

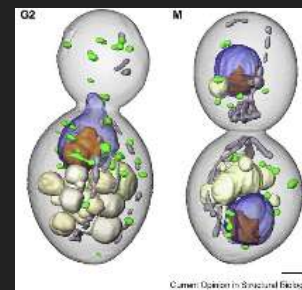
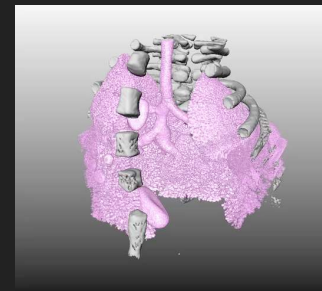
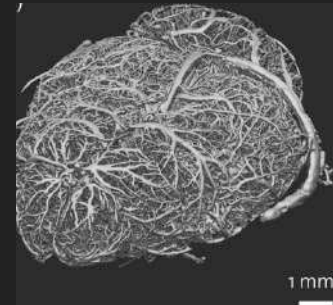
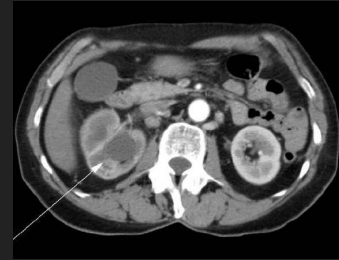


Imaging life with bright X-rays

Rajmund Mokso
Max IV Laboratory

Outline

- I. Introduction to MAX IV
 - I. Source and beamlines
- II. Currently available techniques
 - I. Nano probe and scattering
- III. Selected Science cases (future capabilities at MAX IV)
 - I. In vivo small animals, various fixed tissues



The MAX IV Accelerators

Ring (528 m circumf)

Experimental stations

Ring (96 m circumf)

Properties:

Wide band

High intensity/Brilliance

Polarization

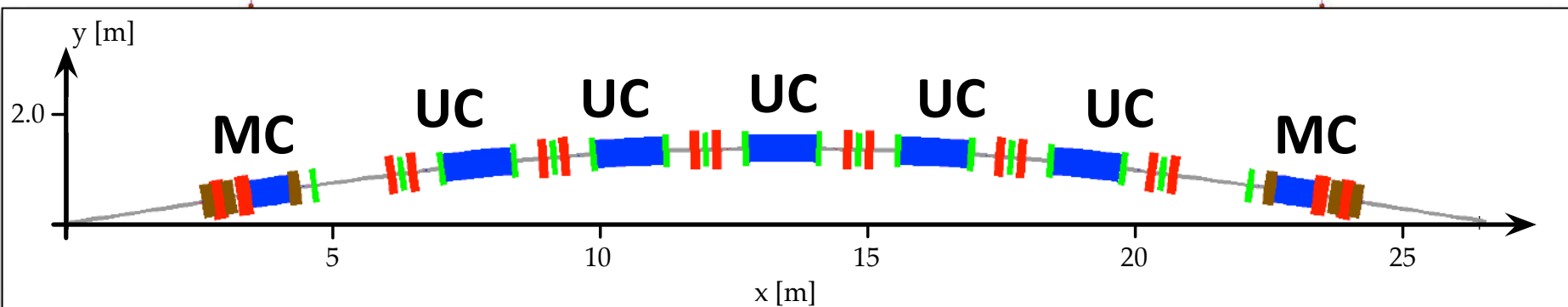
Time structure

Linear accelerator
(ca 250 m)

Electron source

MAX IV magnetic design

- 528 m circumference, 500 mA with top-up, 20 achromats
- 19 long straights (4.6 m) for users, 1 for injection
- 40 short straights (1.3 m) for RF & diagnostics
- 7-bend achromat: 5 unit cells (3°) & 2 matching cells (1.5° LGB)
- 328 pm rad bare lattice emittance (ϵ_y adjusted to 2-8 pm rad)



PRST-AB **12**, 120701 (2009)

IPAC'11, THPC059, p.3029

JSR **21**, 862-877 (2014)

The 7 bend achromat

Each achromat consists of 5
unit cells plus 2 matching cells

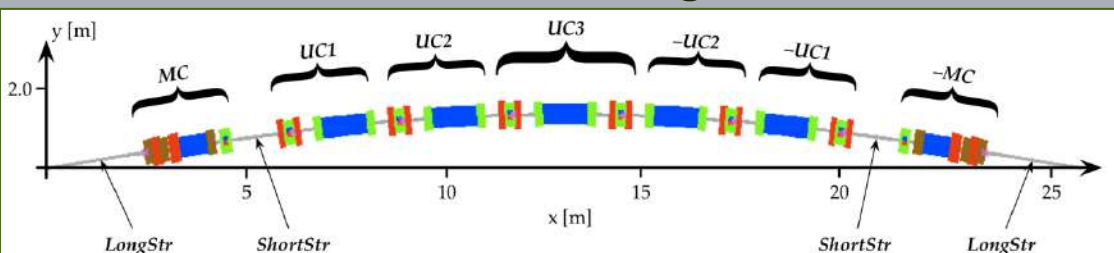


MAX IV magnets

- 528 m circumference, 500 mA with top-up, 20 achromats
- 19 long straights (4.6 m) for users, 1 for injection
- 40 short straights (1.3 m) for RF & diagnostics
- 7-bend achromat: 5 unit cells (3°) & 2 matching cells (1.5° LGB)
- 328 pm rad bare lattice emittance (ϵ_y adjusted to 2-8 pm rad)

Each cell is realized as one mechanical unit containing all magnet elements.

Each unit consists of a bottom and a top yoke half, machined out of one solid iron block, 2.3-3.4 m long.



MAX IV storage ring

- 528 m circumference, 500 mA with top-up, 20 achromats

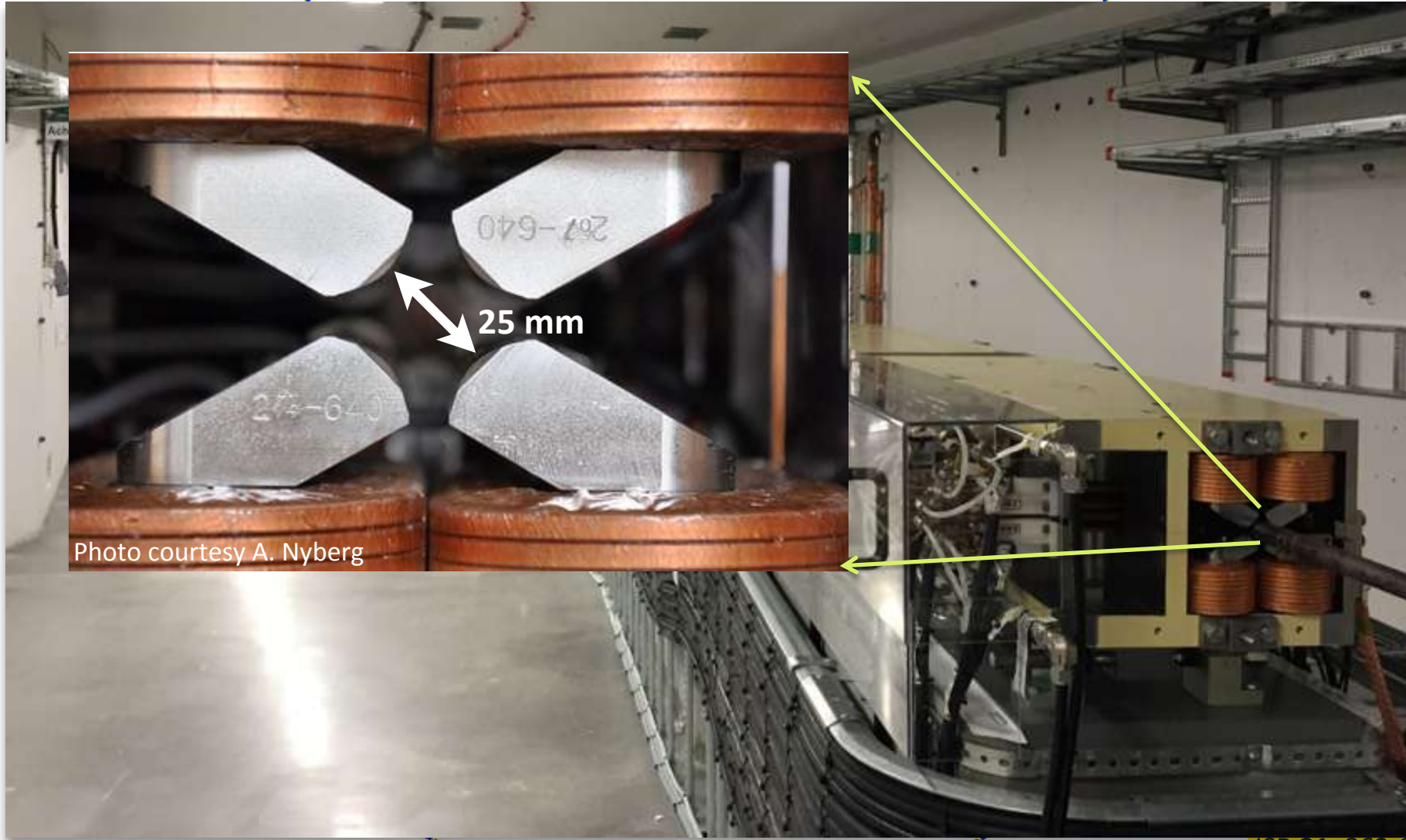
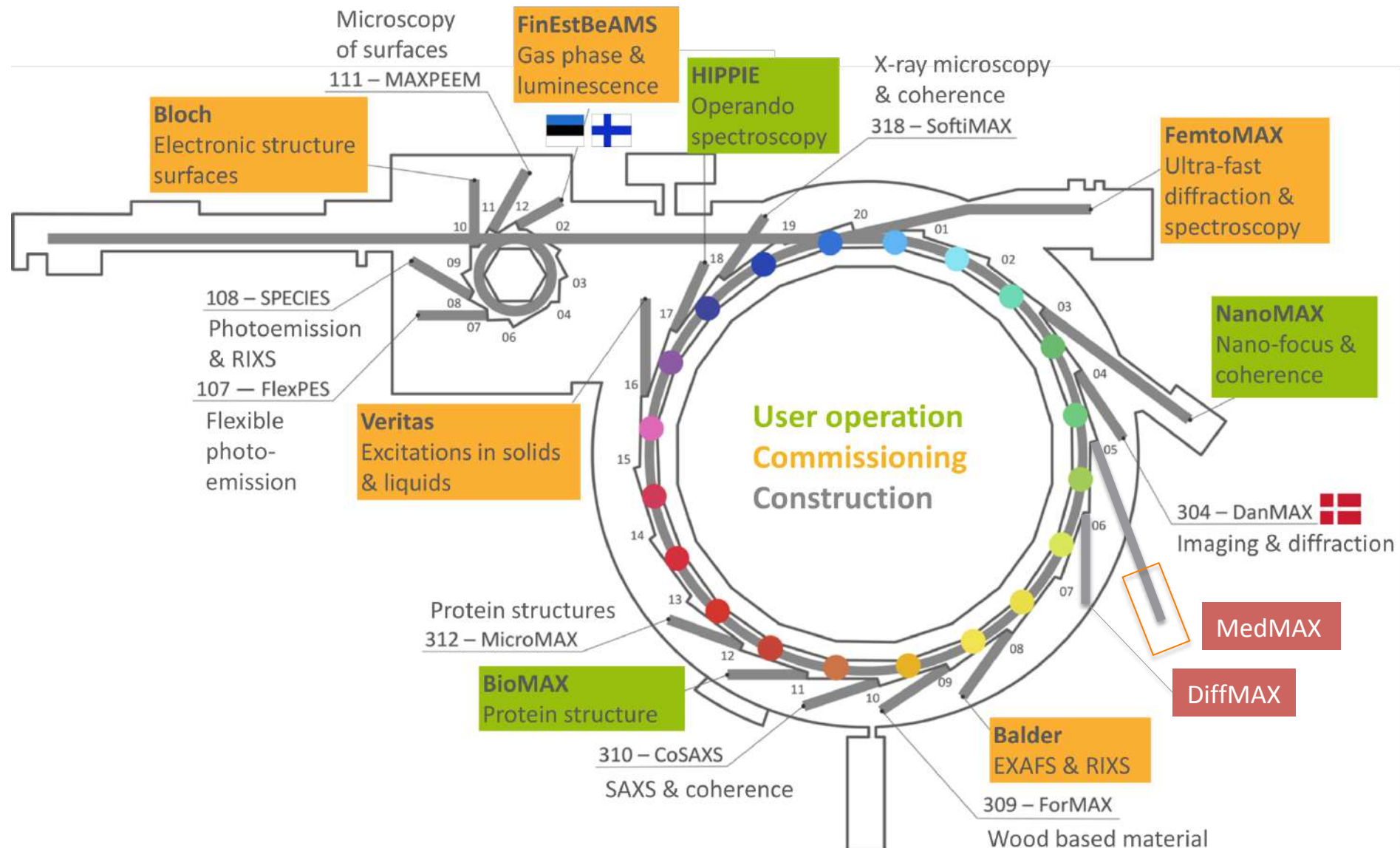


Photo courtesy A. Nyberg



Spectroscopy

SPECIES
HIPPIE VERITAS
FinEstBeaMS
FlexPES BALDER
ARPES

SoftiMAX

FemtoMAX

NanoMAX

BioMAX

DanMAX

MicroMAX

ForMAX

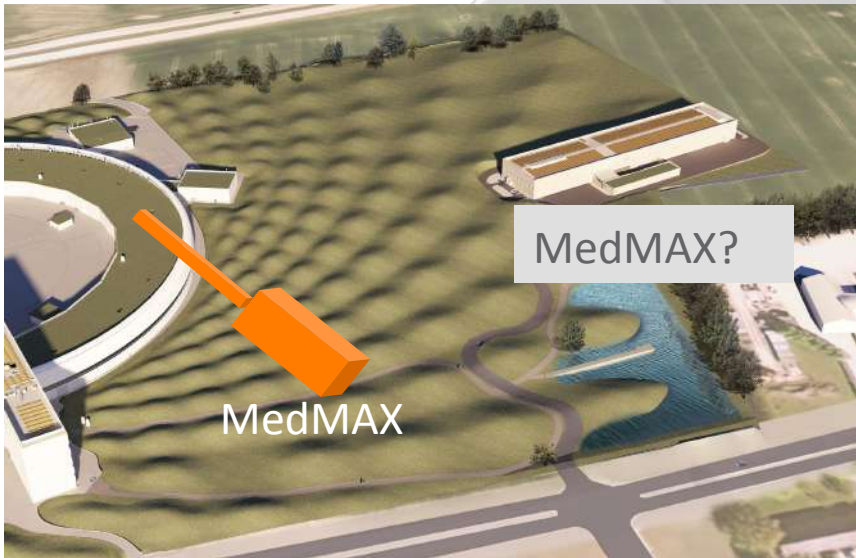
CoSAXS

MedMAX?

MedMAX

Imaging

Diffraction/ Scattering



MAX IV LIFE

Biology at different length and time scales

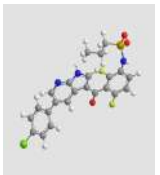
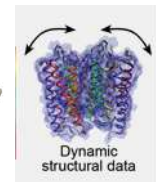
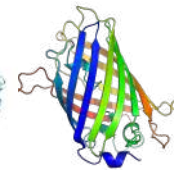
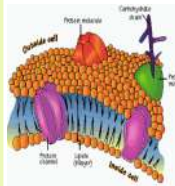
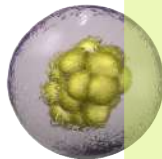
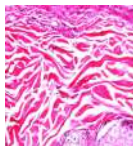
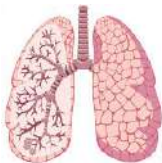
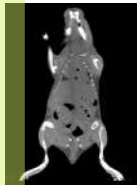


MedMAX III MedMAX II MedMAX I NanoMAX SoftiMAX DiffMAX coSAXS BioMAX MicroMAX FemtoMAX Balder

10^{-6} m

10^{-9} m

10^{-10} m



Animals

Organs

Tissues

Cells

Membranes
Surfaces

Micro-
structures

Molecular
complexes

Biomolecules

Atoms

“in/ex vivo”

“In solution”

Small animals/
disease models

Histology/
Histopathology

Cellbiology

Molecular Medicine/
Chemical Biology

Molecular Biology/
Biochemistry

The Max IV imaging beamlines

NanoMAX

The nanofocus beamline
(users in 2017)

SoftiMAX

Soft X-rays
(in construction -> 2018)

MedMAX

The biomedical imaging beamline
(preparing CDR)

DanMAX

Imaging & diffraction
(in construction -> 2018)

2016

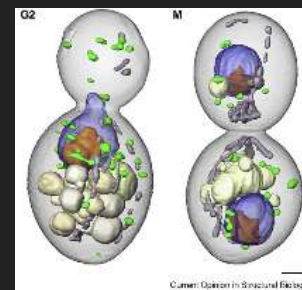
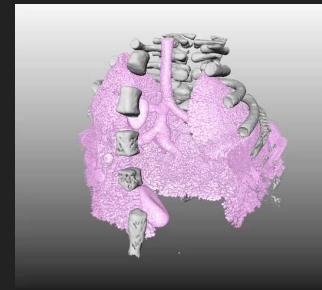
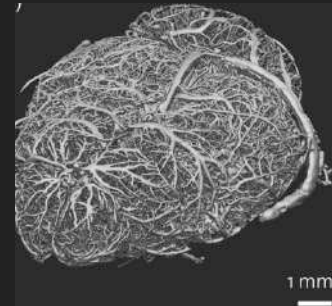
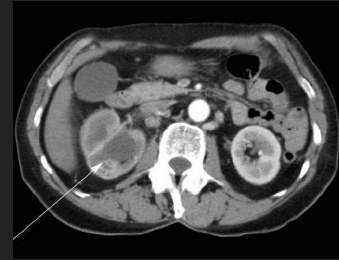
2017

