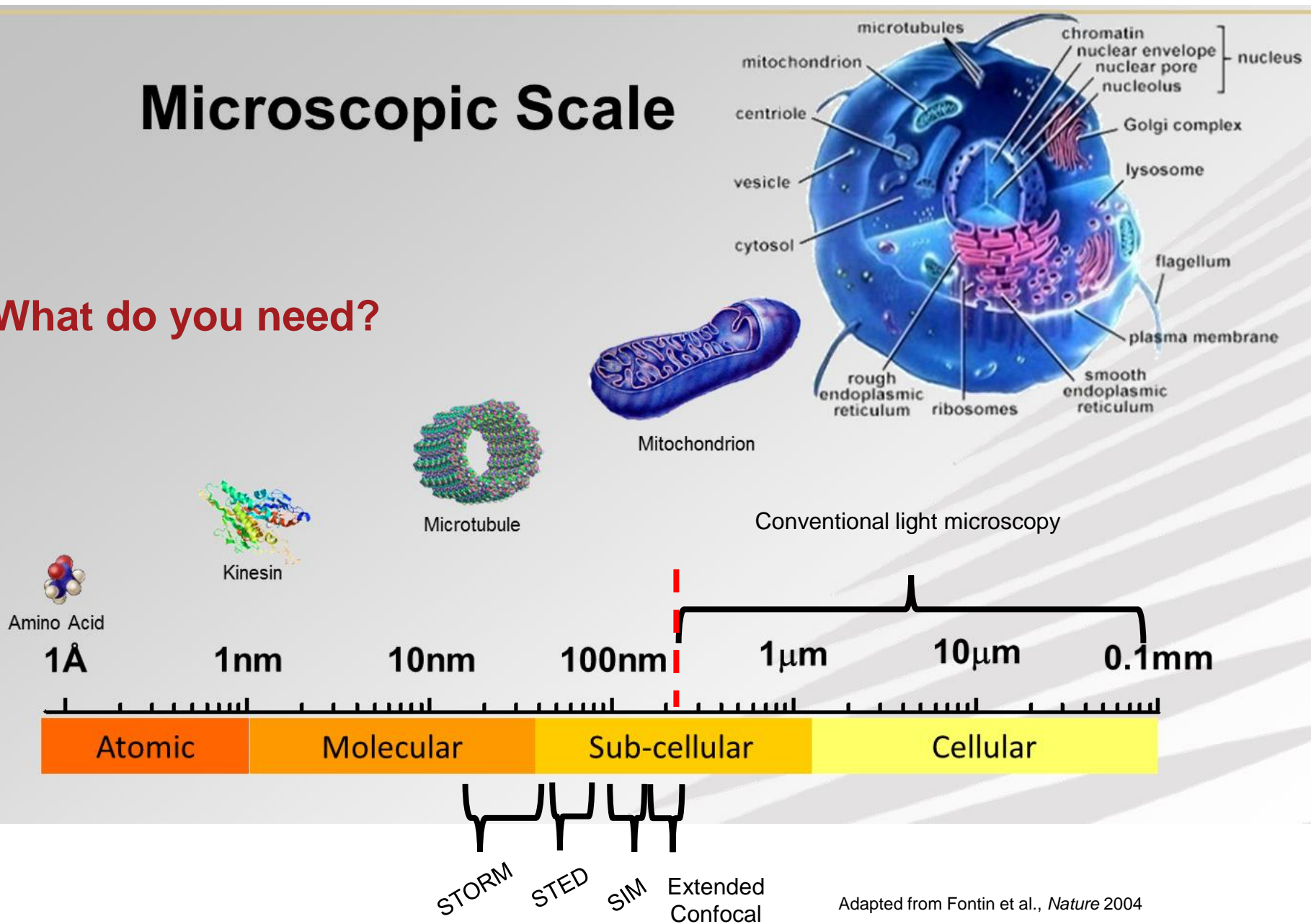
Stefan Hell 



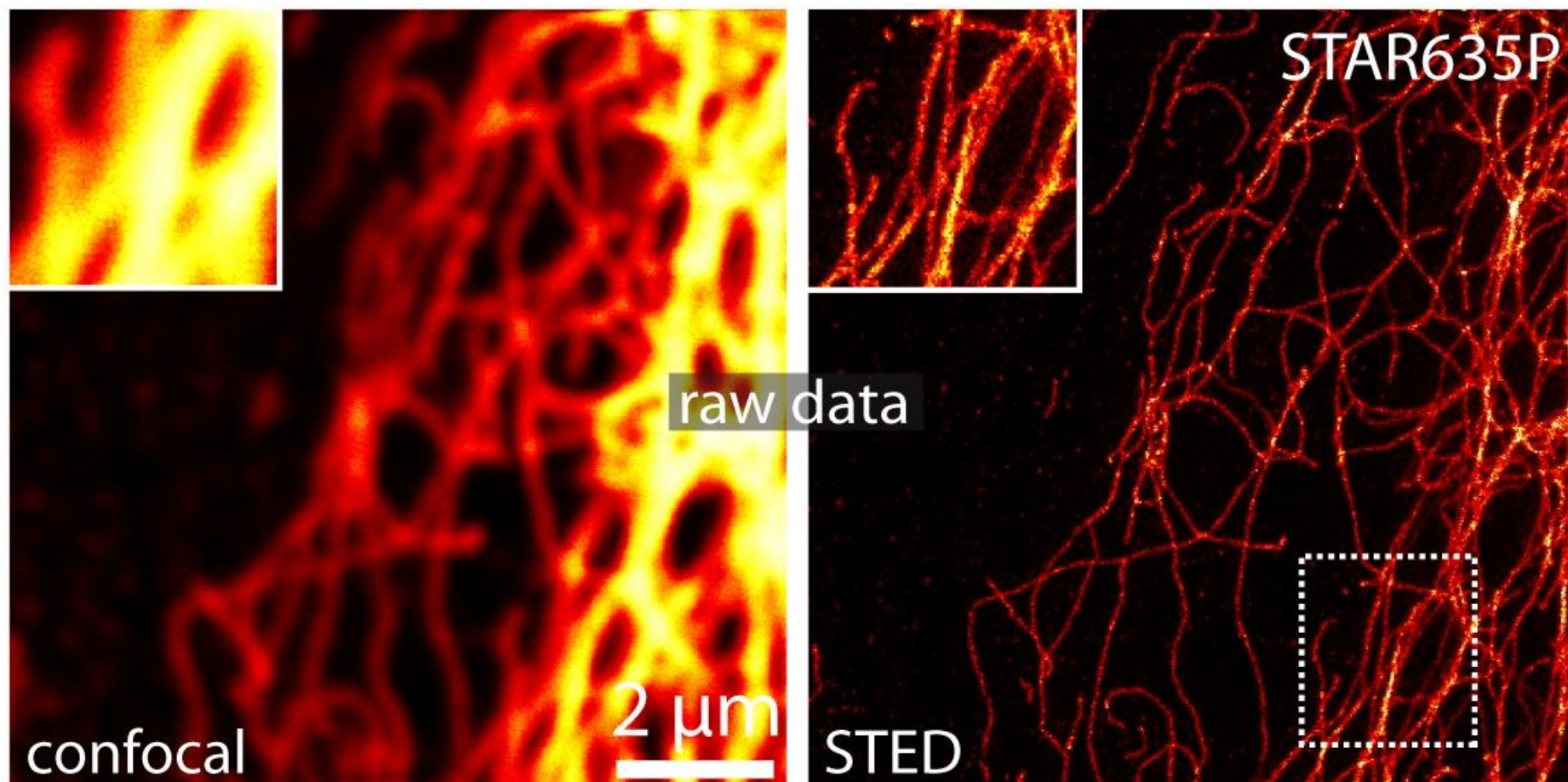
 Nobel prize in Chemistry 2014

Microscopic Scale

What do you need?



STED imaging example

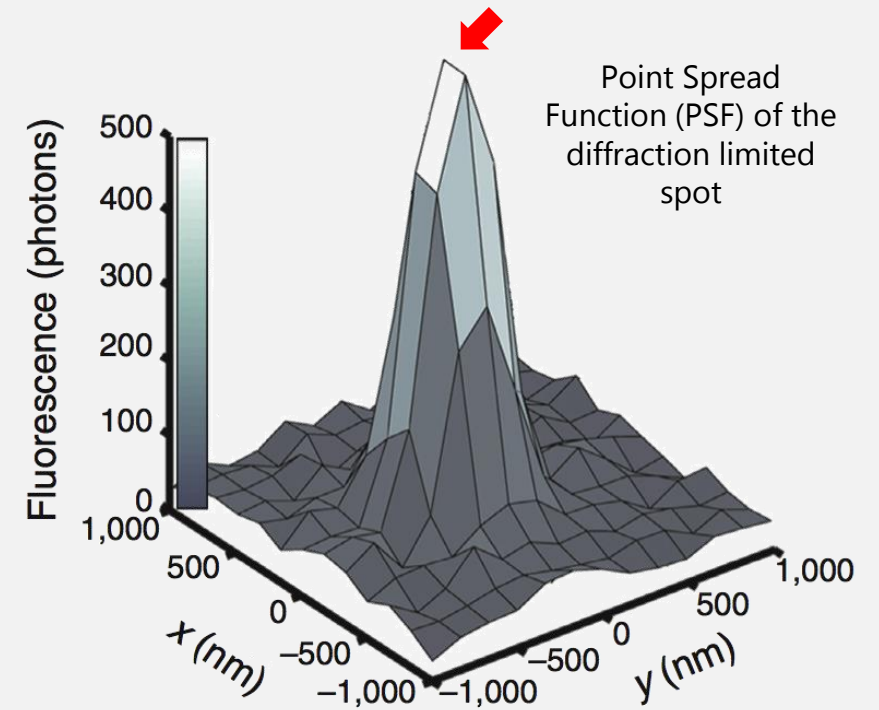
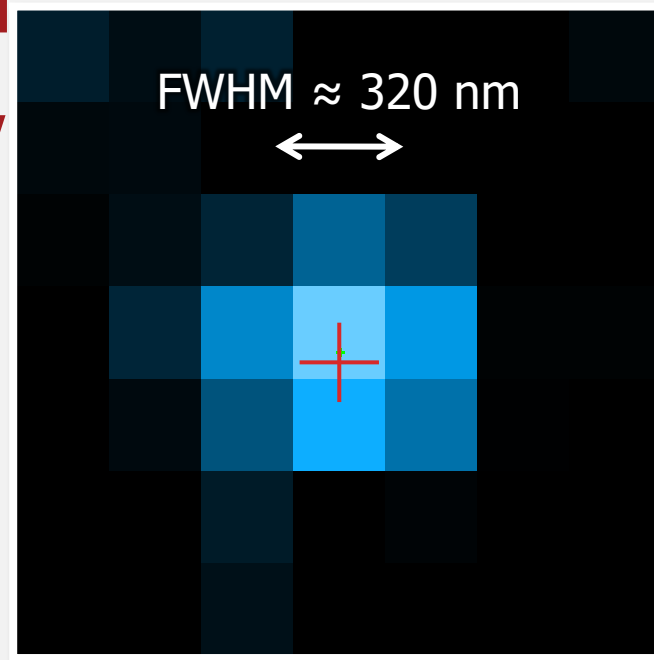