2018

2019-20

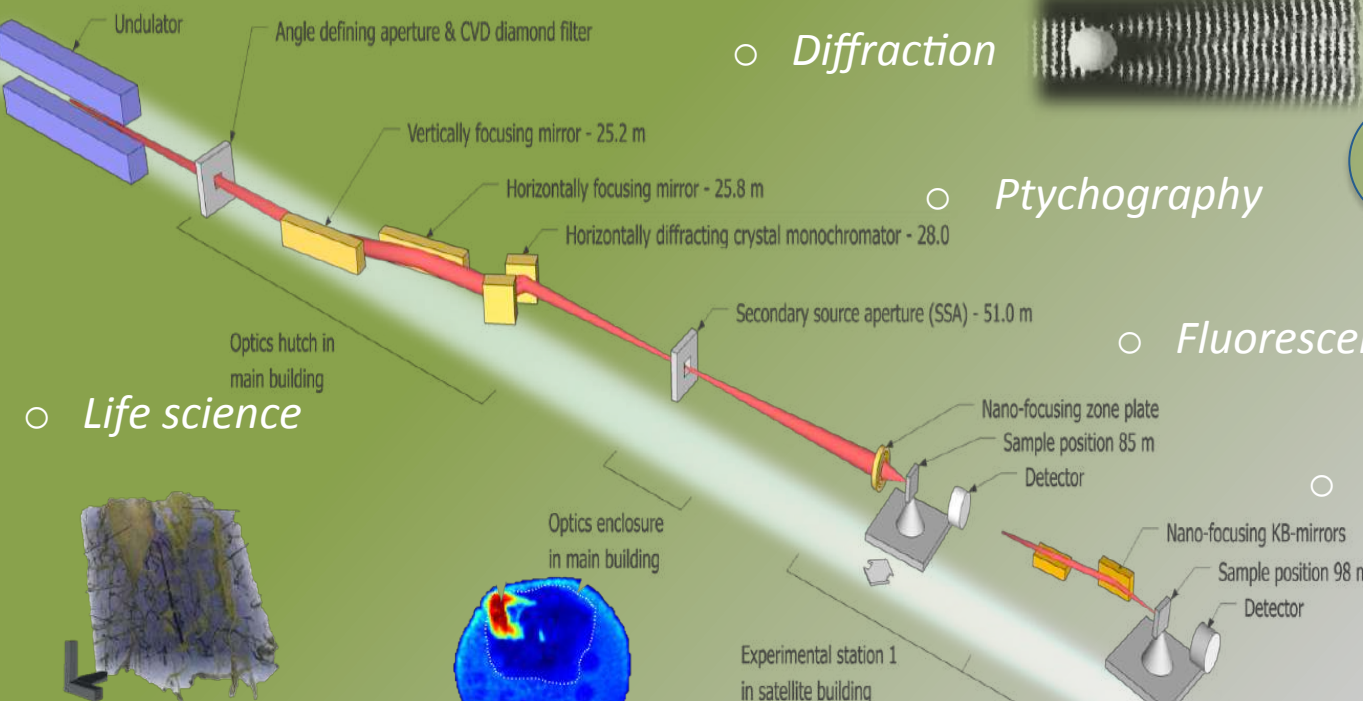
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NanoMAX: scanning X-ray nano-probe

2D & 3D
Structure & strain
Elemental distribution
Morphology



○ Diffraction

○ Ptychography

○ Fluorescence

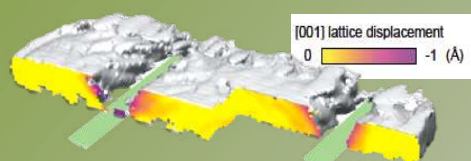
○ Absorption

○ Life science

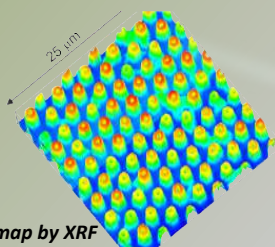
Bone inner structure by Ptychography
 M Dierolf et al. Nature 2010 467, 436-439

Malaria pigment in red blood cell by XRF
 F. Dubasr et al. Chem. Commun 2012 48, 910

○ Materials science

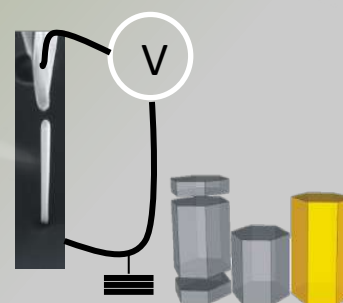


Strain in SiGe device by Bragg ptychography
 S. O. Hruszkewycz et al. arXiv:1506.01262v1
 [cond-mat.mtrl-sci]

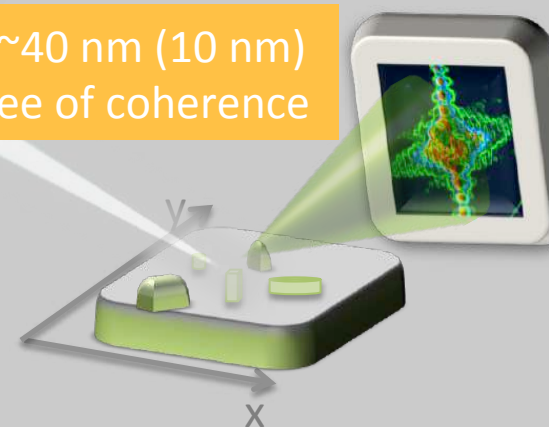


Ga density map by XRF
 G. Martinez-Criado et al
 Nano Lett. (2012) 12, 5829

Focusing beam to ~40 nm (10 nm)
 Clean and with high degree of coherence

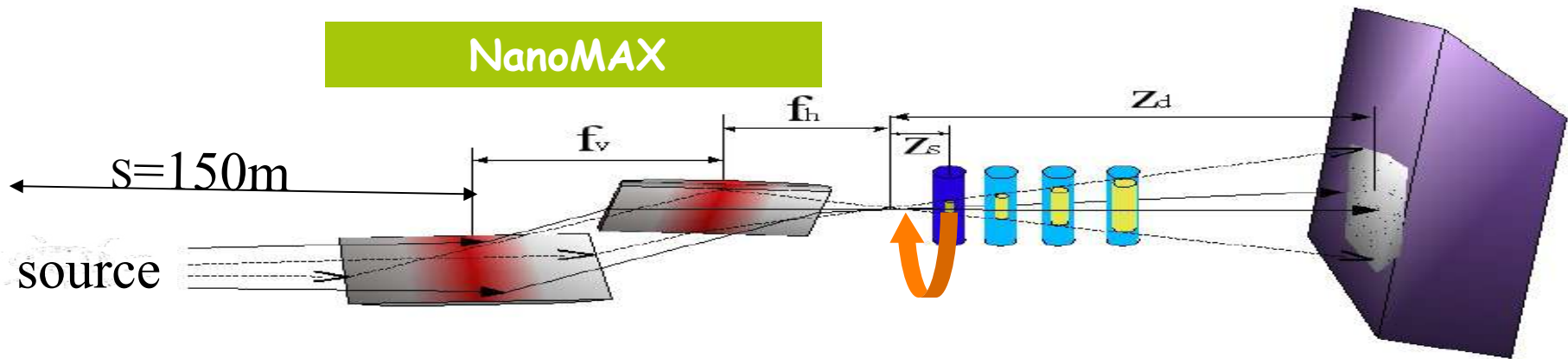


Structure and transport in GaAs by In-Situ XRD
 G. Bussone et al. Nano Lett. 2015, 15 981



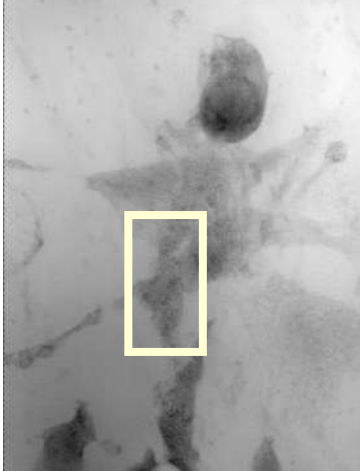
Nanoimaging, Fluorescence spectroscopy

NanoMAX



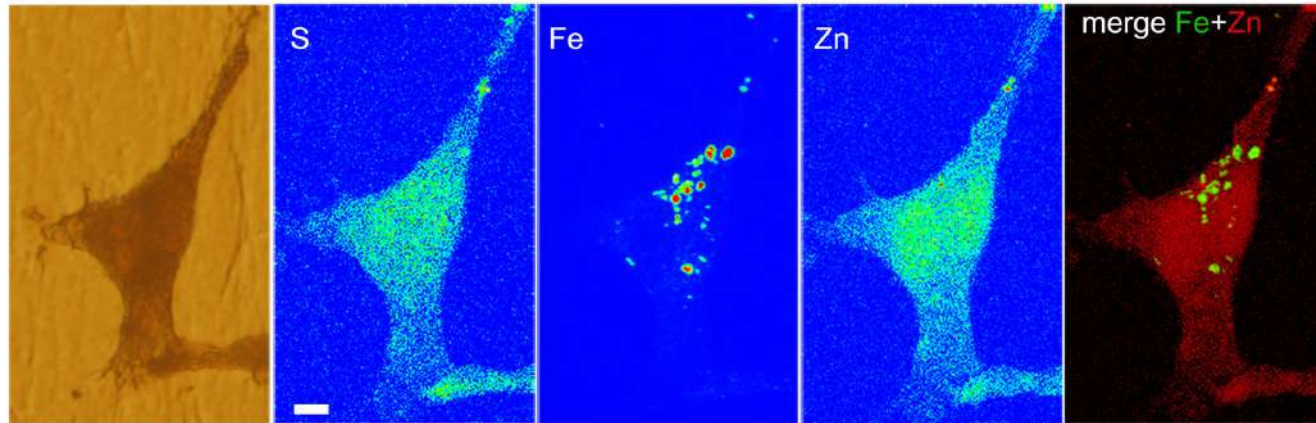
$10\ \mu\text{m}$

A neuron cell phase image



Mokso, Cloetens et al. , APL 2007

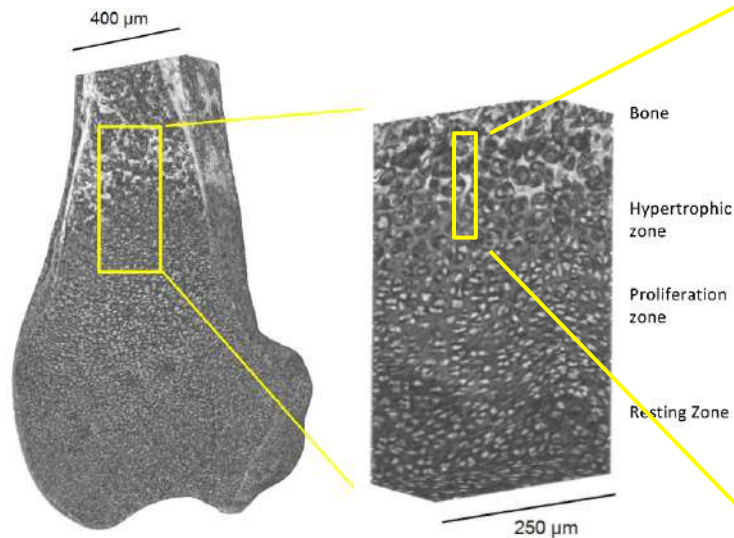
Chemical element distribution



Ortega. , Mol. Neurobiol. 2016

NanoMAX: first users

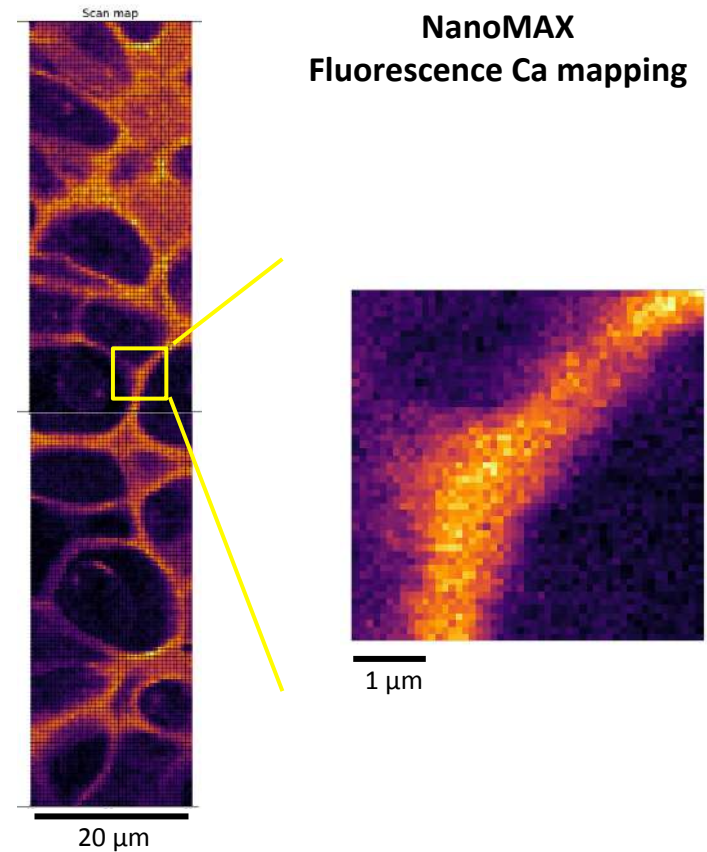
Embryonic mouse growth plate



Phase contrast x-ray tomography (TOMCAT, PSI)
Distal humerus, mouse TS24

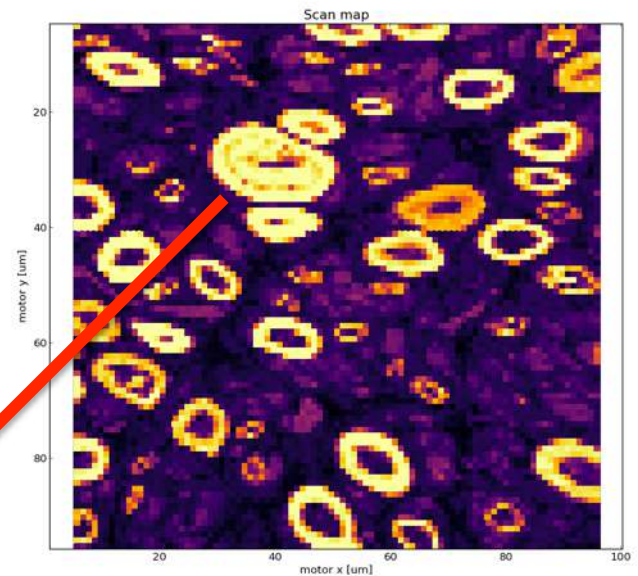
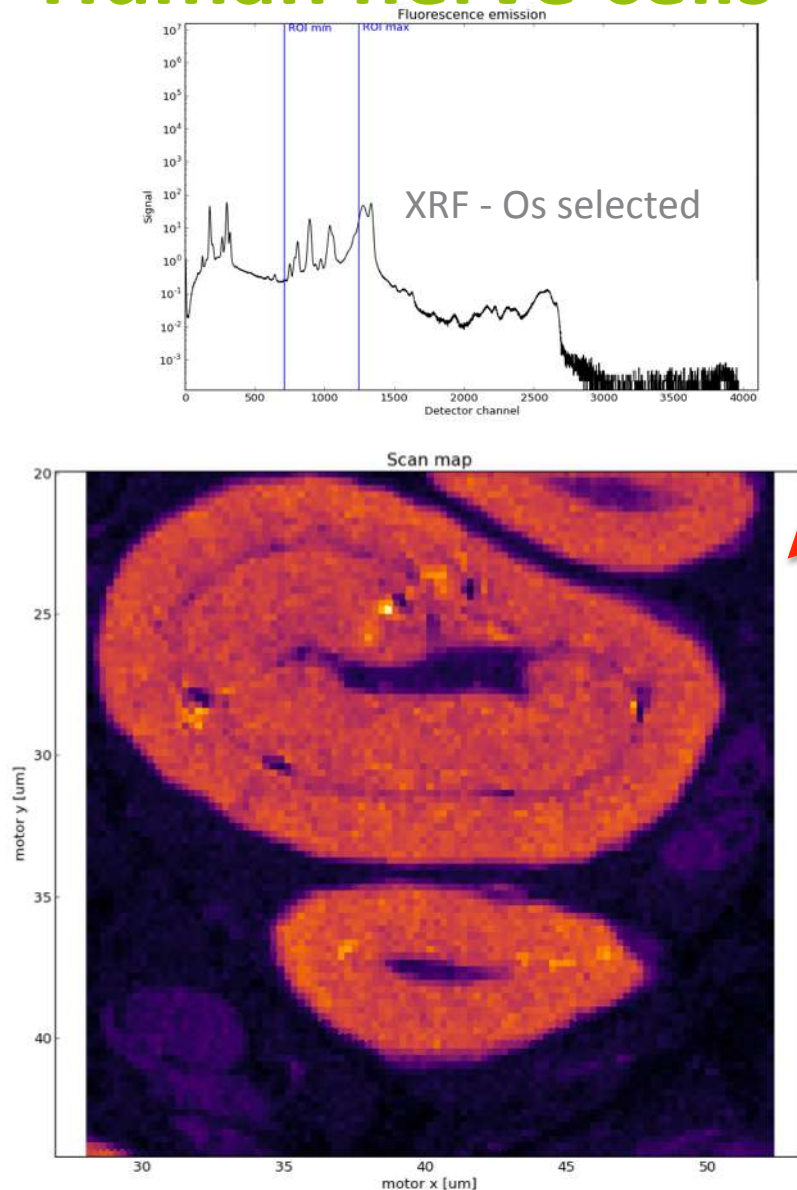
Hanna Isaksson, Lund University
Niamh Nowlan, Imperial Collage, London