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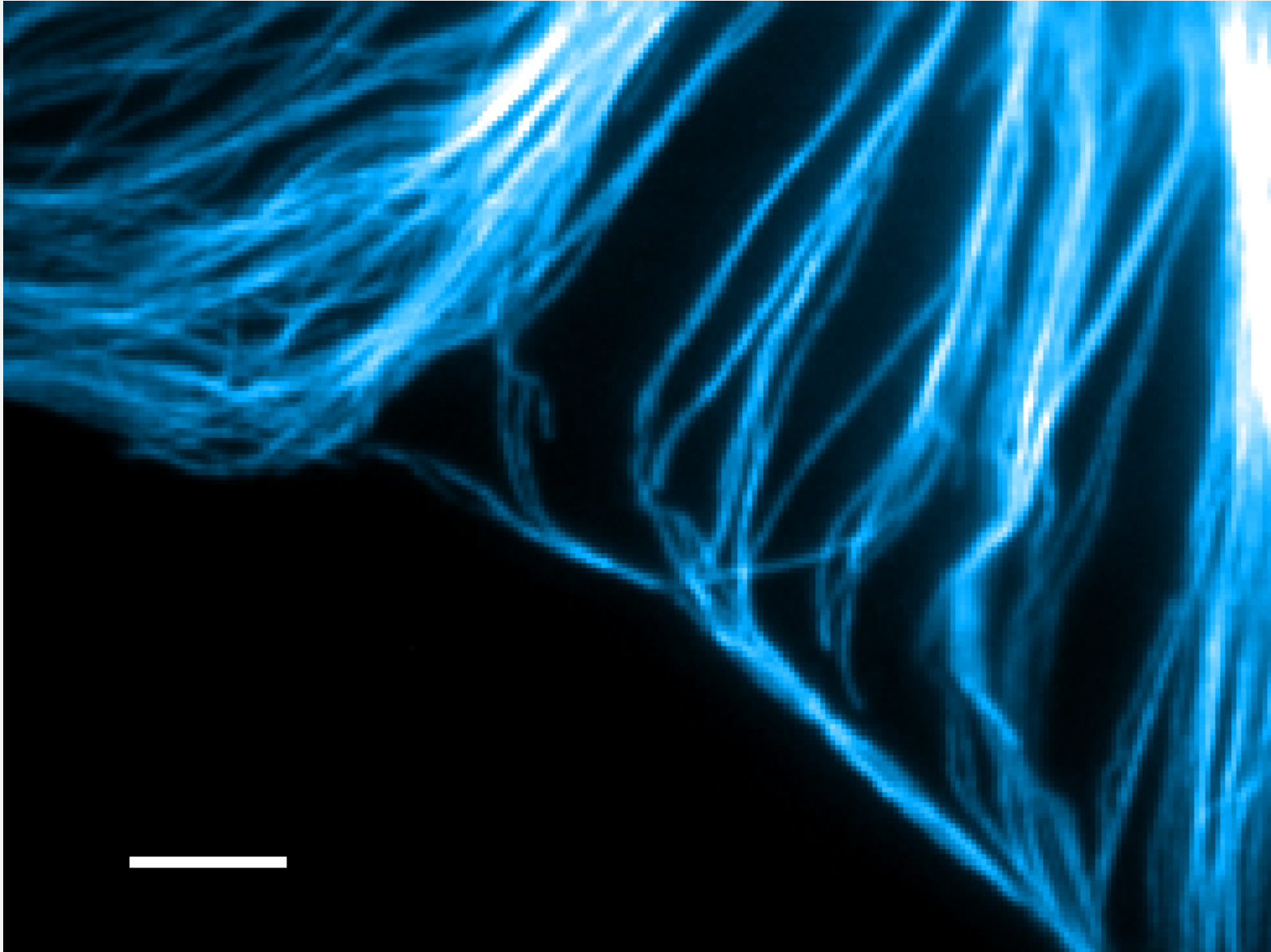
Single molecule localization microscopy

Single-Molecule Localization: Gaussian fitting

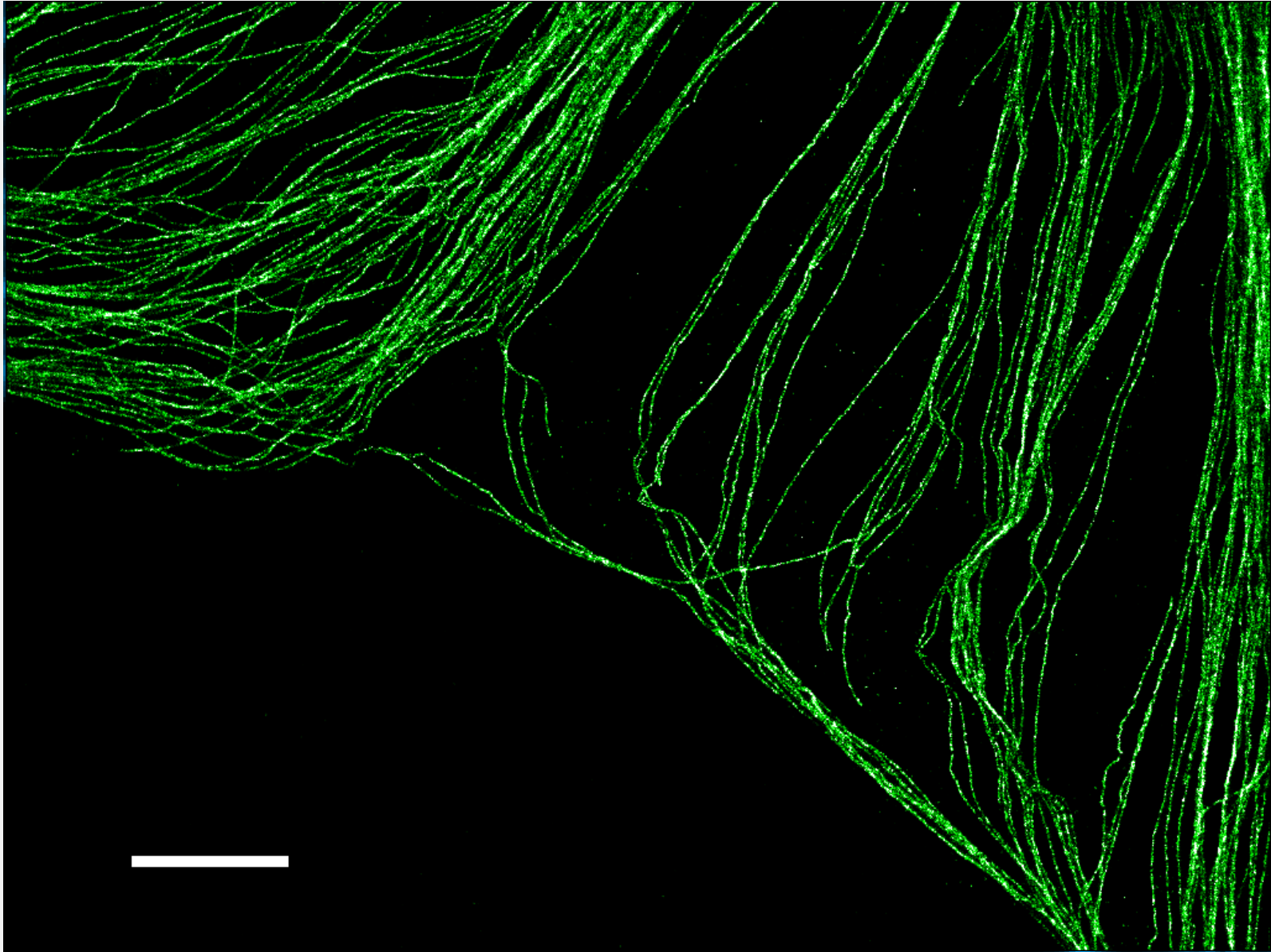
Diffraction limited blinks



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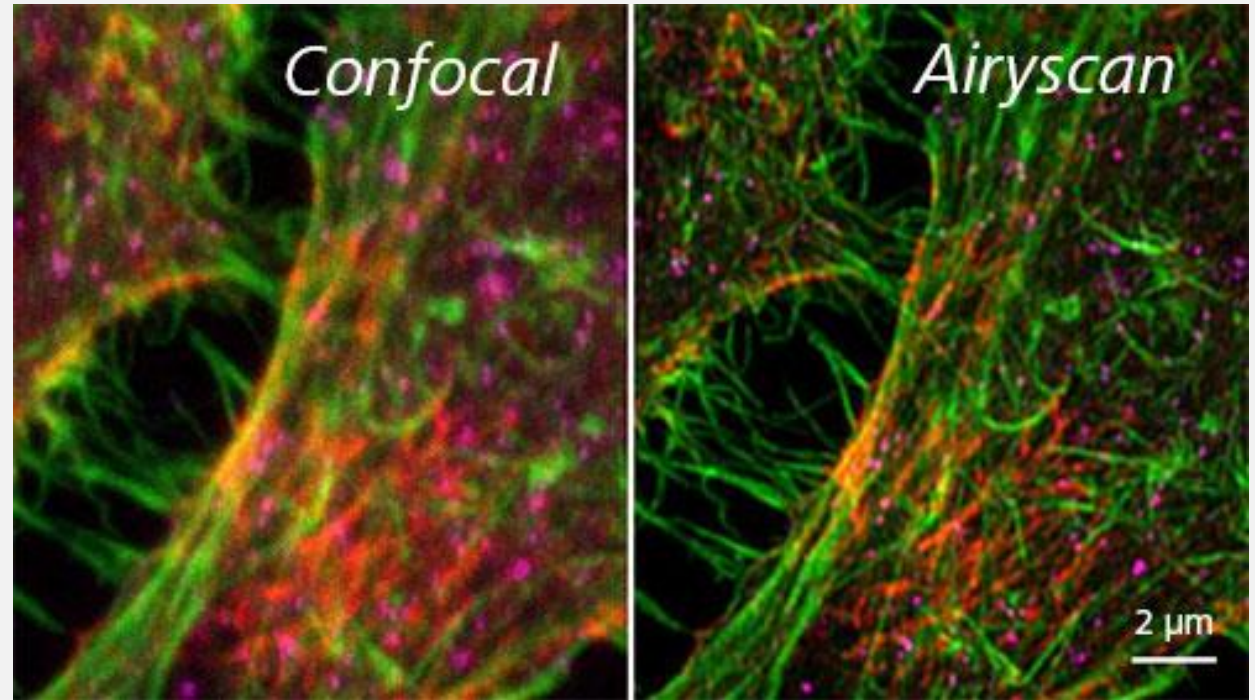
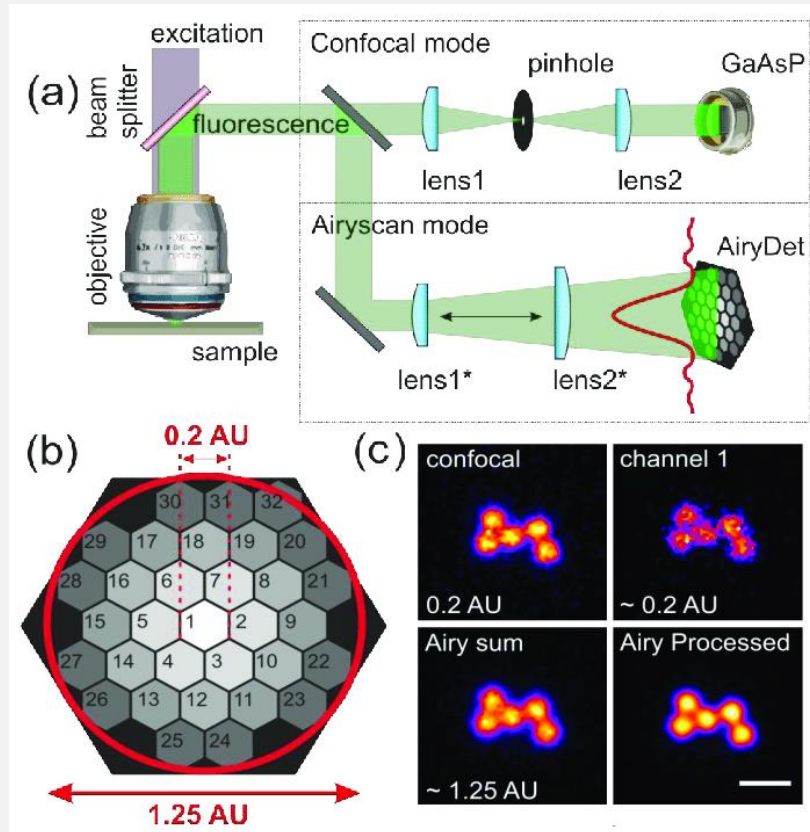
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Airyscan microscopy (LSM900 and LSM980)



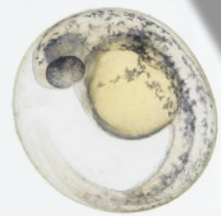
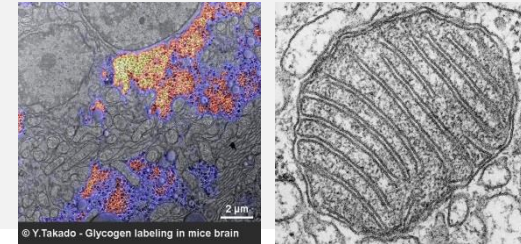
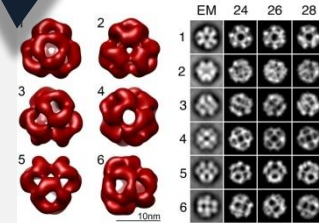
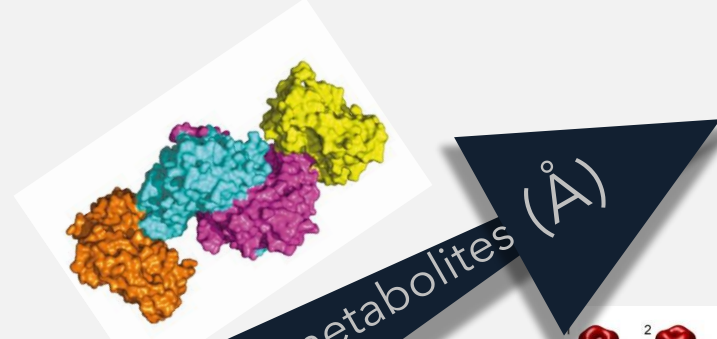
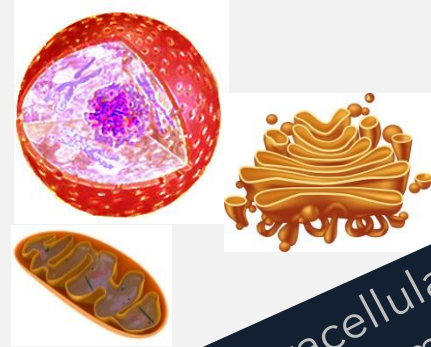
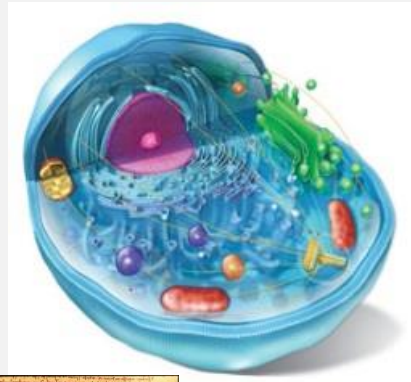
Korobchevskaya et al.

THE CORE FACILITY FOR INTEGRATED MICROSCOPY

Electron Microscopy

+ Ultrastructure resolution

- Static

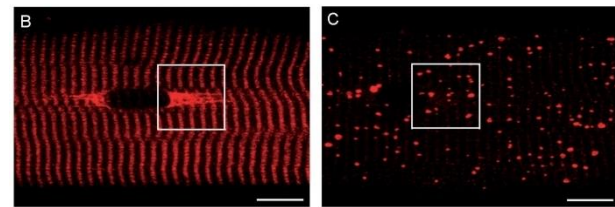
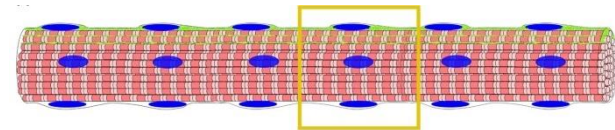
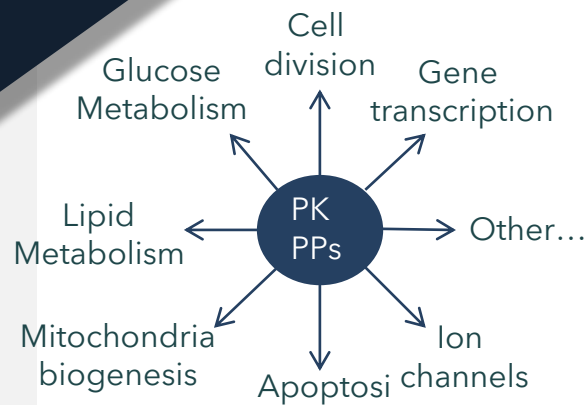


Multicellular Organism Tissue (cm)

Cells (μm)

Intracellular Compartments (nm)

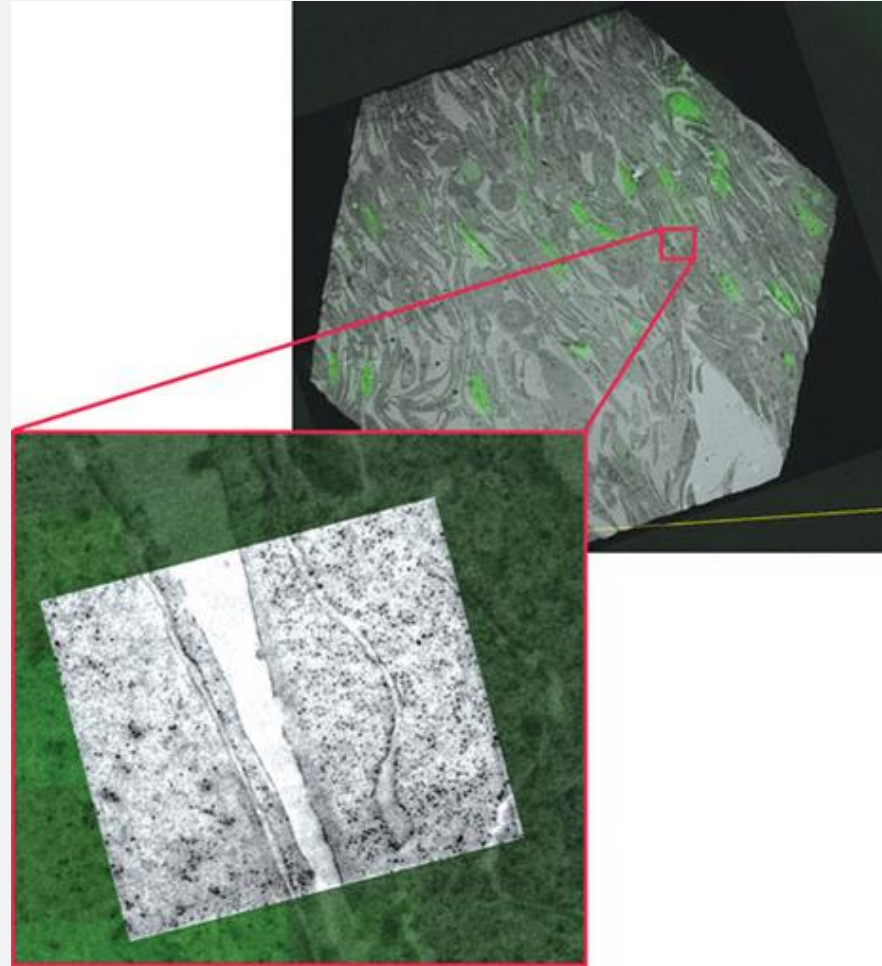
Molecular/ metabolites (\AA)



Prats et al. JBC 2009

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CLEM - Correlative Light and Electron Microscopy

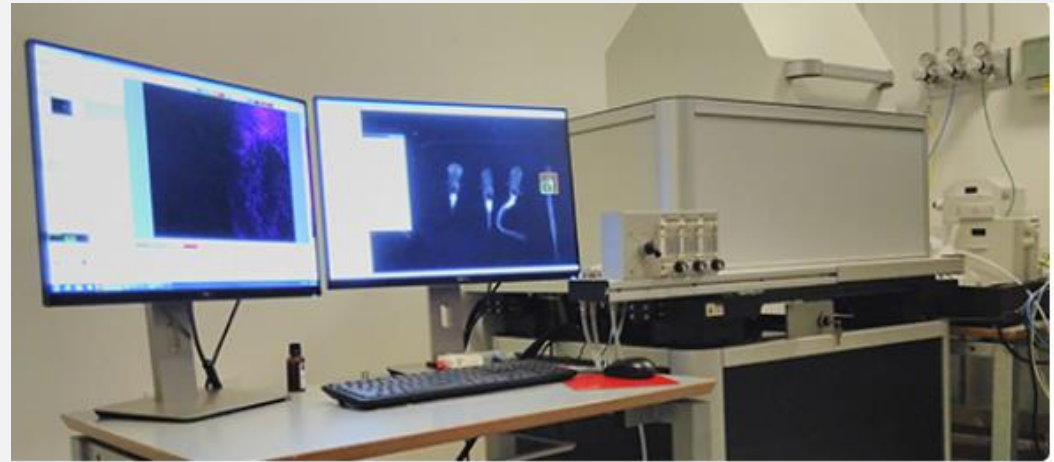
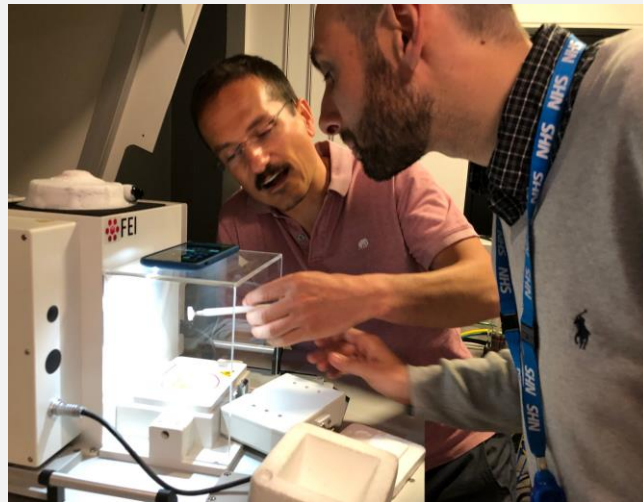
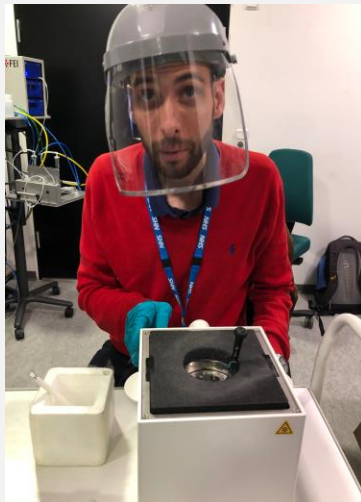