NanoMAX Fluorescence Ca mapping



Growth plate, hypertrophic zone, Mouse TS 23

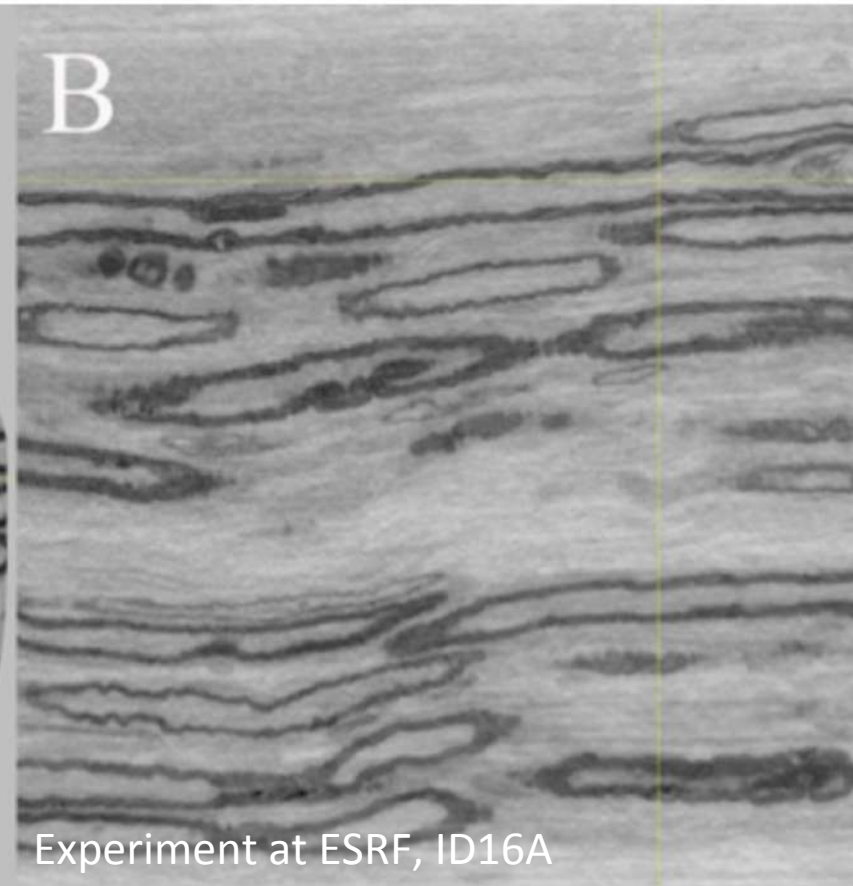
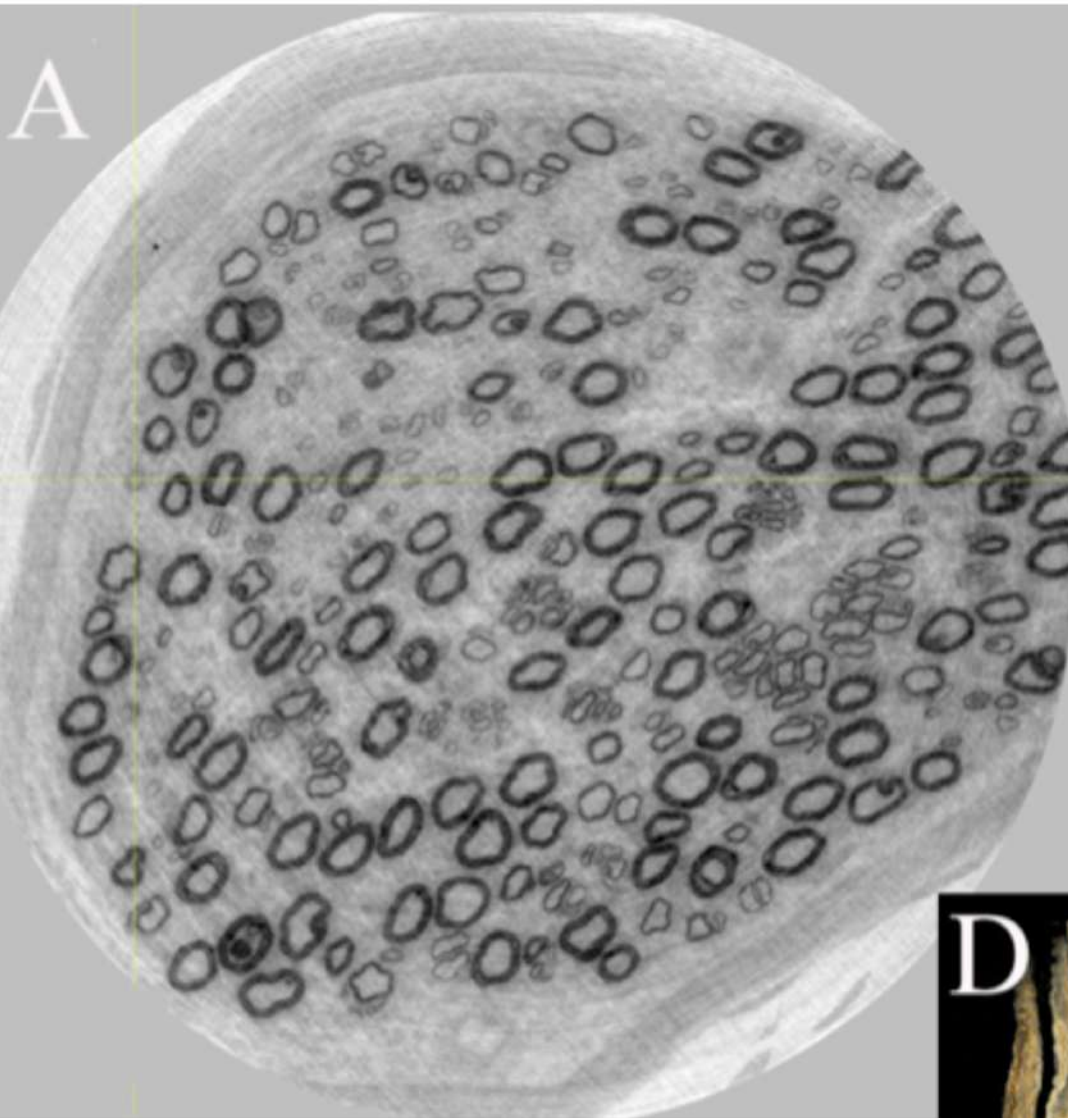
Human nerve cells – diabetes type 2



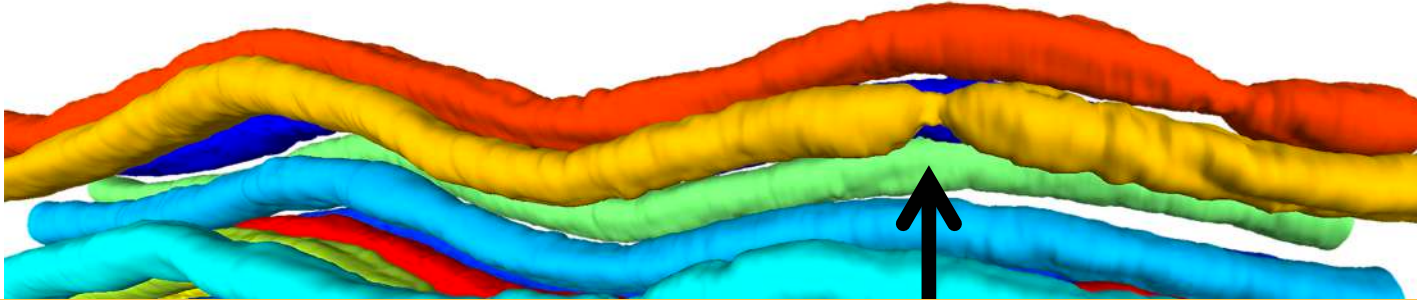
XRF imaging of human nerve cells of diabetes type 2 person. Samples stained with Osmium.
Left image 25x20 μm^2 .

Courtesy of Lars Dahlin & Martin Bech,
BMC Lund University.

Human peripheral nerves



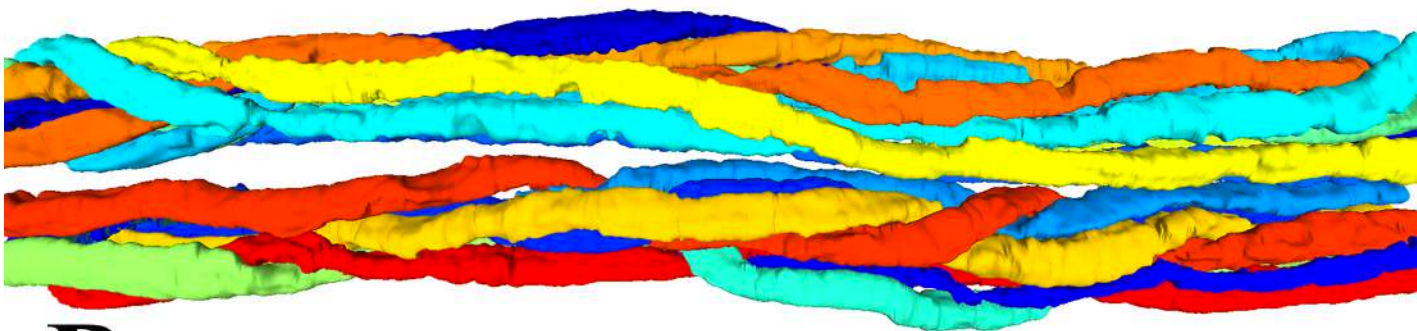
Human peripheral nerves (diabetes)



MedMAX : sub-micrometer resolution 3D histology

A = normal nerve fibers

Node of Ranvier



B

= regenerative clusters

Fluorescence Tomography 3D

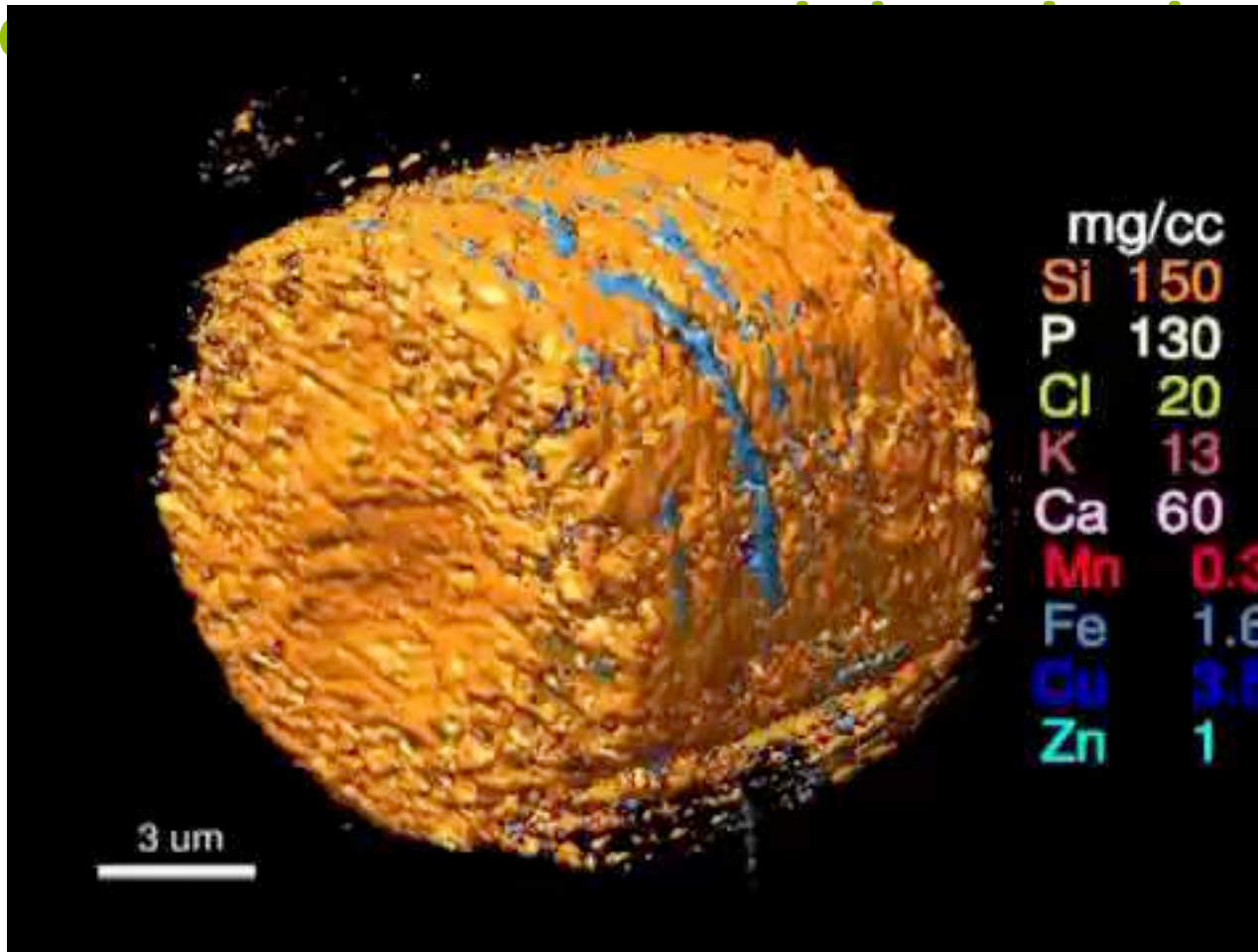


Figure 24. X-ray fluorescence tomography of an air-dried single-celled freshwater diatom *Cyclotella meneghiniana*. The silicacious frustule has been removed to show inner detail. Adapted with permission from ref 76. Copyright 2010 National Academy of Sciences.

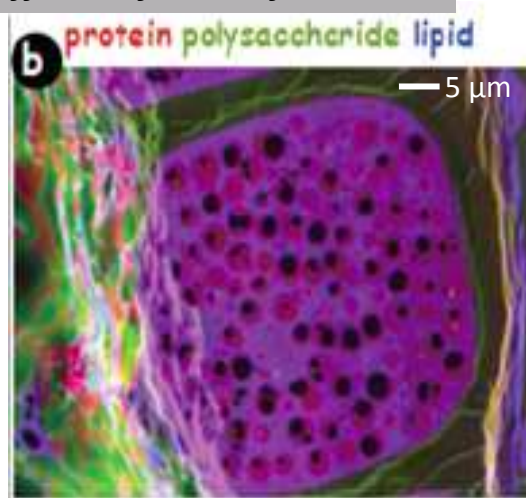
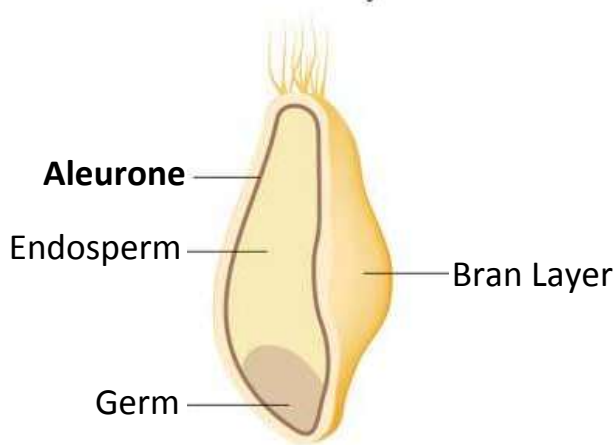
SoftiMAX: Soft X-ray scanning microscopy



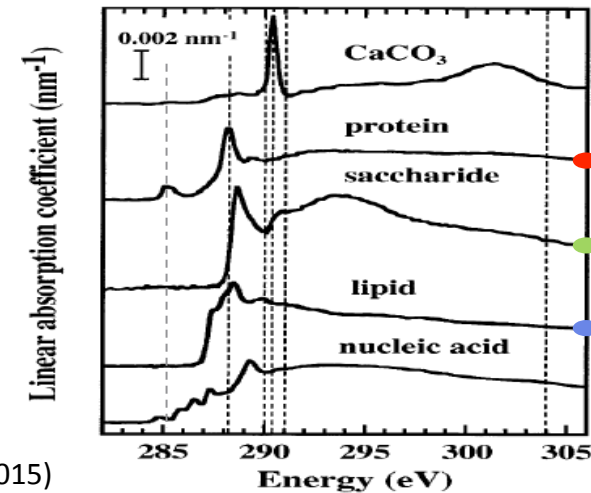
Currently finalizing the design of the scanning transmission microscopy endstation

A grain's anatomy by chemical contrast: different forms of carbon

Grain anatomy



C. Karunakaran, *et al.*, PLoS One 10, e0122959 (2015)

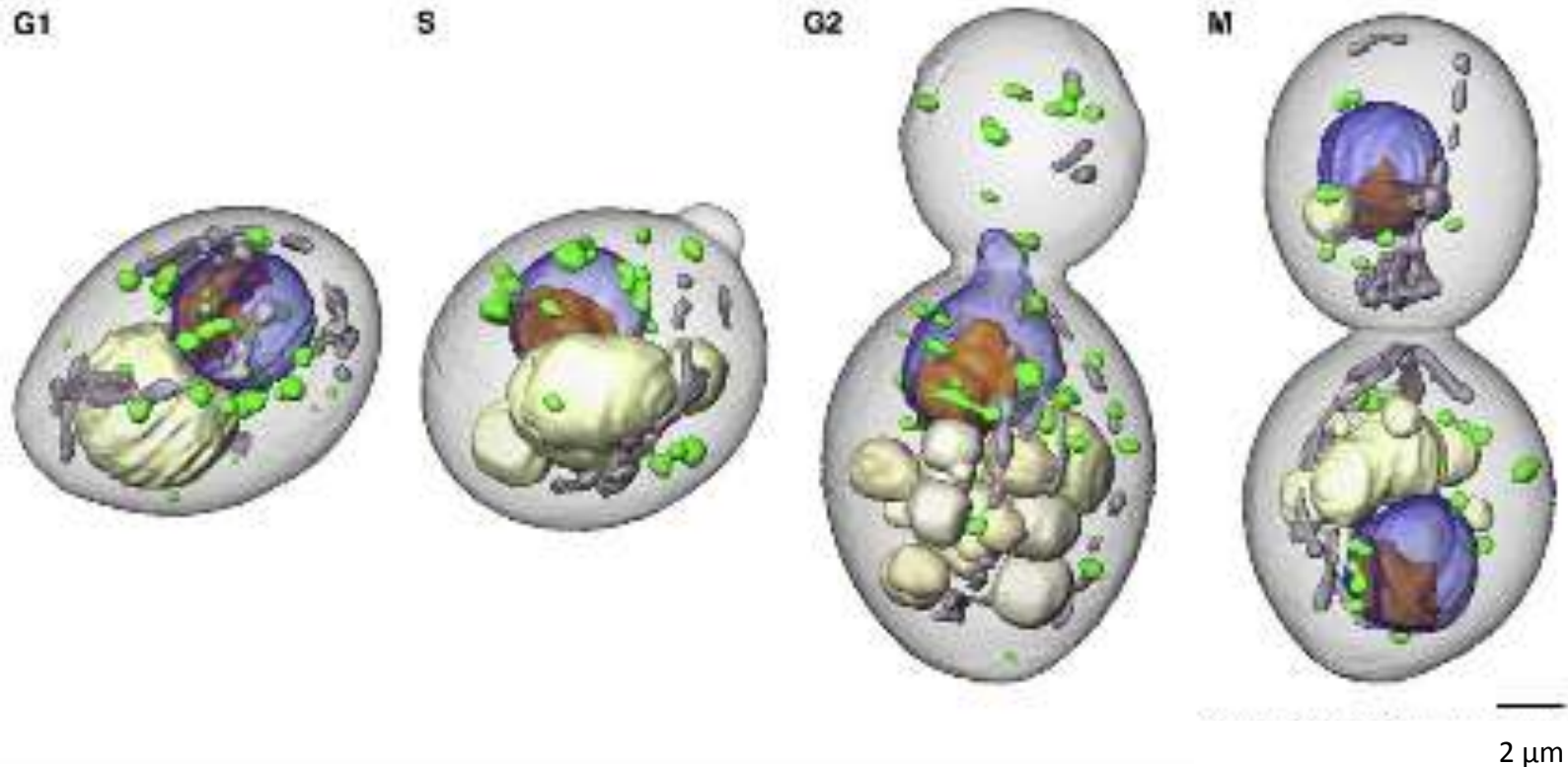


Imaging cells with hard X-rays still a big challenge.