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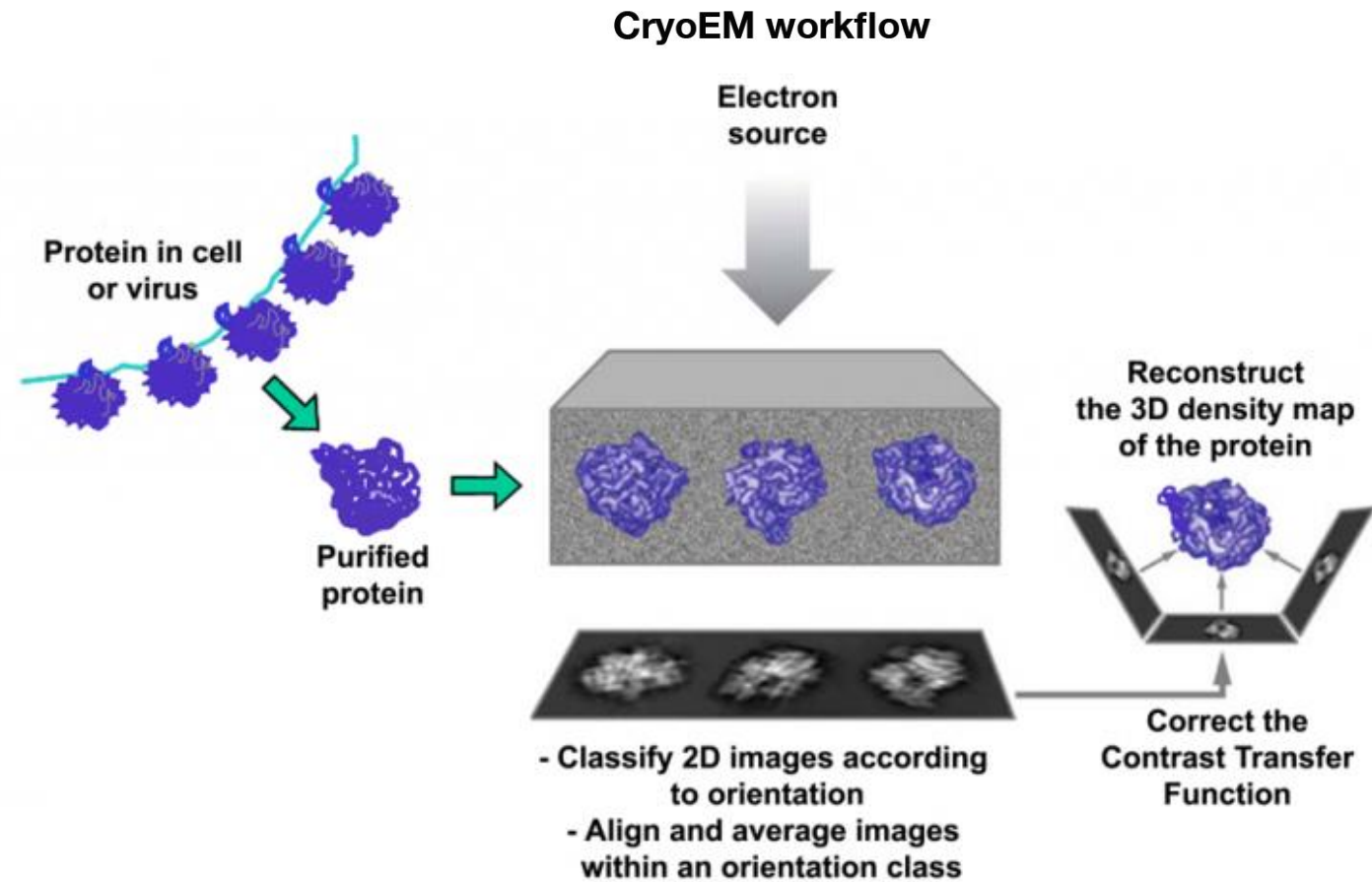
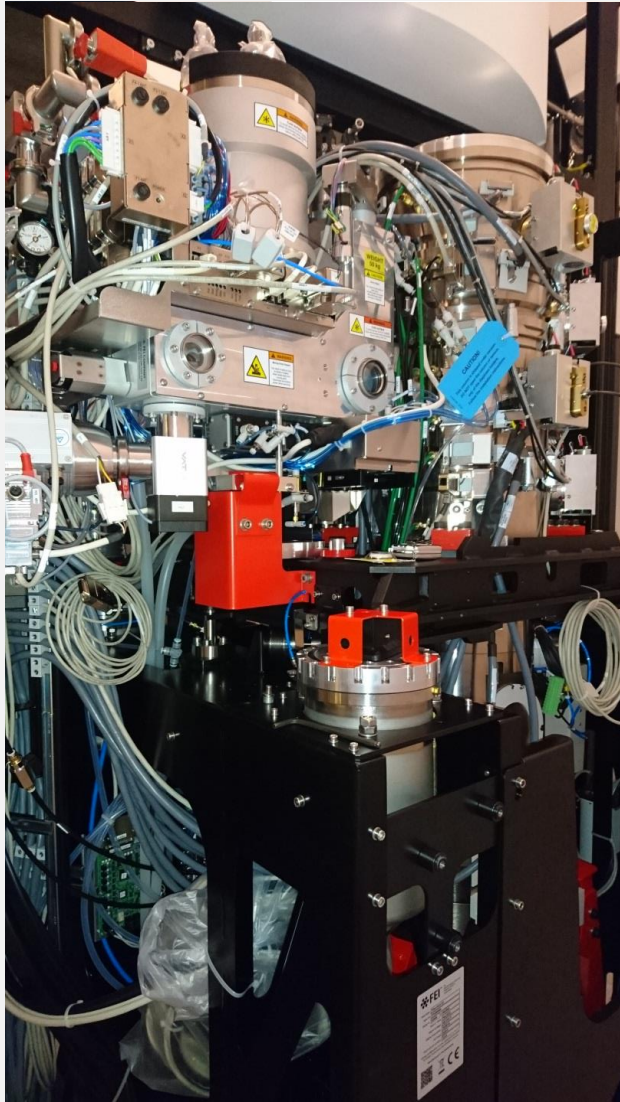


FEI (ThermoFisher) CorrSight

The CorrSight is a light microscope built for **live microscopy** as well as **correlative light and electron microscopy** experiments. It's run by the MAPS software, which is also present on some of CFIM electron microscopes, allowing registration between the images recorded in electron and light microscopy.

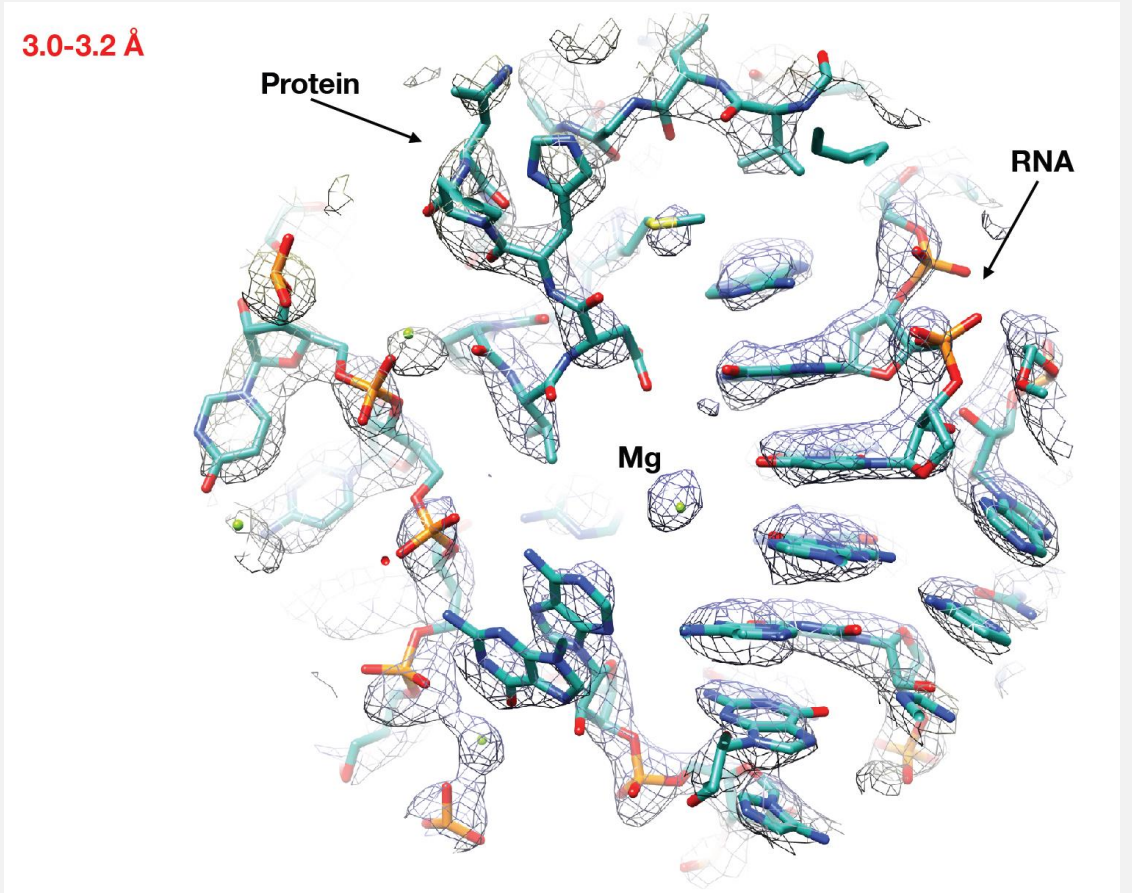
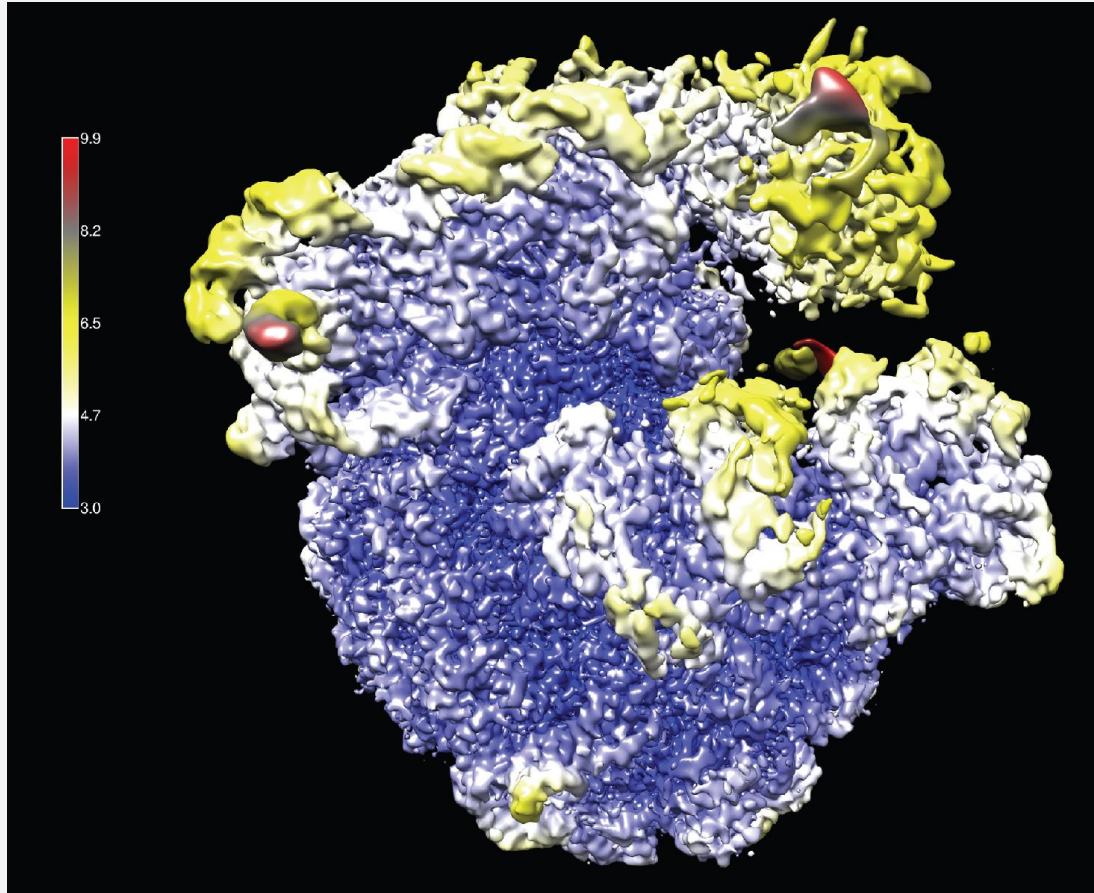


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70S structure

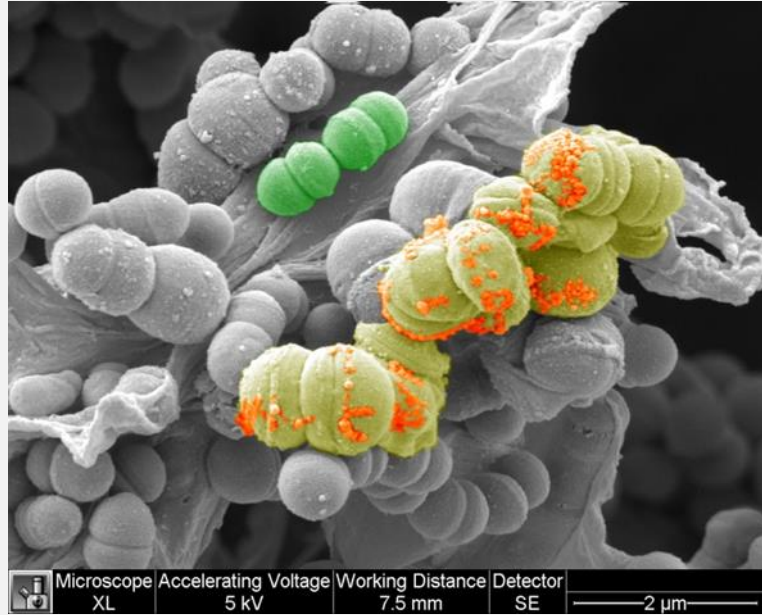


Courtesy of Pablo Mesa and Guillermo Montoya

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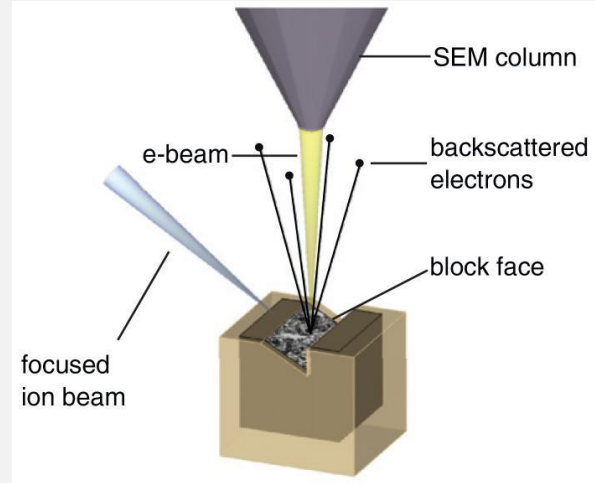
Scanning Electron Microscopy (SEM)

Topography

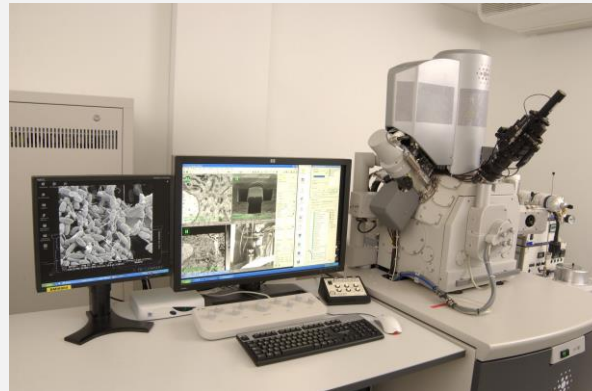
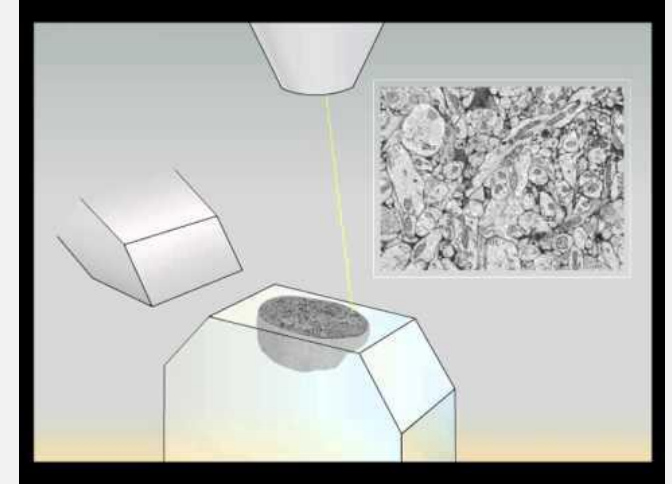