While it is almost a routine with soft X-rays

Soft X-ray microscopy

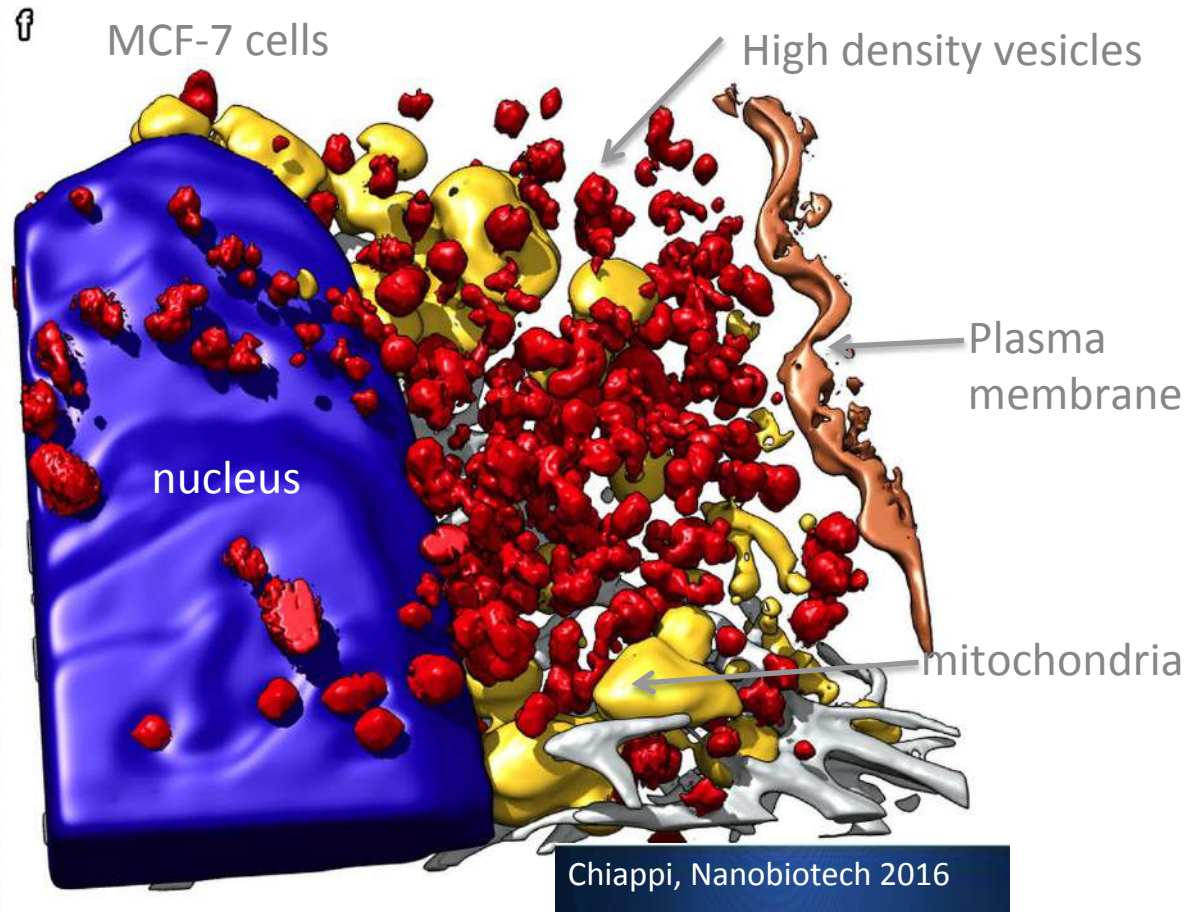
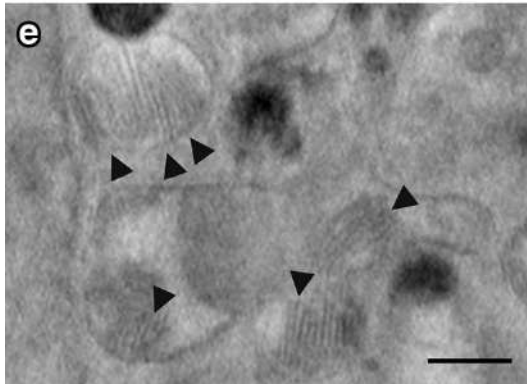
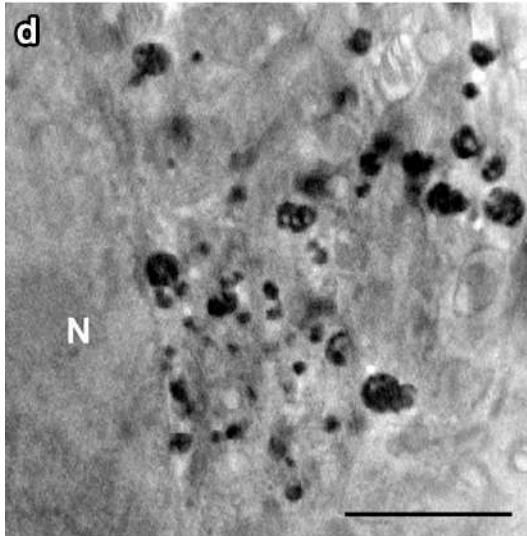
spatial resolution ~ 30 nm



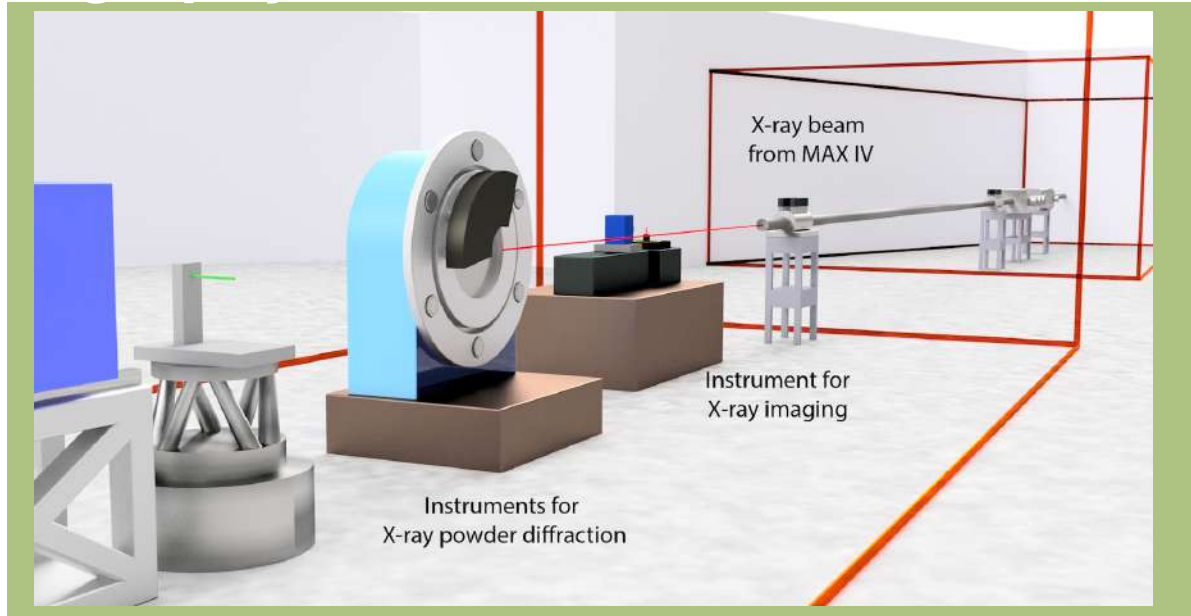
Larabell & Nugent, Current Opinion in Structural Biology (2010)

Soft X-ray tomography of rapidly frozen *Saccharomyces cerevisiae* cells imaged at each phase of cell cycle – G1, S, M, and G2. Organelles are color-coded as follows: blue, nucleus; orange, nucleolus; gray, mitochondria; ivory, vacuoles; green, lipid bodies.

Cryo soft X-ray microscopy of cells



DanMAX: tomography and diffraction for materials

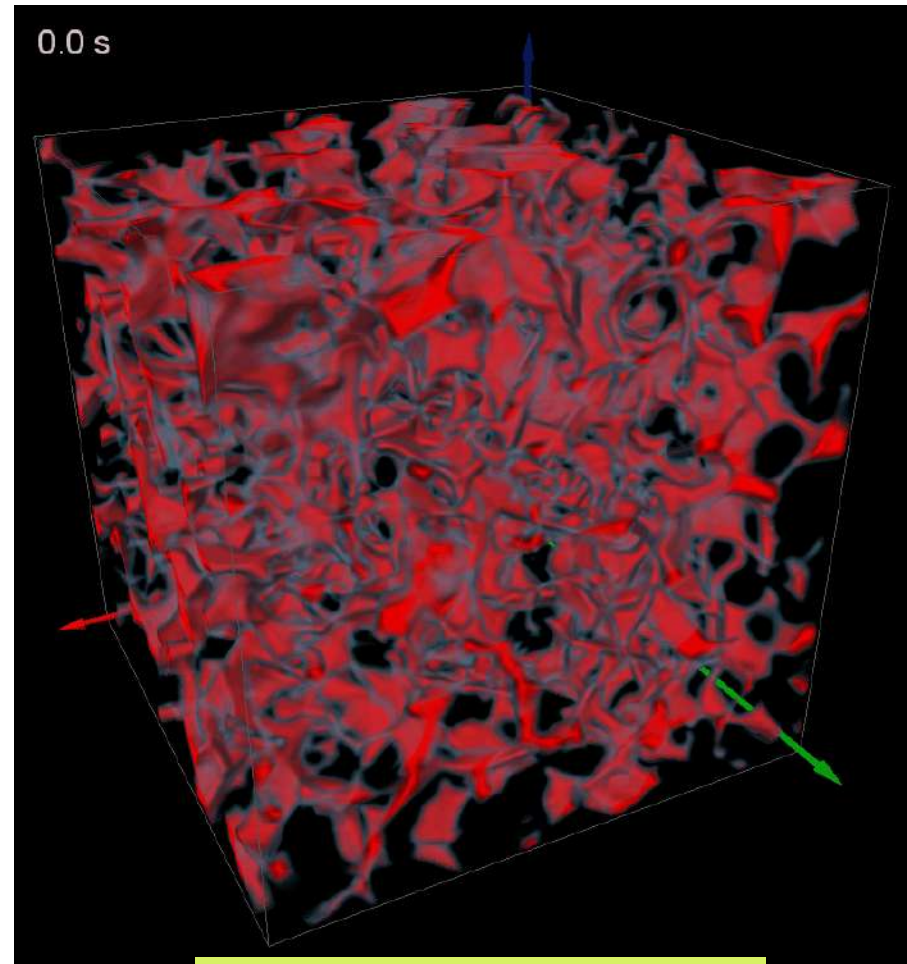


- study real materials at real conditions at real timescales.
- Combining powder X-ray diffraction (PXRD) and full field imaging

DanMAX: 1/2 tomographic microscopy



Crystal melting

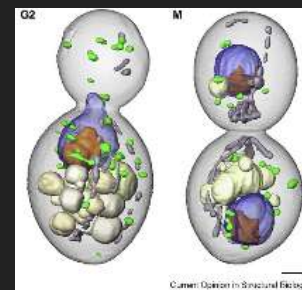
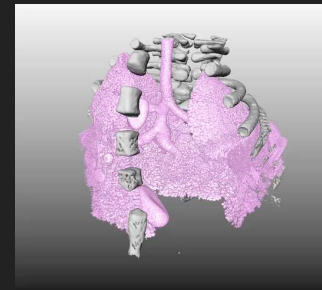
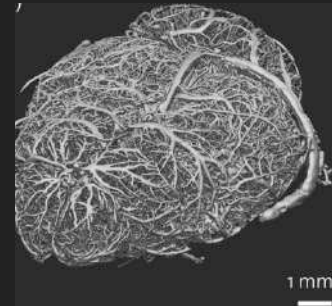
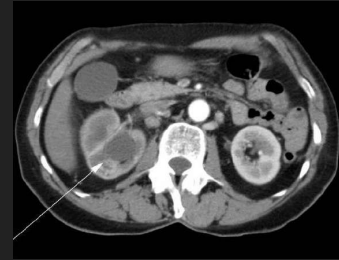


Exploiting oil from the rock

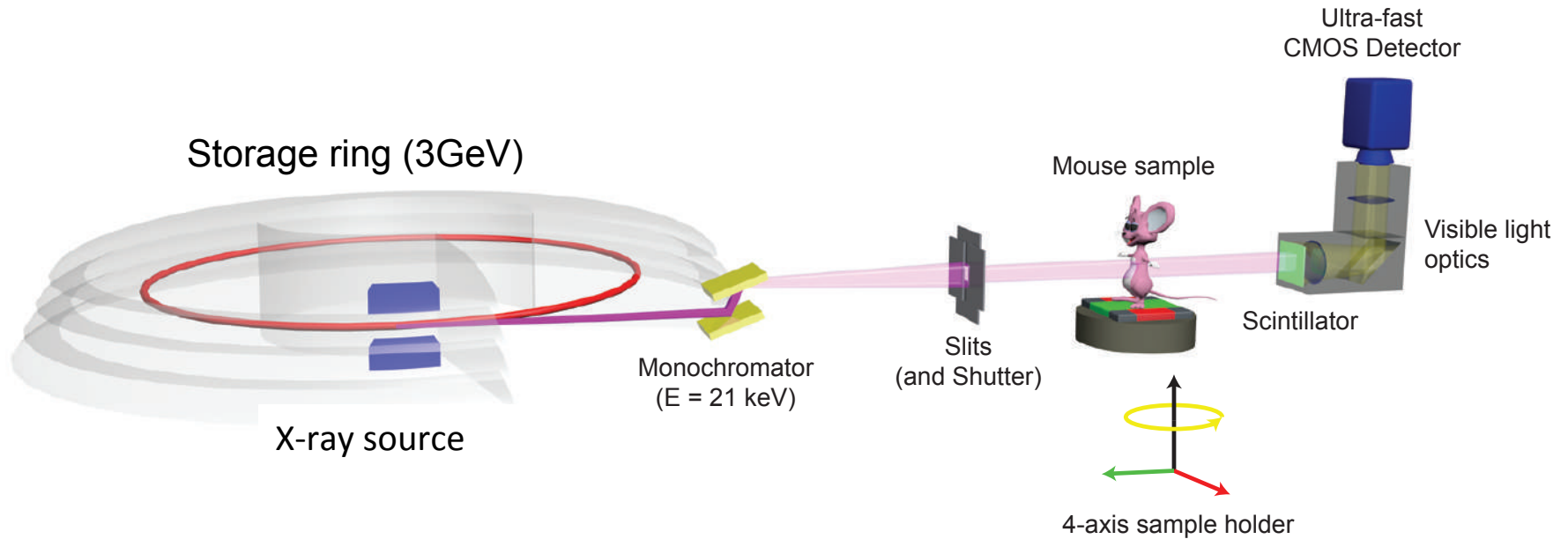
Seeing the dynamics of matter in 3D at the micrometer scale

Outline

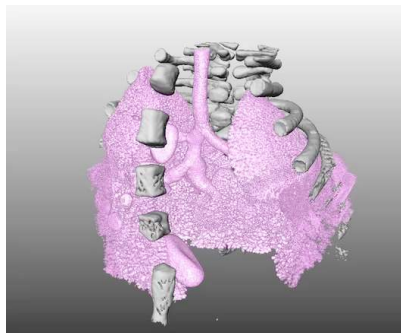
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Tomographic microscopy at synchrotron