Serial Block face SEM

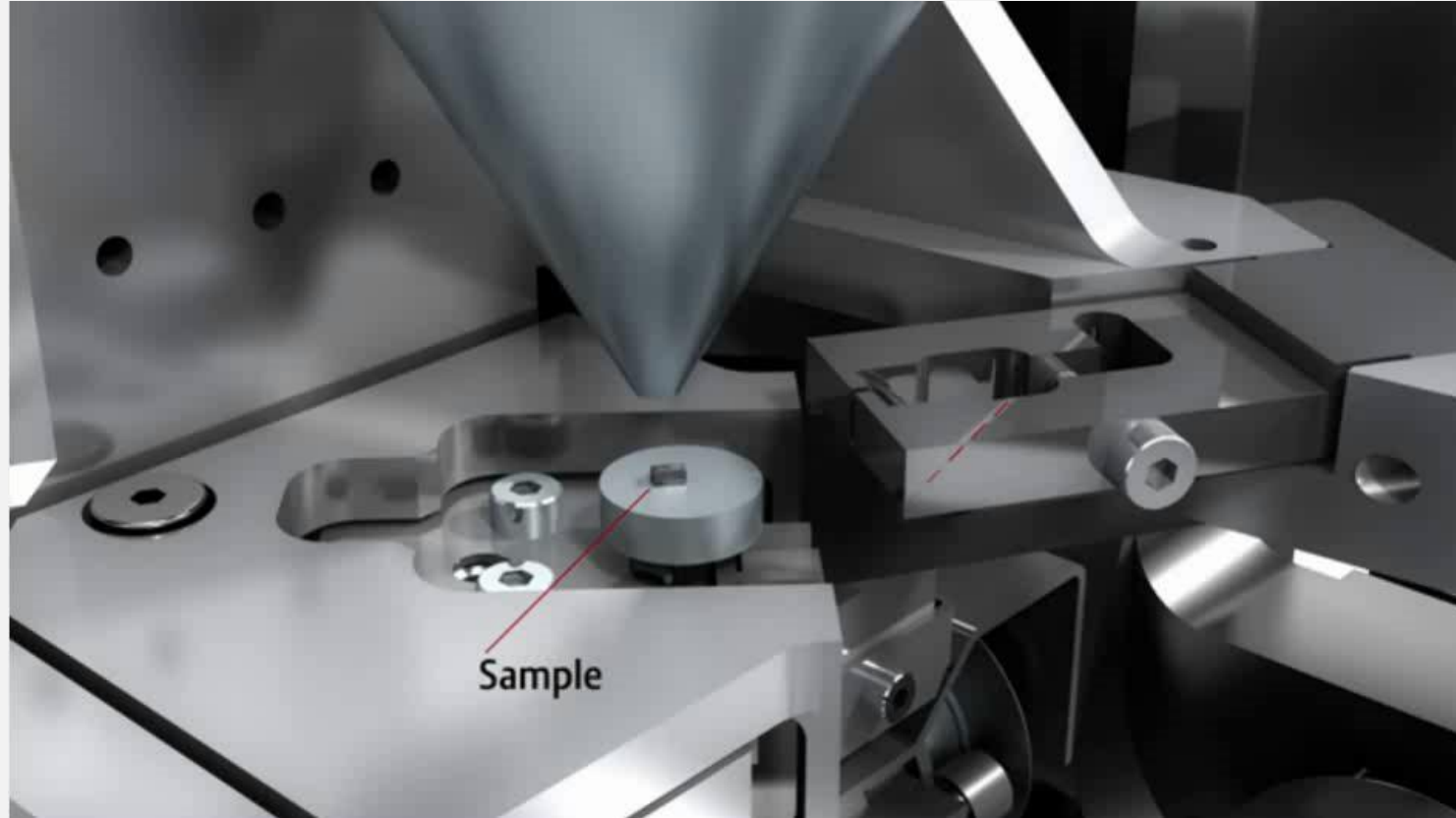
FIB/SEM



Serial block face



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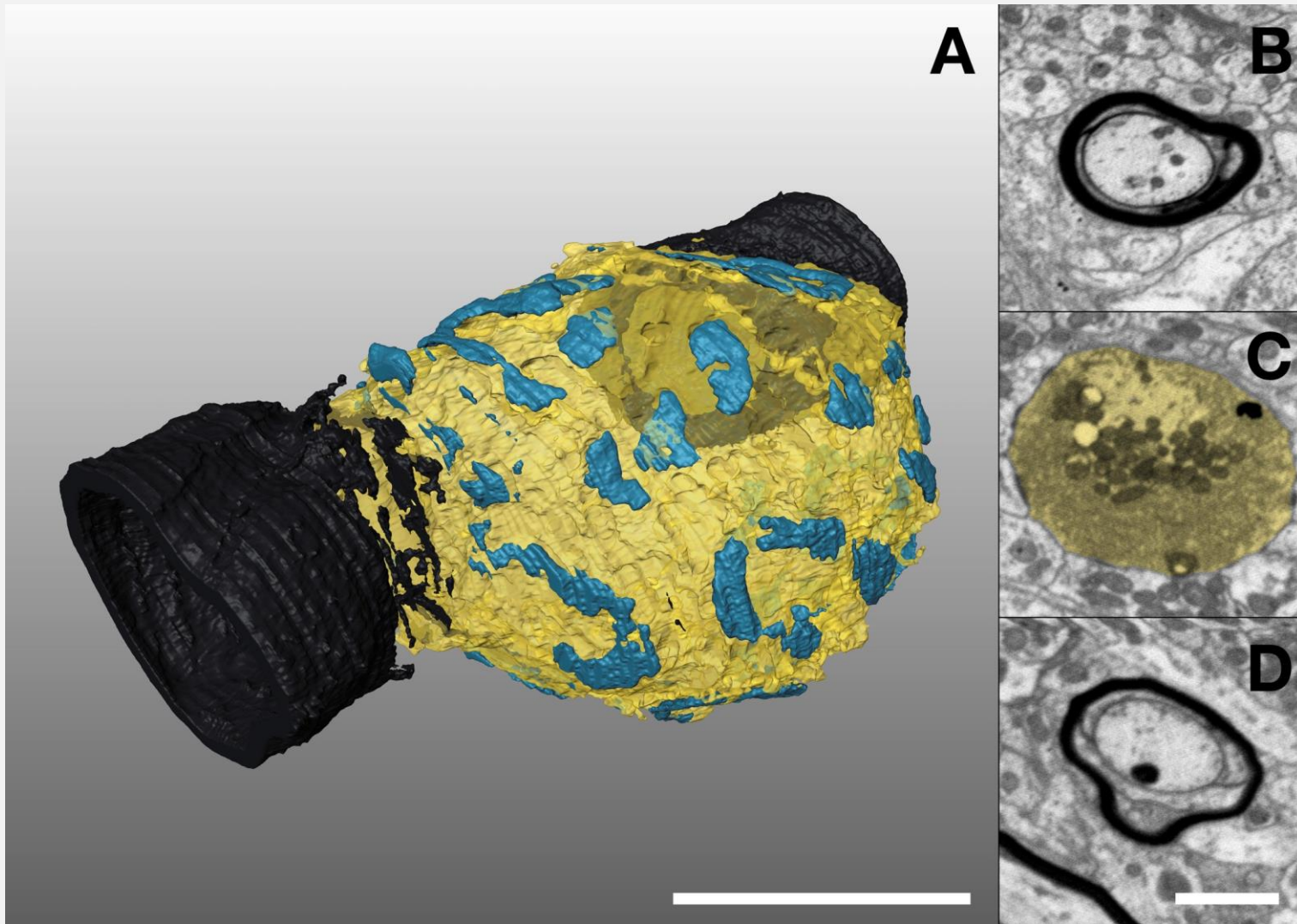


THE CORE FACILITY FOR INTEGRATED MICROSCOPY

FIB/SEM



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Electron Microscopy Department Staff

MD, PhD, Head of
Electron Microscopy



Professor
Klaus Qvortrup
qvortrup@sund.ku.dk

Electron Microscopy
Technical Director



Michael Johnson
michael.james@sund.ku.dk

PhD, Applications
Specialist



Tillmann Pape
tillman.pape@cpr.ku.dk

PhD, Applications
Specialist



Nicholas Heelund Sofos
nicholas.sofos@cpr.ku.dk

Electron Microscopy
Lab Manager



Zhila Nikrozi
zhilan@sund.ku.dk

Electron Microscopy
Lab technician



Cristiano Di Benedetto
zhilan@sund.ku.dk

THE CORE FACILITY FOR INTEGRATED MICROSCOPY

Light Microscopy Department Staff

PhD, Head of Light
Microscopy



Associate Professor
Clara Prats
cprats@sund.ku.dk

PhD, Applications
Specialist



Thomas H Braunstein
thobra@sund.ku.dk

PhD, Applications
Specialist



Pablo Hernandez Varas
pablo.varas@sund.ku.dk

PhD, Applications
Specialist



Jana Harizanova
j.harizanova@sund.ku.dk

Student Assistance

Master Student



Iva Svecová
iva.svecova@sund.ku.dk

Coordination

Secretary



Jacqueline van Hall
jacq@sund.ku.dk

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Image Analysis

PhD, Image Analyst



Richard Denis Maxime De Mets
richard.demets@sund.ku.dk

Computers

AUGUST KÖHLER

CPU Score : 45826
GPU Score : 4352
RAM : 128GB
Zen 3.4, Fiji, QuPath, CellProfiler



FRITZ ZERNIKE

CPU Score : 15750
GPU Score : 4352
RAM : 96GB
Zen 3.4, Fiji, QuPath, CellProfiler



ERNST ABBE

CPU Score : 45826
GPU Score : 4352
RAM : 128GB
Zen 3.5, Fiji, QuPath, Amira 2019.2



ERNST RUSKA

CPU Score : 45826
GPU Score : 10496
RAM : 128GB
Zen 3.5, Fiji, QuPath, CellProfiler, R



MANFRED VON ARDENNE

CPU Score : 45826
GPU Score : 4352
RAM : 128GB
Zen 3.6, Fiji, QuPath, Amira 2019.2, CellProfiler



SANTIAGO RAMÓN Y CAJAL

CPU Score : 81343
GPU Score : 10496
RAM : 256 GB
Zen 3.5, Fiji, Huygens, Arivis, Imaris, Matlab, ...



ADA LOVELACE (SOON)

CPU Score :
GPU Score :
RAM :
Zen 3.5, Fiji, QuPath, CellProfiler, Amira 2019.2



Free software



CELLPOSE



CELLPROFILER



FIJI



ICY



ILASTIK



NAPARI



QUPATH



ZEROCOSTDL4MIC



Commercial software

Avizo

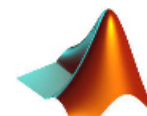
AMIRA-AZIVO



ARIVIS



IMARIS



MATLAB



HUYGENS



ZEN



The Core Facility for Integrated Microscopy



Training and support

Since 2010 – around 2000 users
Currently – 319 unique active users in 2021

Education



Principles Light and confocal Microscopy - PhD course

The course is suitable not only for beginners in microscopy, but also for those who already use microscopy in their work and now want to extend their knowledge of basic principles and more specialised techniques.

The wide range of microscopes available at CFIM allows for a strong practical element, with time for each student to gain hands-on experience coupled with lectures given by renowned scientists.

The course is run in collaboration with The Royal Microscopical Society.

Content

The course in Light Microscopy consists in two modules:

Basic Principles of microscopy

- Optics of the microscope
- Diffraction, Resolution and Contrast

Fluorescence and Confocal Microscopy

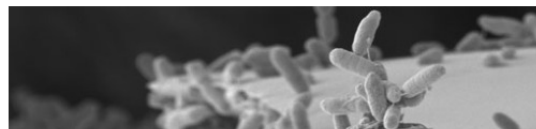
- Fluorescence microscopy (fluorophores, illumination...)
- Basic Confocal microscopy (Lasers, multi-dimensional Image Recording ...)
- Advanced techniques in fluorescence microscopy (FRET, FLAP, FCS)

Next course 9th - 13th & 21st-27th January 2017 - to be confirmed

Preliminary programme

For further information please contact Ragnhild Mostert, rmostert@sund.ku.dk
Registration: search for Light microscopy in the Course catalogue

Electron Microscopy - PhD Course



The course is suitable not only for beginners in microscopy, but also for those who already use microscopy in their work and now want to extend their knowledge of basic principles and more specialised techniques.

The wide range of microscopes available at CFIM allows for a strong practical element, with time for each student to gain hands-on experience coupled with lectures given by renowned scientists.

The course is run in collaboration with The Royal Microscopical Society.

Course content

The course provides an essential grounding in the basic principles of electron microscopy, covering topics such as:

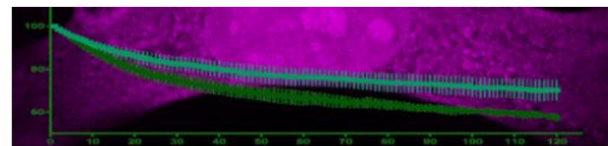
- Principles of Scanning and Transmitted Electron Microscopy
- Biological Specimen Preparation
- Advanced electron microscope techniques (immunogold labeling, electron tomography, and data analysis/visualization).

Next course 17th - 21st of October 2016

Last edition's Programme

For further information please contact Ragnhild Mostert, rmostert@sund.ku.dk
Registration: search for Electron microscopy in the Course catalogue

Image Processing - PhD Course



The course is suitable not only for beginners in image analysis with no experience, but also for those who want to extend their knowledge of basic principles and more specialised tools.

The course is mostly based on interactive lectures given by renowned scientists, where the students will perform themselves all processing and analysis tasks in parallel with the lecturers.

Course content

This course is an introduction to image processing (both light and electron microscopy images), analysis tools and basic Macro programming. The course will focus mainly on Fiji software but Cell profiler, Matlab, Amira, iMOD and ITEM will be demonstrated.

Main topics:

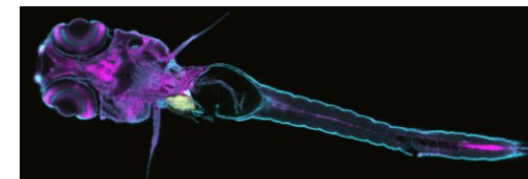
- what is a digital image? How and when to perform basic image processing tasks?
- Tools: segmentation, 2D and 3D measurements, tracking...
- Basic Macro programming for automated image analysis with Cell profiler and Fiji.

Next course: to be announced

Preliminary programme will be available soon.
Former edition's programme

For further information please contact Ragnhild Mostert, rmostert@sund.ku.dk
Registration: search for Image Processing in the Course catalogue

Light microscopy two-day crash course January 2022



January 2022 - 17th and 18th of January from 9.00 to 16.30.

This condensed course is aimed at everybody, who wants to understand the basic principles of optics and fluorescence in microscopy. Theoretical lectures are followed by practicals. The course will cover:

Basic Principles of microscopy

- Optics of the microscope
- Diffraction, Resolution and Contrast

Fluorescence and Confocal Microscopy

- Fluorescence microscopy (fluorophores, illumination...)
- Basic Confocal microscopy (lasers, multi-dimensional image acquisition, live imaging ...)

Overall satisfaction of previous students





Core Facility for Integrated Microscopy

Department of Biomedical Sciences

 All  

Core Facility for Integrated Microscopy

- Governance
- Booking
- Services
- Using the Core Facility
- Publications
- Public outreach
- Events
- Links
- Contact



CFIM - Core Facility for Integrated Microscopy - is a technology platform offering access to a wide range of state-of-the-art light and electron microscopes. Besides hosting equipment, CFIM provides expertise, training, and support.

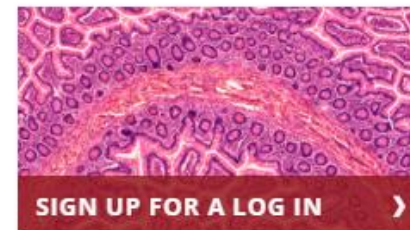


Visit www.cfim.ku.dk for more information

or contact:

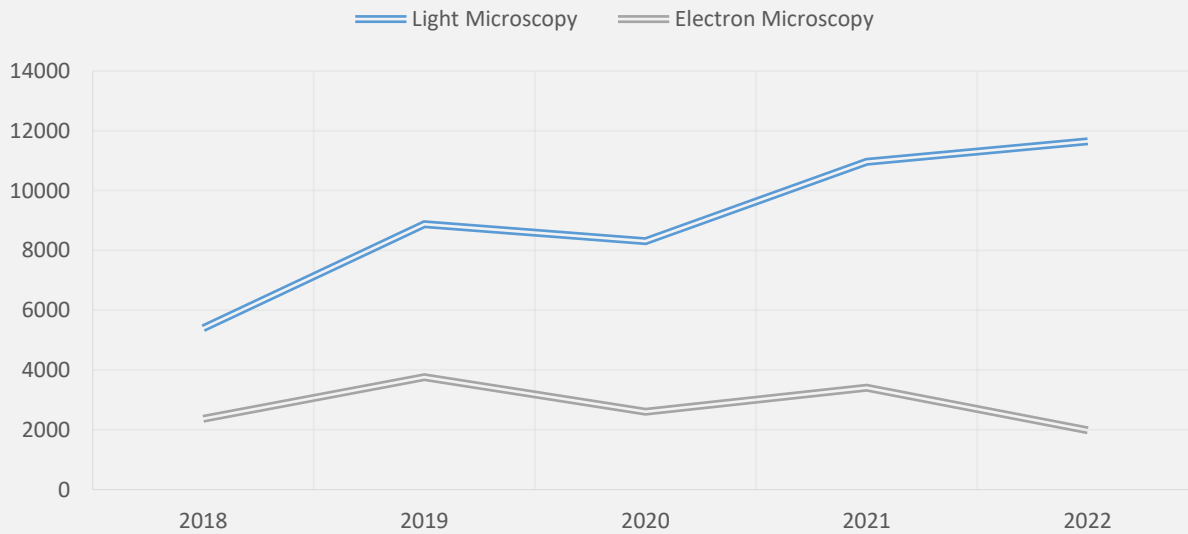
Klaus Qvortrup for EM

Clara Prats for LM

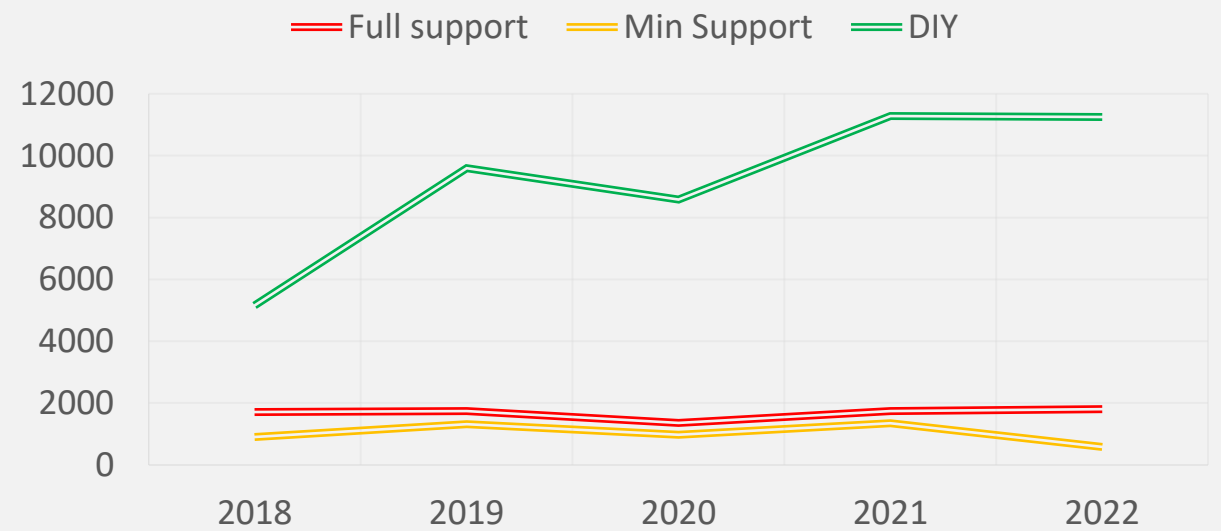


THE CORE FACILITY FOR INTEGRATED MICROSCOPY

USAGE (OPERATIONAL HOURS)



SUPPORT HOURS



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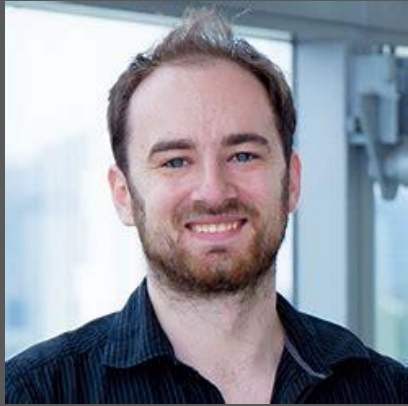
THE CORE FACILITY FOR INTEGRATED MICROSCOPY

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Image analysis

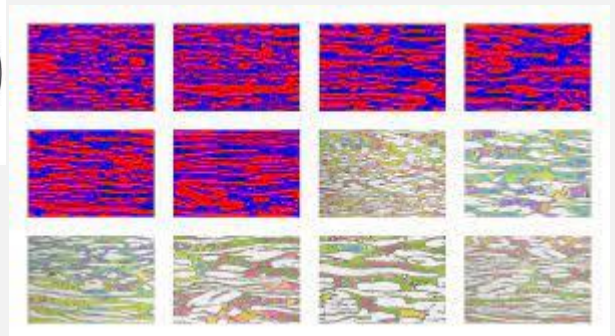
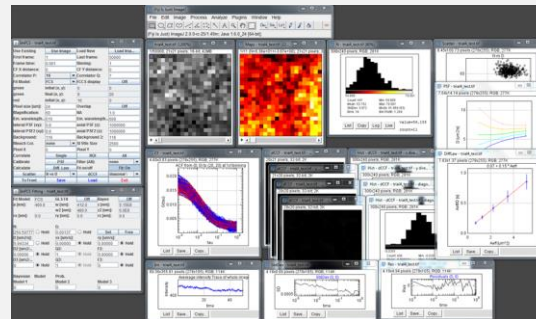
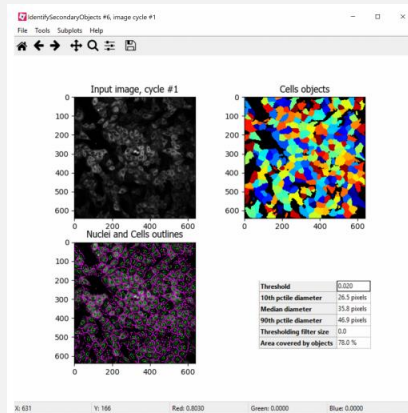
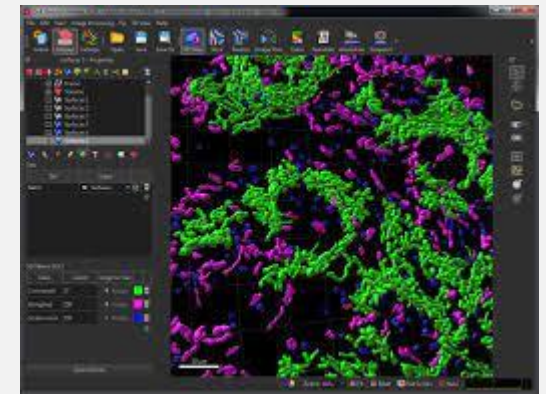
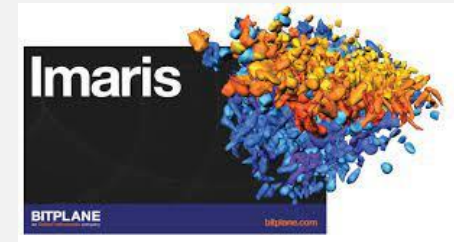


Richard De Mets

Bioimage Analyst at CFIM

- PhD in BioPhysics
- MSc Imaging Robotics and Life Engineering

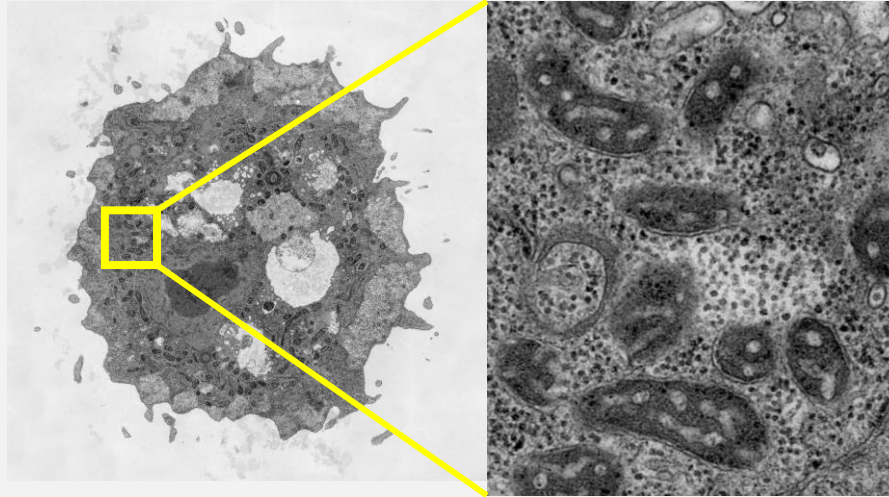
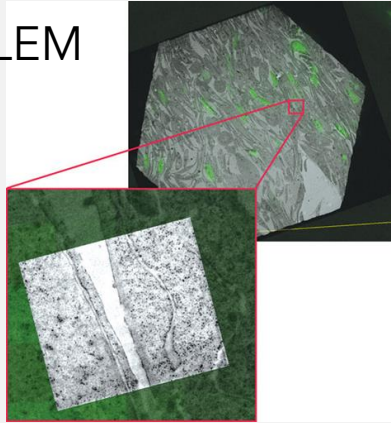
- Five powerful workstations for booking
- Image analysis software packages (Incl. Commercial and open source)