Radiographic projection
clinics



Tomographic reconstruction
Synchrotron

Pixel size: 11 $\rightarrow$ 0.3 μm

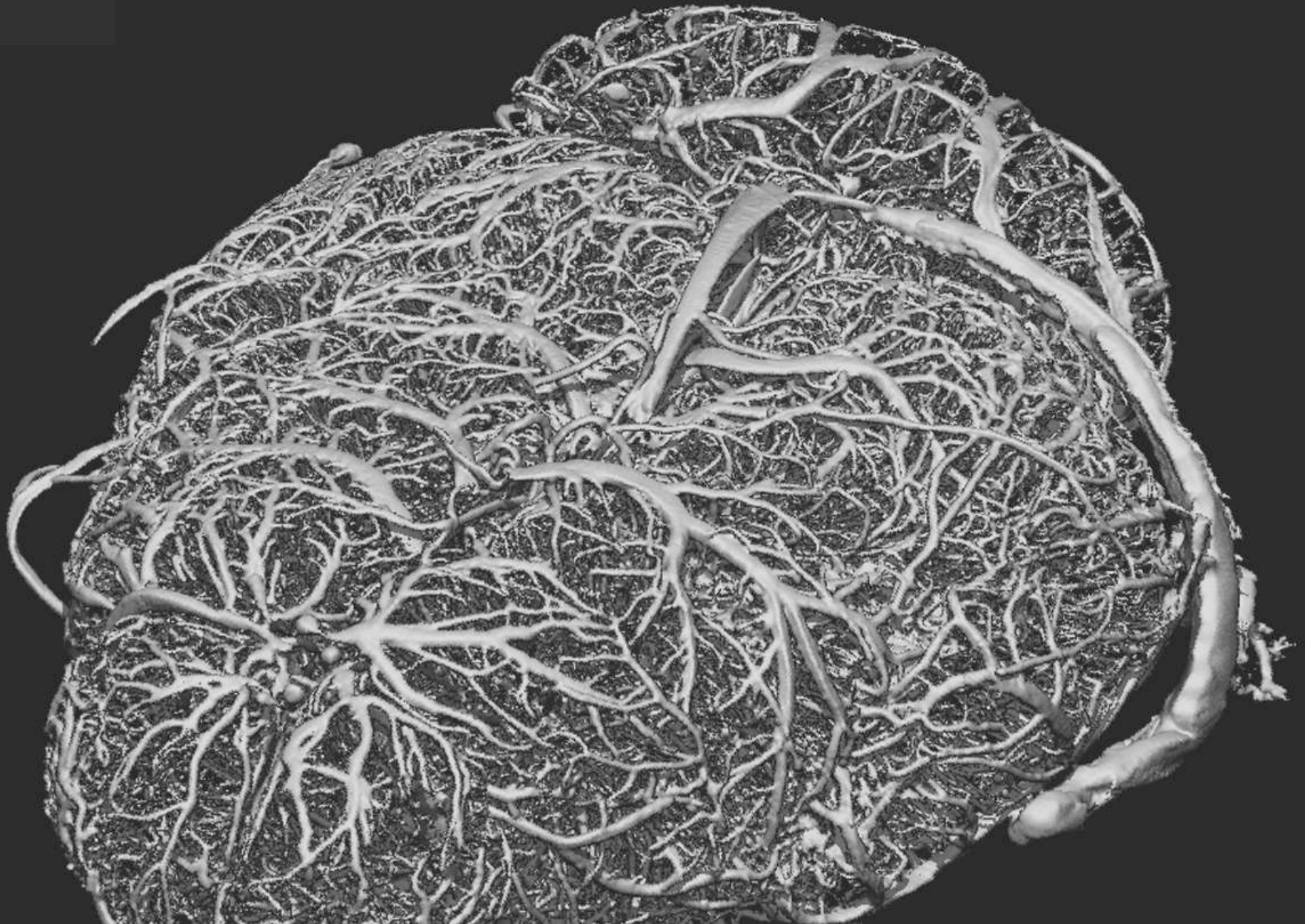
FOV: 22 x 22 $\rightarrow$ 1 x 1 mm^2

Projections: 300–2000

Exposure times: 0.1–300 ms

Total scan time: 0.05 to 500 s

The MedMAX project



Medical imaging: CT

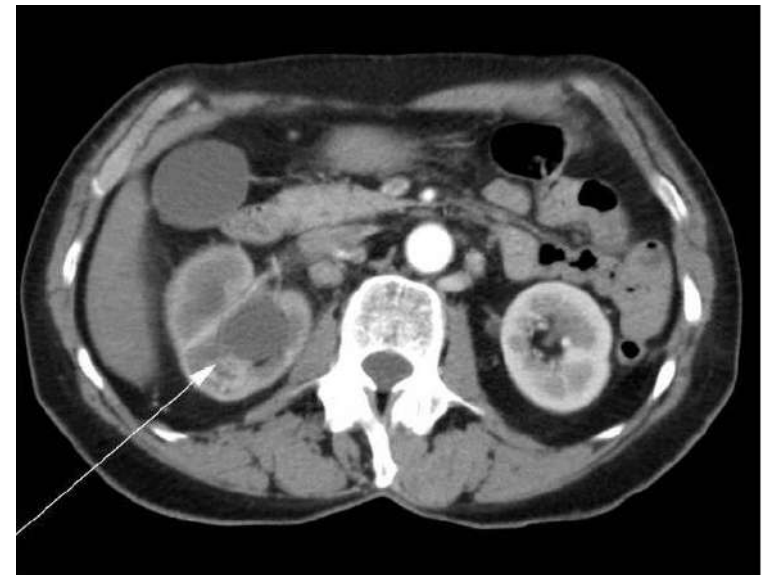
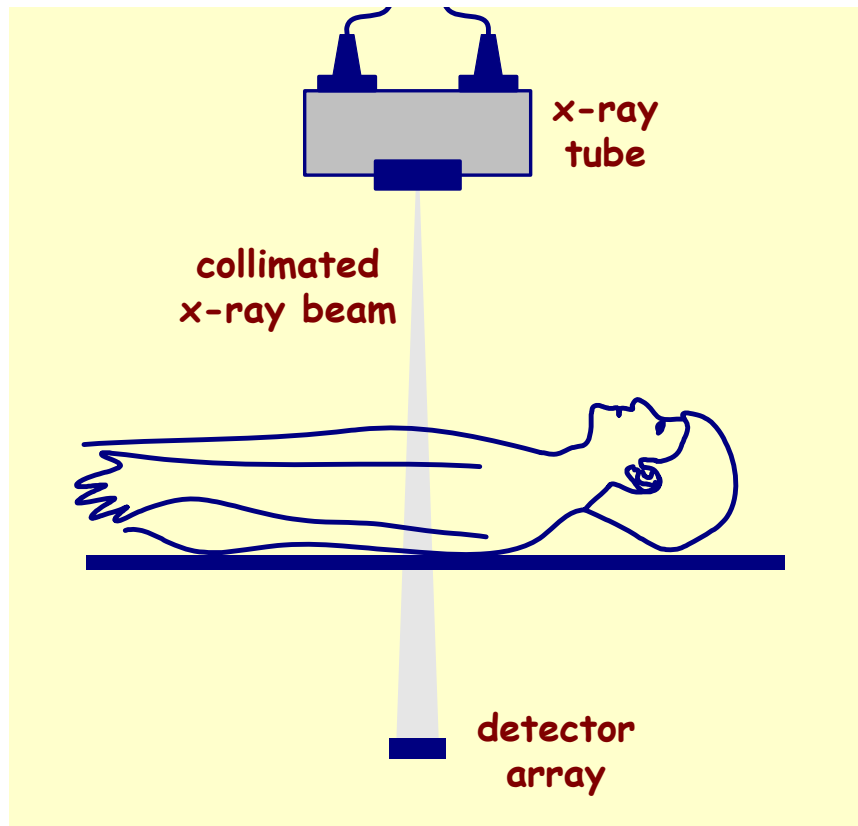
Resolution: 0.5 mm

Speed: seconds

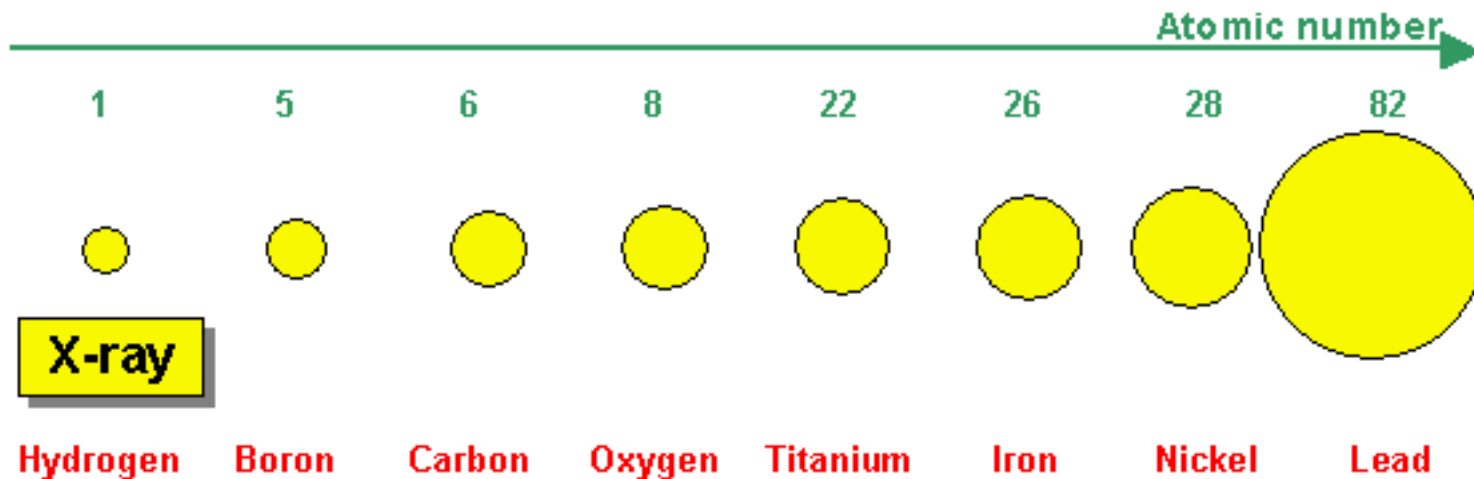
+ high resolution

- Radiation dose

Radiation dose per CT scan $\sim 3\text{-}5$ mGy



Probing matter with X-rays



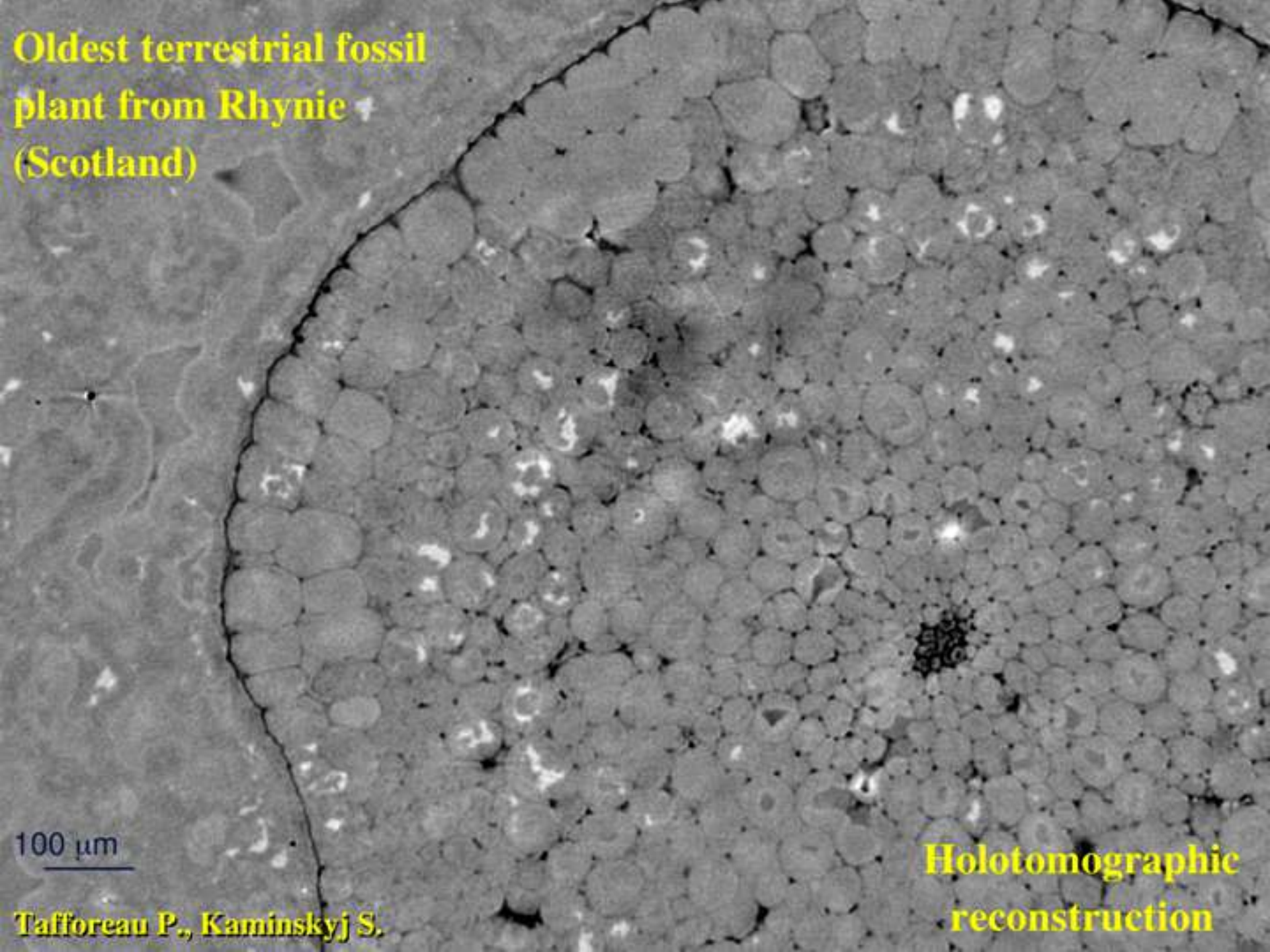
X-rays $\sigma_{abs} \propto Z^4 \cdot (\hbar\omega)^{-3}$

**Oldest terrestrial fossil
plant from Rhynie
(Scotland)**

100 μm

Tafforeau P., Kaminskyj S.

**Holotomographic
reconstruction**

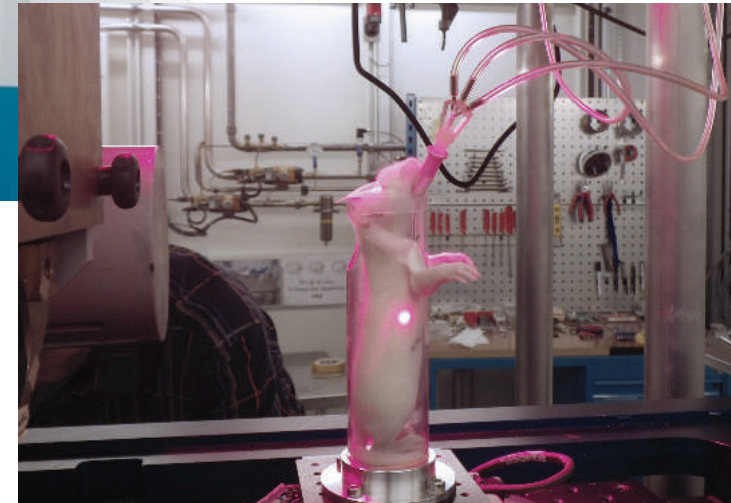


Medical and preclinical CT

The X-ray dose scales with resolution³

Resolution: 300 μm $\rightarrow$ 3 μm

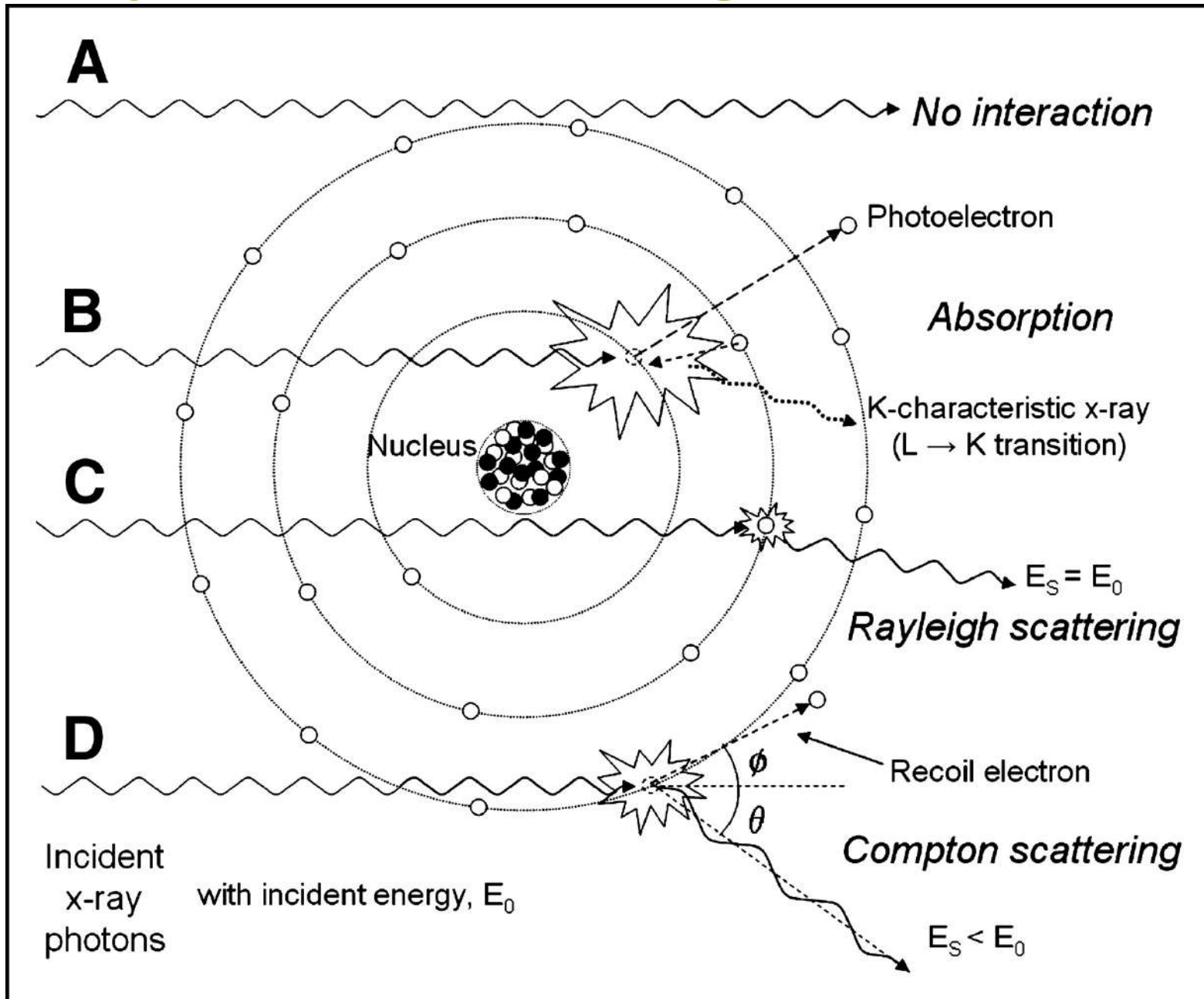
Dose: 3 mGy $\rightarrow$ 3 kGy



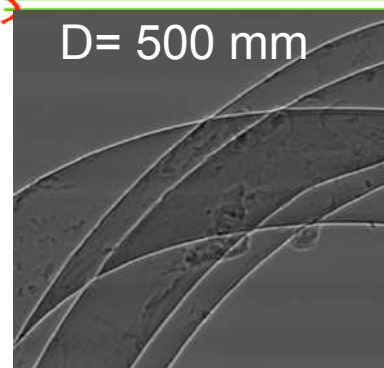
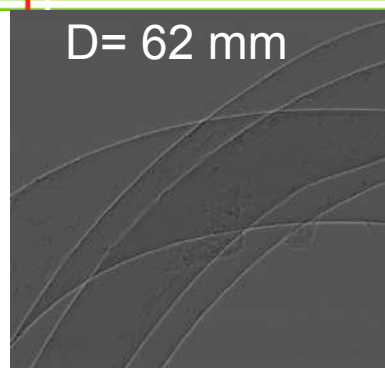
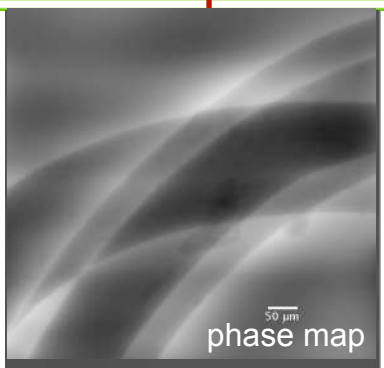
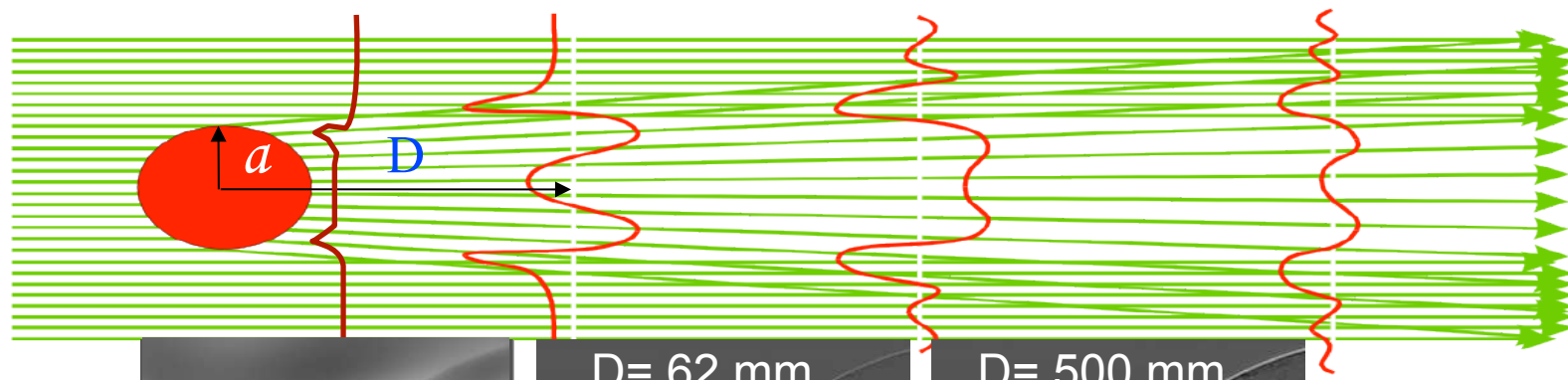
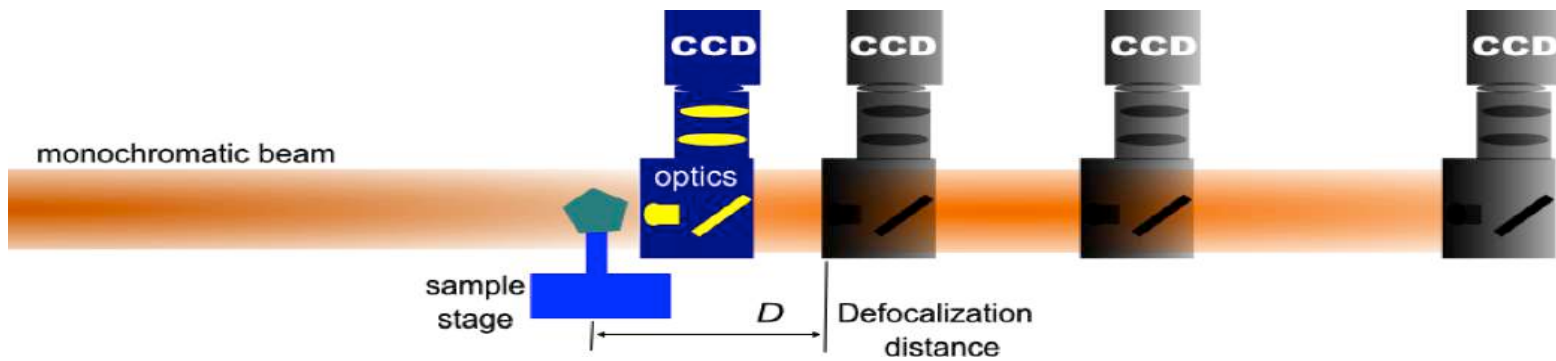
Absorbed dose:

$$D = \frac{E_{\text{abs}}}{m} = \frac{\Delta I h \nu}{V \rho} = \frac{(I_0 - I_1) h \nu}{\rho A \Delta x}$$

Absorption and scattering



Phase contrast in free space propagation



Three phase fibers

Fresnel diffraction
Partially coherent illumination

Cloetens et al. J. Phys. 1999

Rajmund.Mokso@maxiv.lu.se

Phase tomography

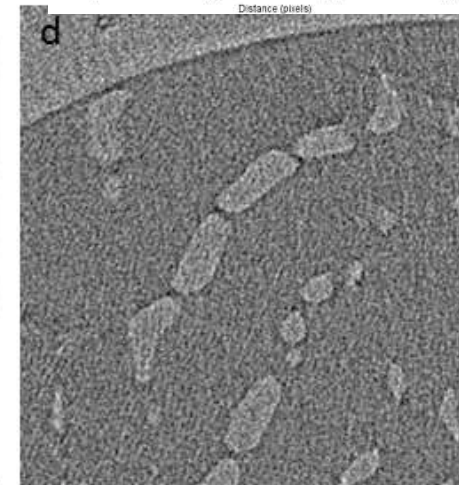
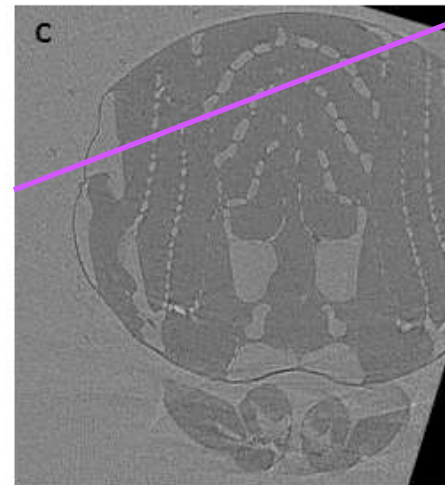
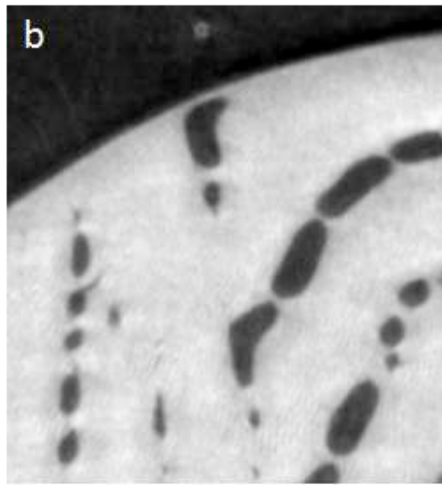
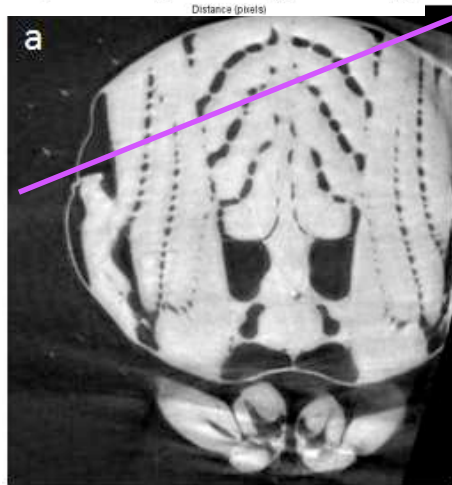
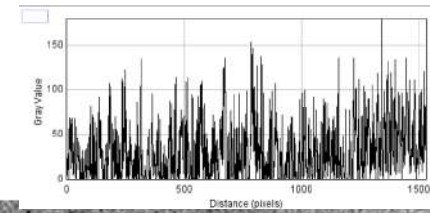
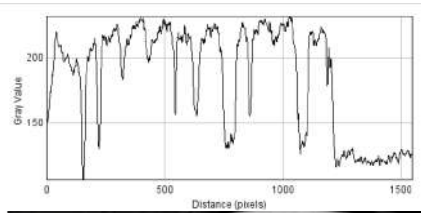
- Refractive index n

$$n = 1 - \frac{\delta}{\beta} + i \frac{\delta}{\beta}$$

$\delta \gg \beta$

fast tomography
30% of detector dynamic range

flight muscles of a fly



phase map

absorption & edge

Phase tomography

Phase contrast tomography

Mokso et al., J. Phys. D, 2013

Outline

I. Existing and soon available instruments

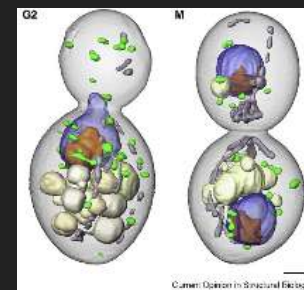
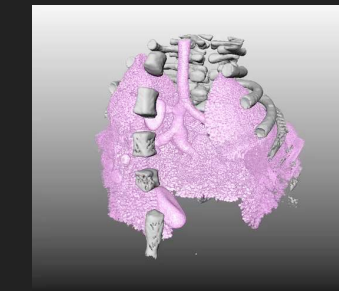
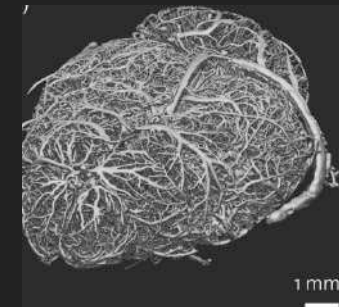
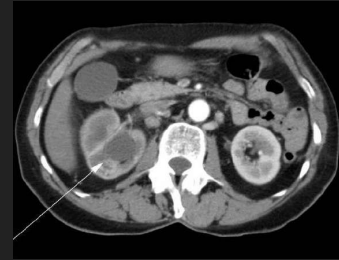
- I. NanoMAX, SoftiMAX, DanMAX

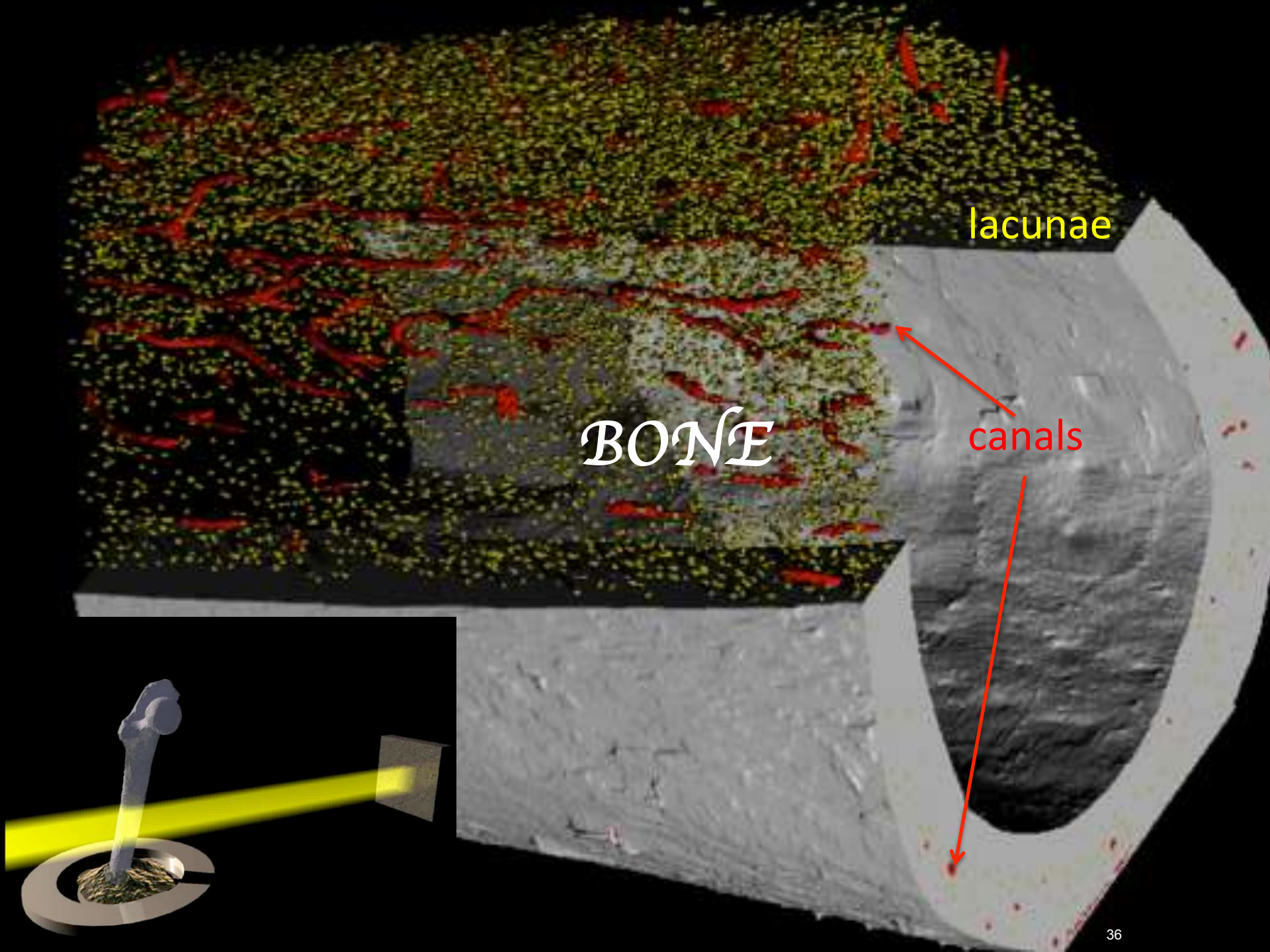
II. Biomedical tomographic microscopy

- I. The MedMAX project

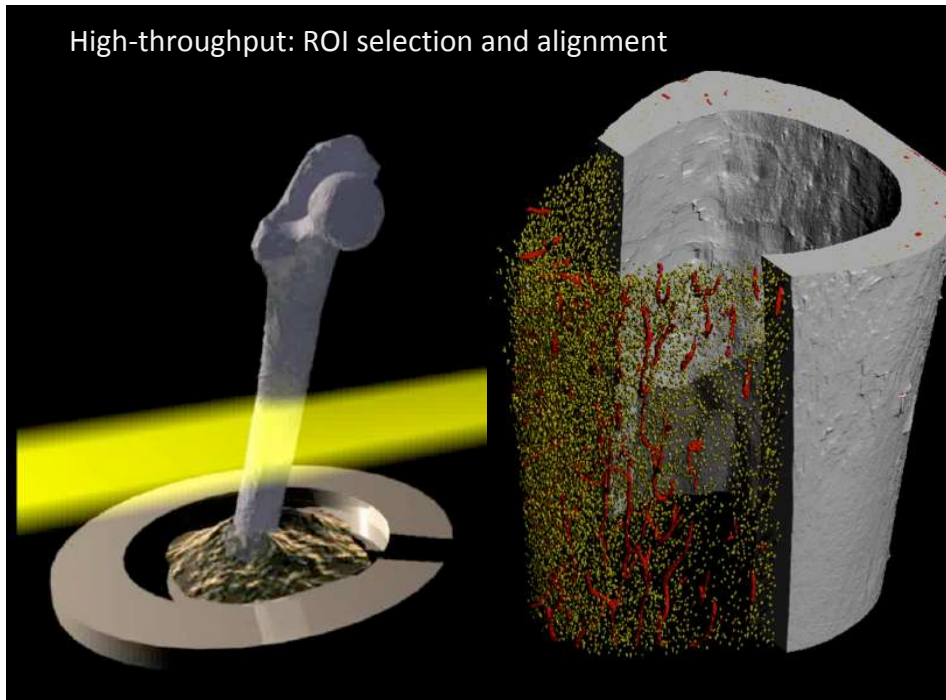
III. Examples

- I. In vivo small animals, various fixed tissues

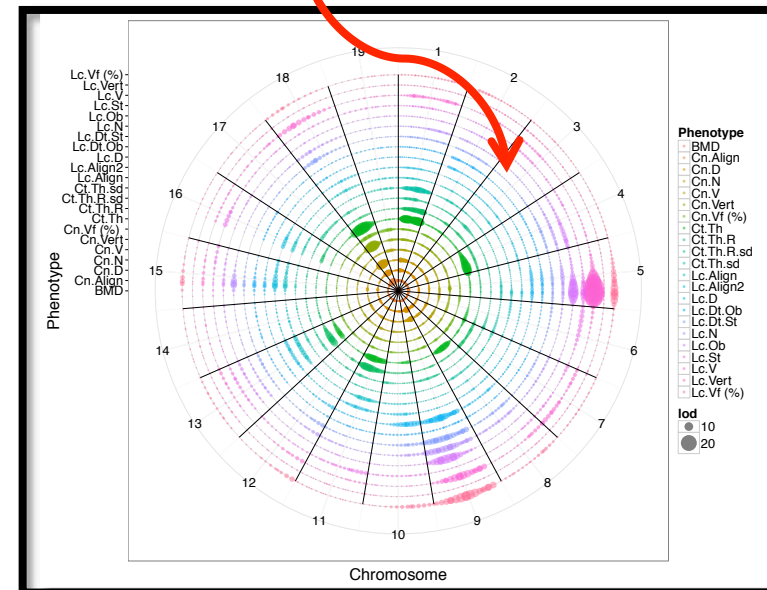
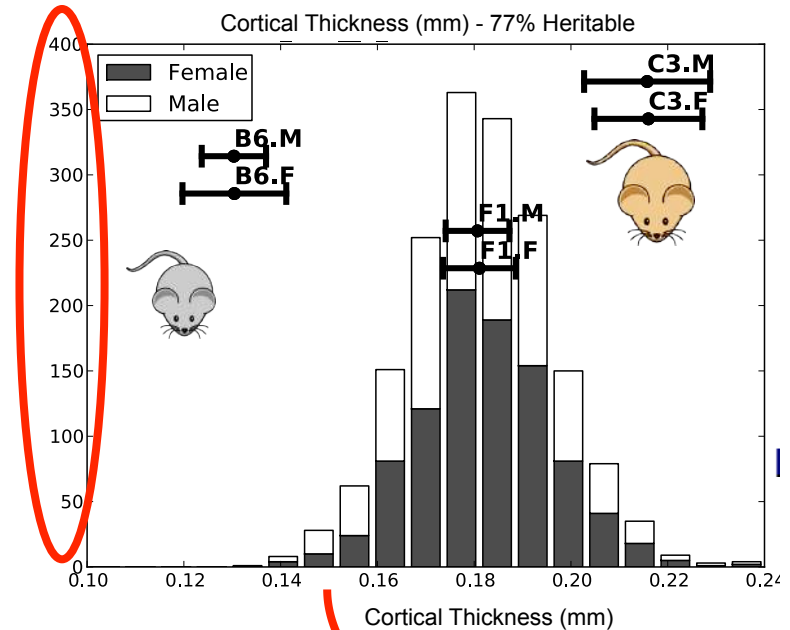




High-throughput imaging: from sample alignment to QTL analysis



Femur samples automatically aligned using goniometer and moved to region of interest using projections and image processing scripts



Alzheimer plaques

X-ray DPC scan, $(7.4\mu\text{m})^3$ voxel size



2-photon fluorescence, $2\times 2\times 4\mu\text{m}^3$ voxel size

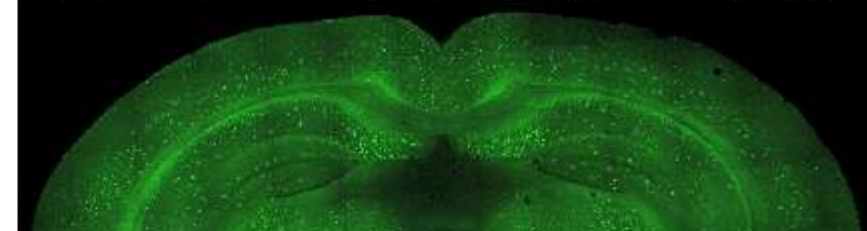


Alzheimer plaques

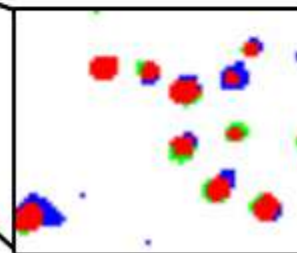
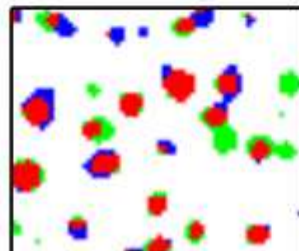
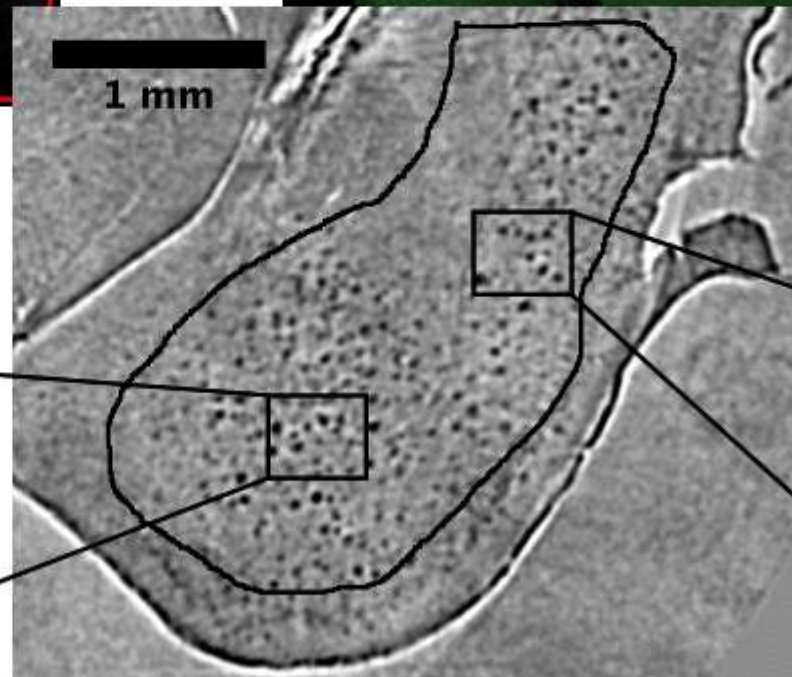
X-ray DPC scan, $(7.4\mu\text{m})^3$ voxel size