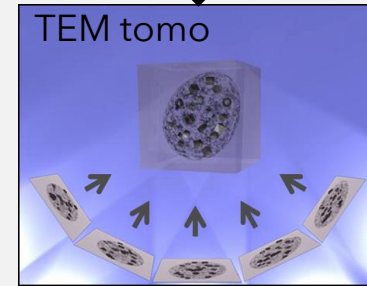
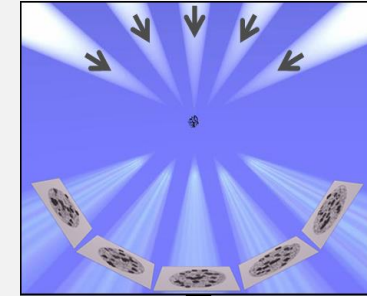
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Transmission Electron Microscopy (TEM)

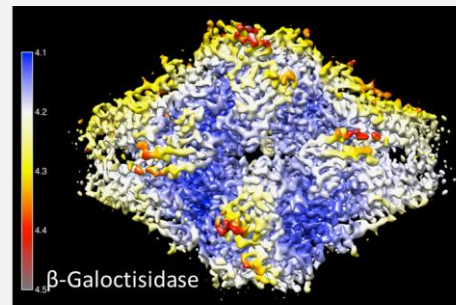
CLEM



TEM Tomography

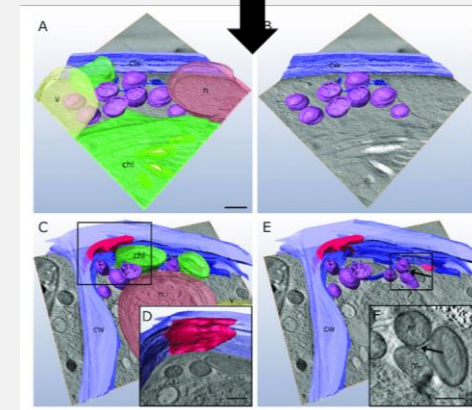
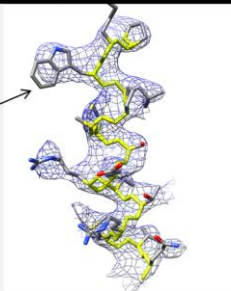


Cryo-TEM
NNF CPR











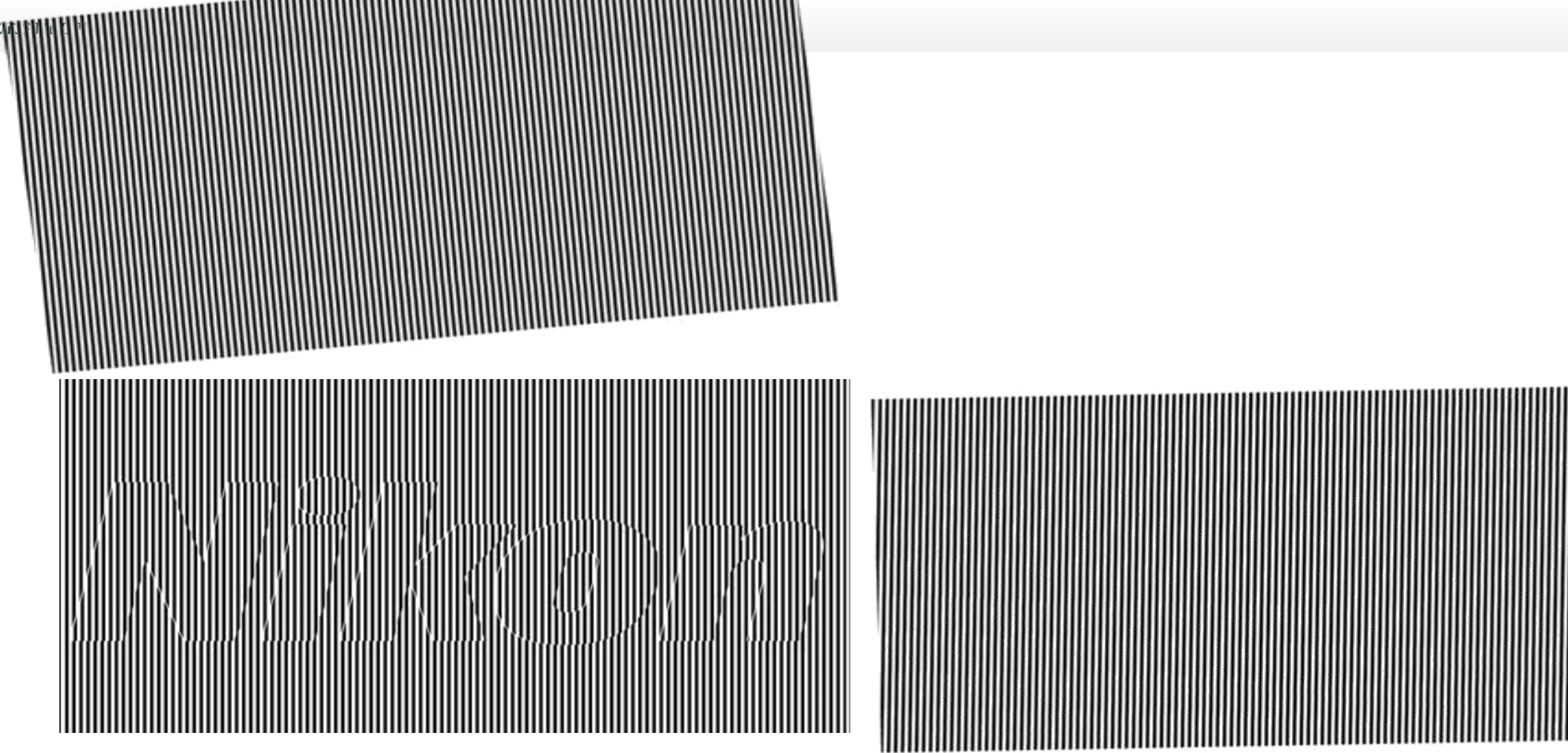
Resolution 4.2 Å
65,000 particles in total
26,000 particles for this model
by Pablo Mesa, CPR-SUND

lateral chains on the β -galactosidase



Improving resolution

		<u>@ 488 nm</u>		<u>Minimal object size</u> <u>(PSF)</u>
		xy	z	
1873 Abbe	Diffraction limit	250 nm	850 nm	 
1957 Minsky	Confocal	200 nm	500 nm	 
1994 Gustafsson	3D-SIM	100 nm	250 nm	 
2008 Zhuang	3D-STORM	20 nm	50 nm	 



- SR information is observable as a Moiré pattern created by the interaction of sub-resolution structures in the sample and the illumination pattern.
- The interference pattern is *coarser* than the sample and can be picked up by the optical system

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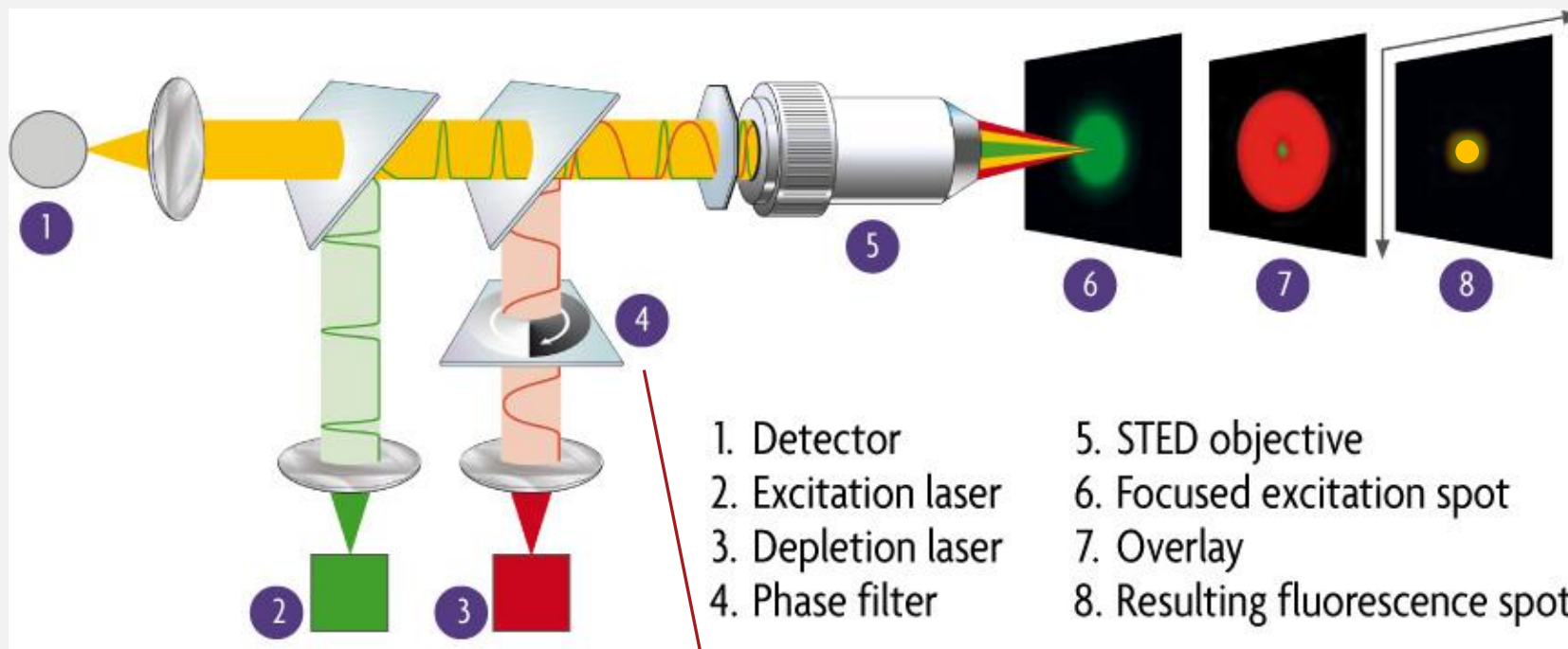


Image source: Leica

**STimulated
Emission
Depletion**