2-photon fluorescence, $2\times 2\times 4\mu\text{m}^3$ voxel size

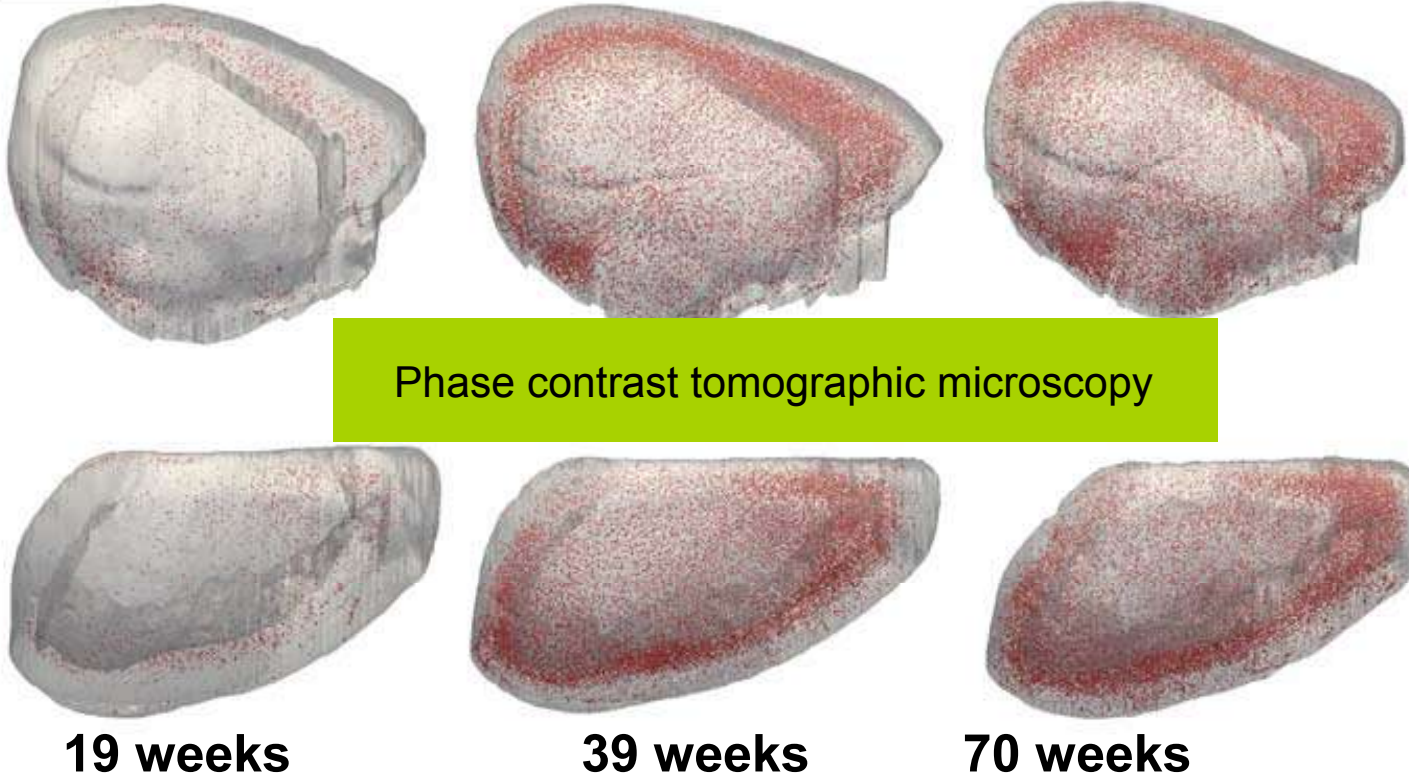


MedMAX : sub-micrometer resolution 3D histology



Alzheimer plaques

2 mm

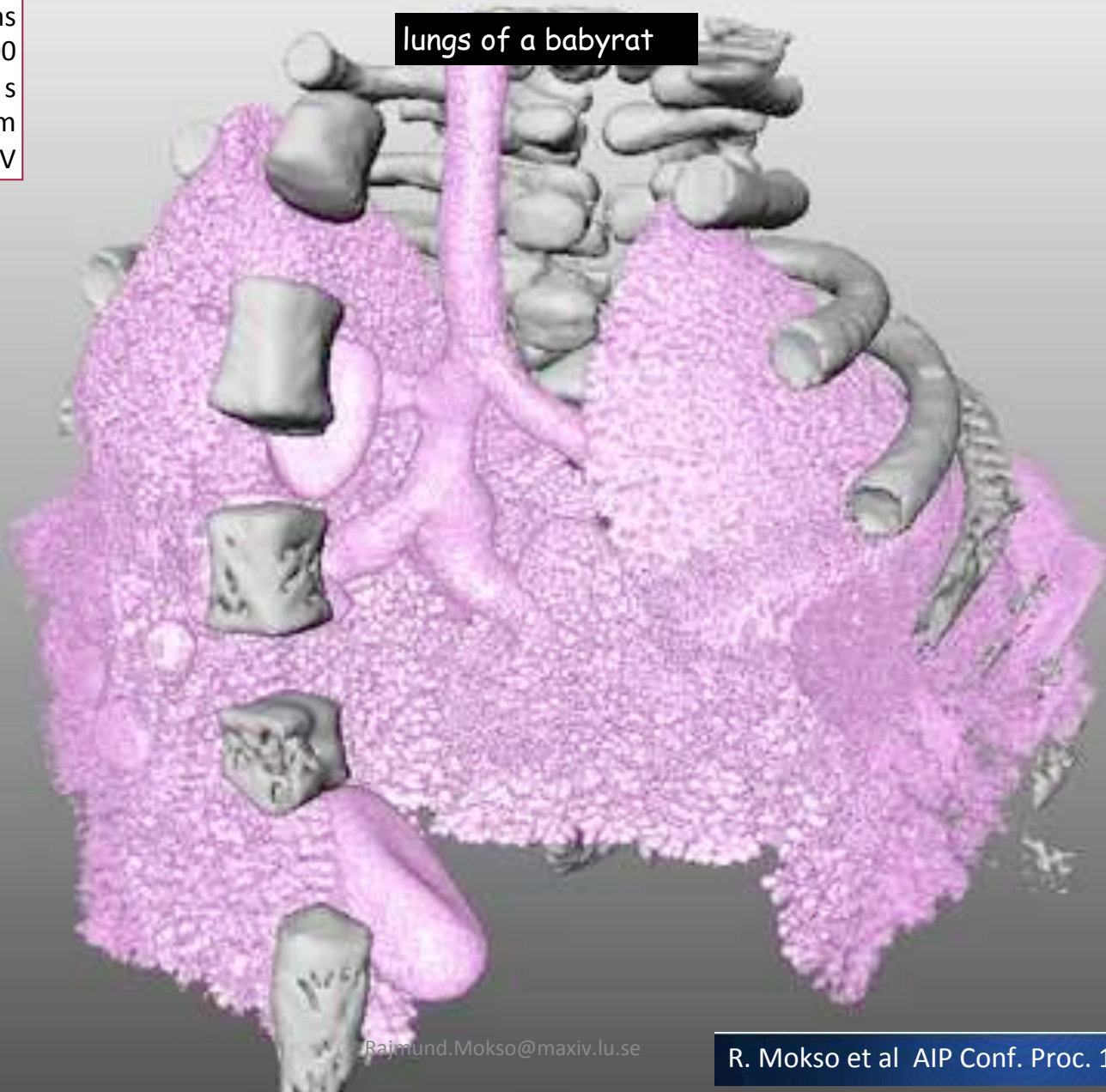


- Full 3D structural analysis is possible:
 - Size distribution
 - Plaque load (volumetric density)
 - Nearest neighbour distance (and much more)

In-vivo lung imaging

exposure time= 1.1 ms
projections= 500
total scan time= 0.57 s
voxel size= 11 μ m
E= 20 keV

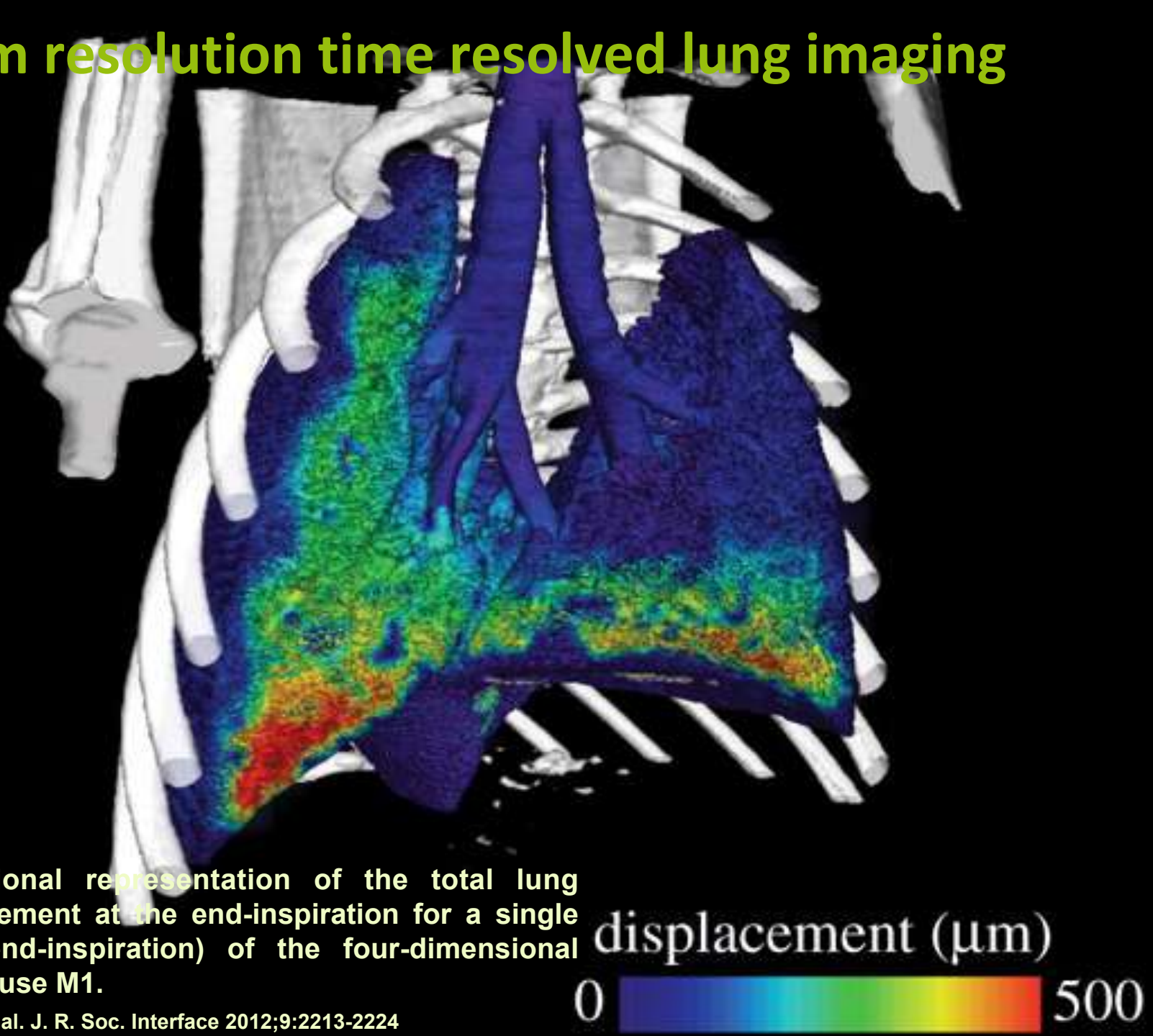
lungs of a babyrat



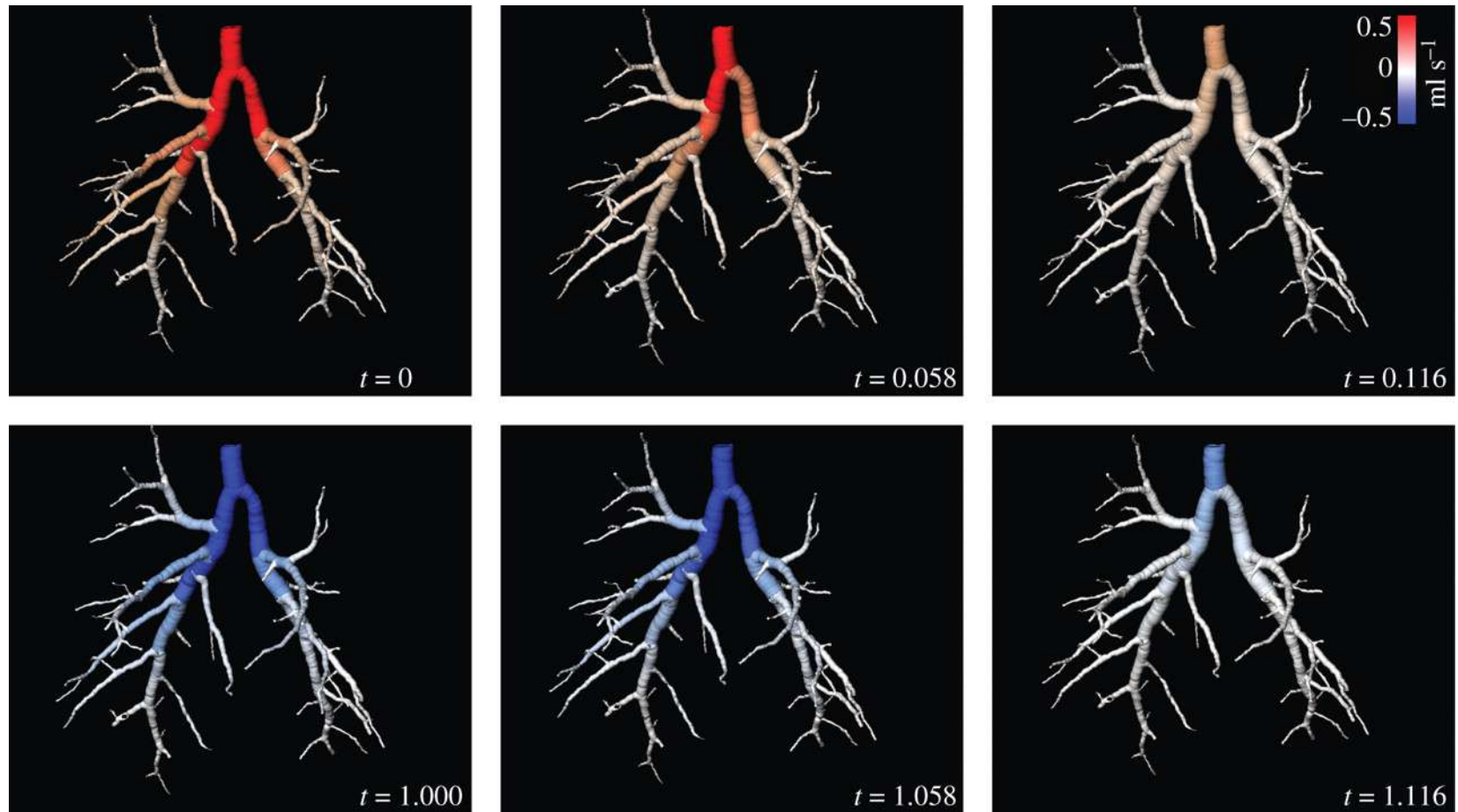
Medium resolution time resolved lung imaging

Three-dimensional representation of the total lung tissue displacement at the end-inspiration for a single time point (end-inspiration) of the four-dimensional dataset for mouse M1.

Stephen Dubsky et al. J. R. Soc. Interface 2012;9:2213-2224



Distribution of air flow through an airway tree in rabbit lungs

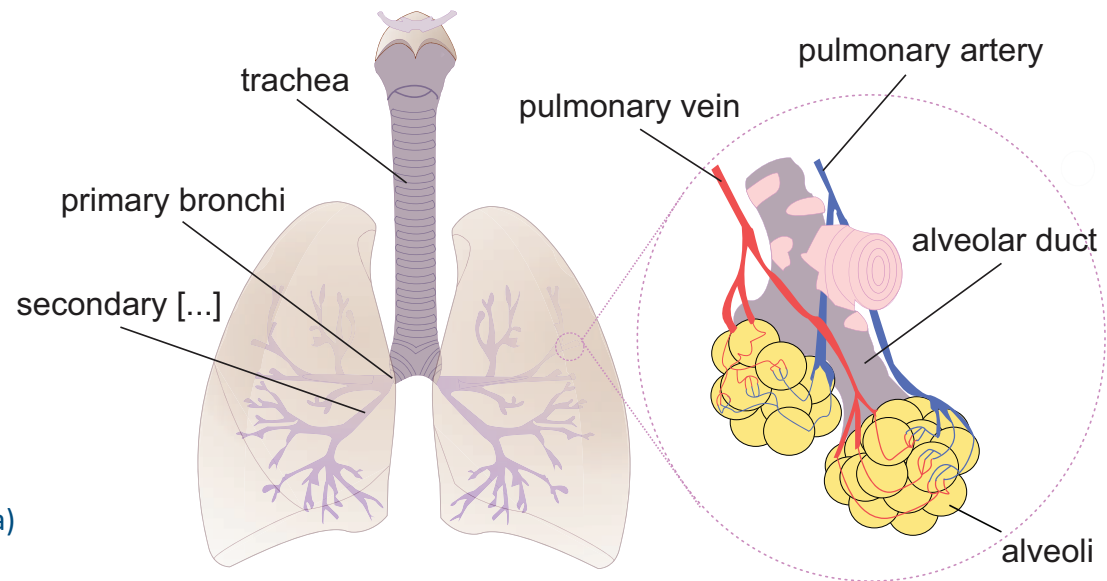


Stephen Dubsy et al. J. R. Soc. Interface
2012;9:2213-2224

Dynamic imaging of lungs at the micrometer scale: motivation

Ventilator-induced lung injury (VILI)

- Overextension of lung tissue in certain lung regions (mechanical damage, biotrauma)
- Still unclear how ventilation induces its deleterious effect [4]
- Hypothesis: local strains in the alveolar wall cause hotspots (overstretching regions)



► Human lungs (Source: Wikipedia)

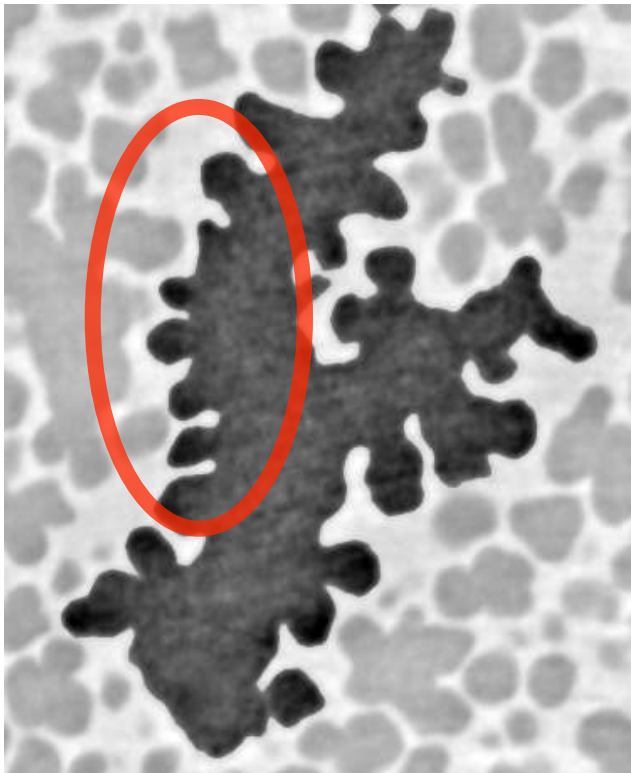
[4] Rausch, S. M. K., Haberthür, D., Stampanoni *et al.*, Ann Biomed Eng **39** (11), 2835 (2011).

Lung tissue quantification

Challenges in lung tissue analysis

- How to detect non-linear and regional changes in the lung?
- How to quantify them?

G. Lovric , PhD thesis



$$\Delta p = 5 \text{ cmH}_2\text{O}$$

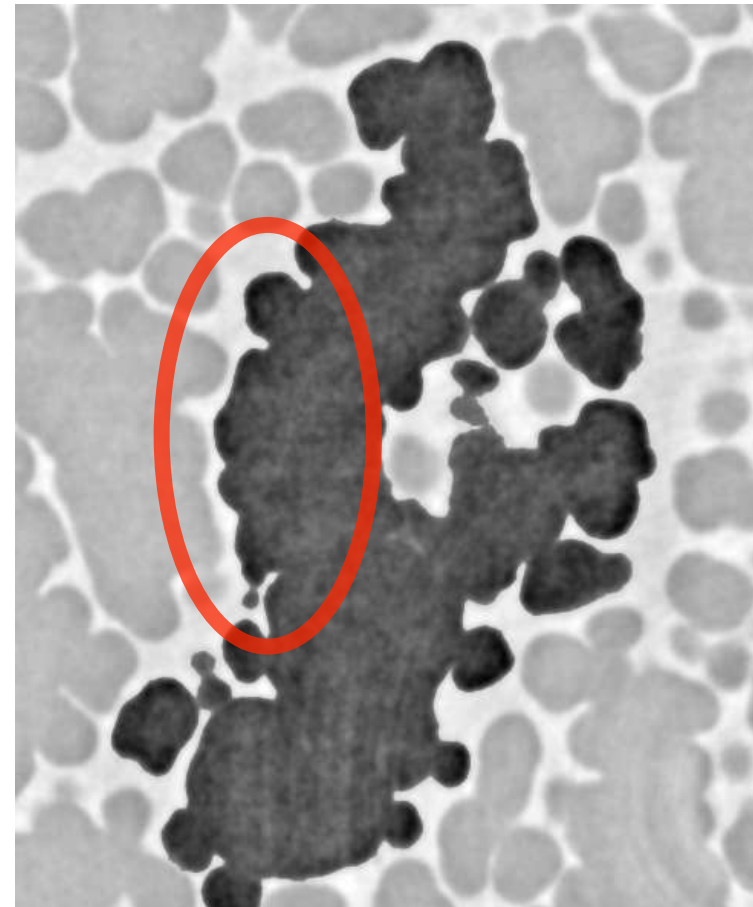


Image quality vs. radiation damage

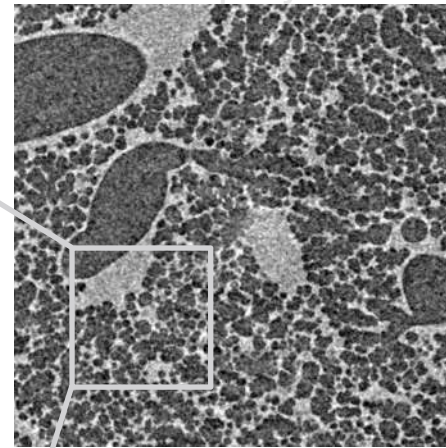
2.9 μm pixel-size optics

901 projections

901 projections

901 projections

361 projections



Scan time

0.24 s

Scan time

0.17 s

CNR

2.3

CNR

1.5

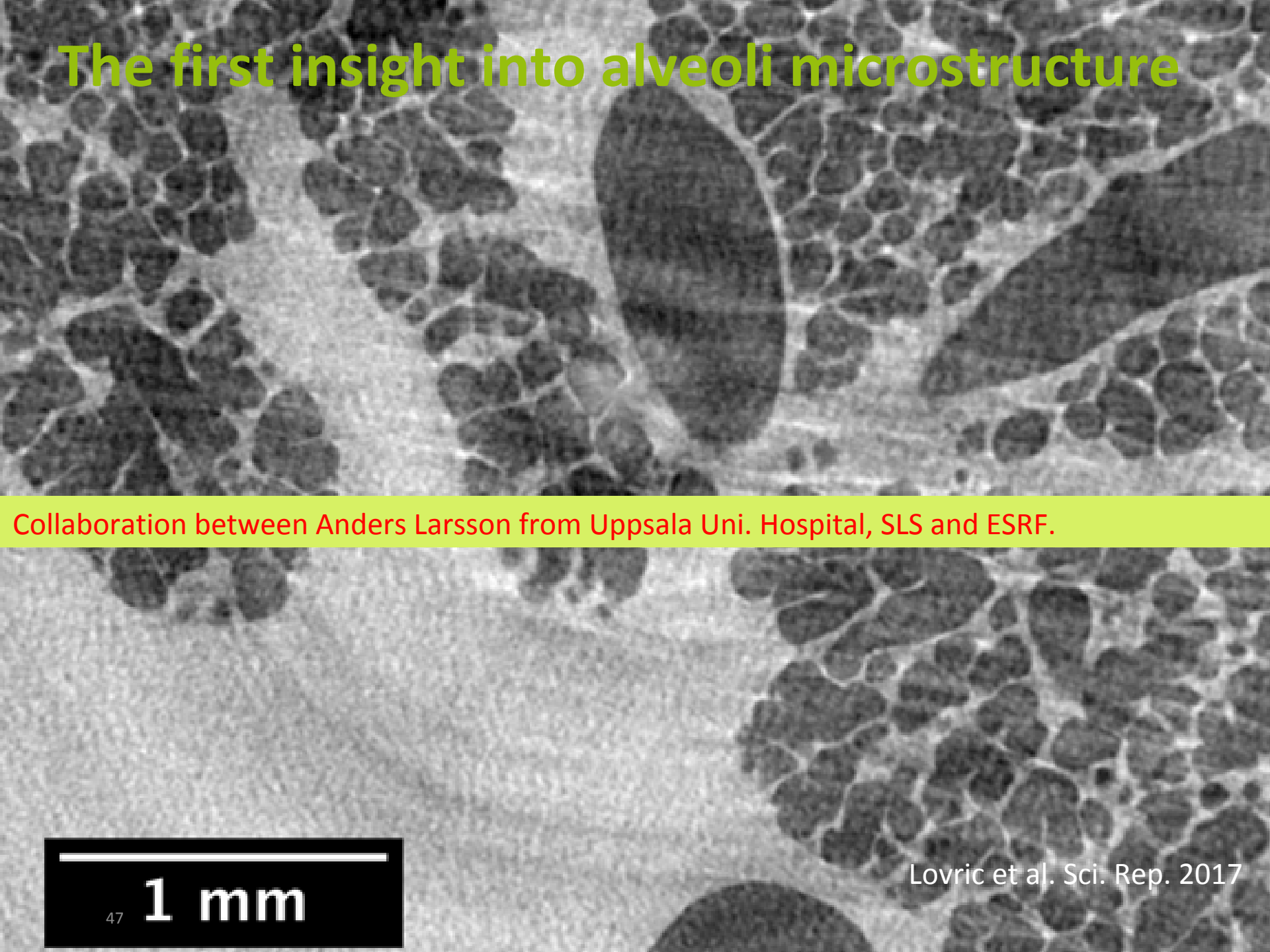
Entrance dose

12 Gy

Entrance dose

9 Gy

The first insight into alveoli microstructure

A grayscale micro-CT scan of lung tissue showing the intricate, honeycomb-like structure of alveoli. The alveoli are represented as dark, rounded clusters separated by a lighter, more fibrous network of connective tissue. The overall texture is highly porous and complex.

Collaboration between Anders Larsson from Uppsala Uni. Hospital, SLS and ESRF.

1 mm

47

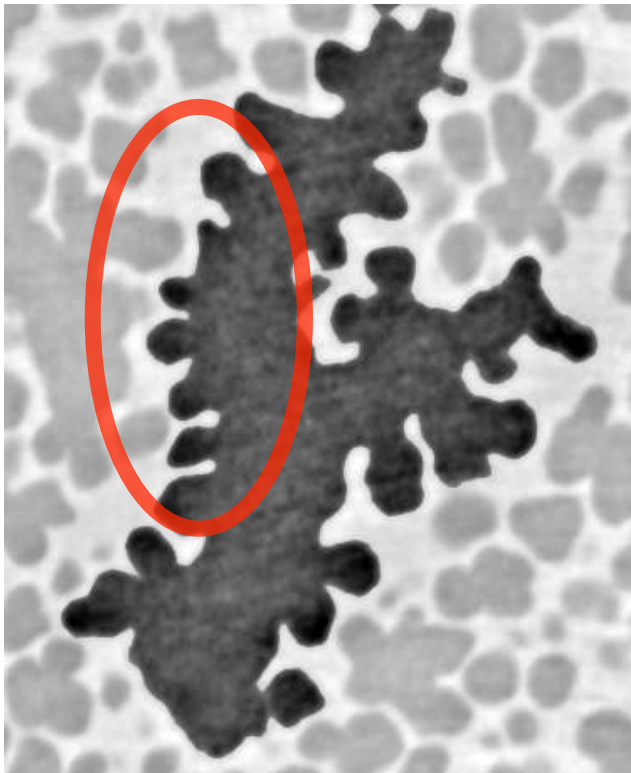
Lovric et al. Sci. Rep. 2017

Lung tissue quantification

Challenges in lung tissue analysis

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- How to quantify them?

G. Lovric , PhD thesis



$$\Delta p = 5 \text{ cmH}_2\text{O}$$

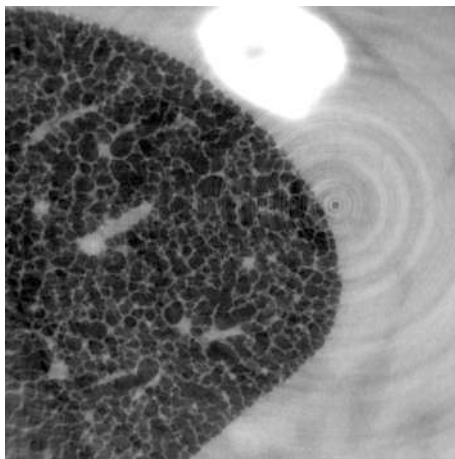


Alveoli dynamics quantification

A detailed insight into the lung

- Apply *quantification* and *labeling* tools, previously used for foam and bone data [14] and dendritic microstructures [15]

Original tomographic slice



1 mm

Thickness map



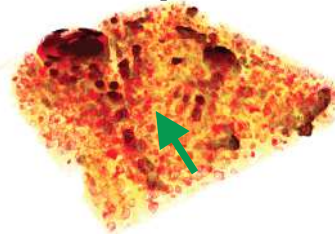
Curvature



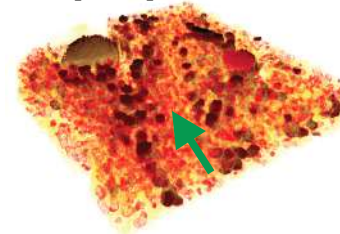
[3.5 x 3.5 x 0.6 mm³]

[3.5 x 3.5 x 0.6 mm³]

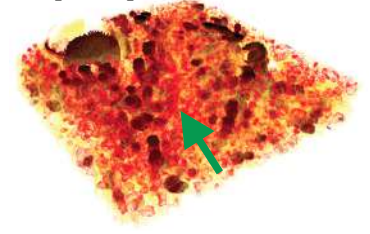
[3.5 x 3.5 x 0.6 mm³]



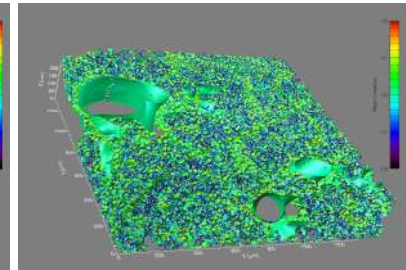
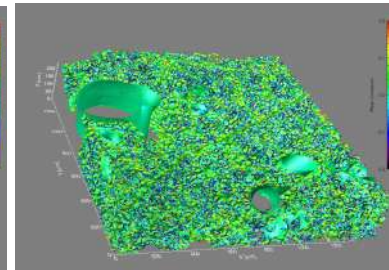
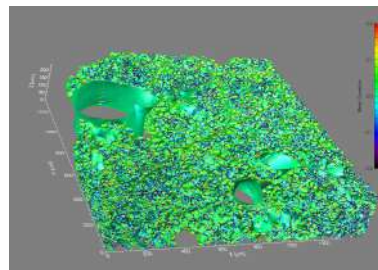
25 cmH₂O



30 cmH₂O

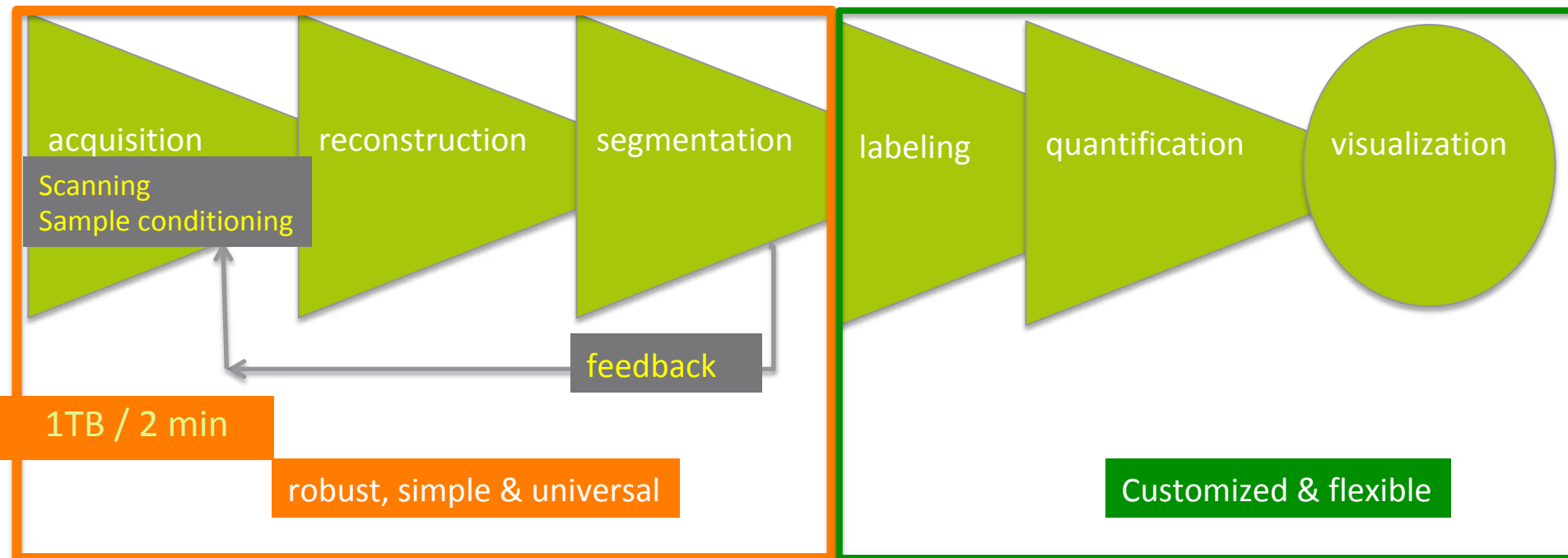
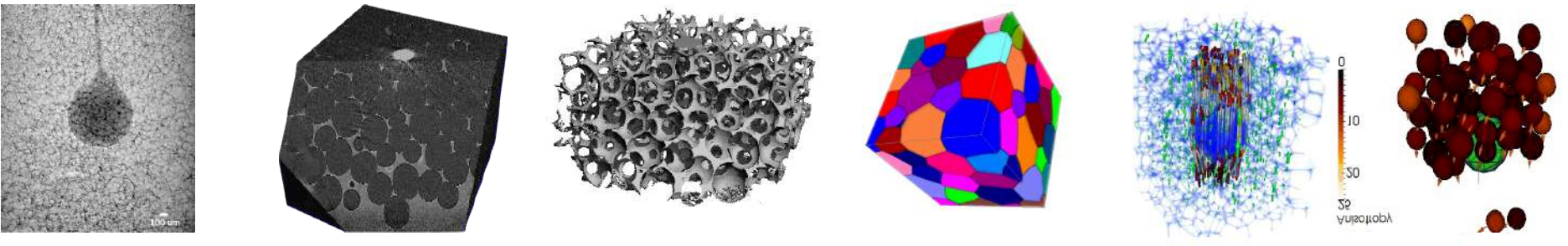


35 cmH₂O

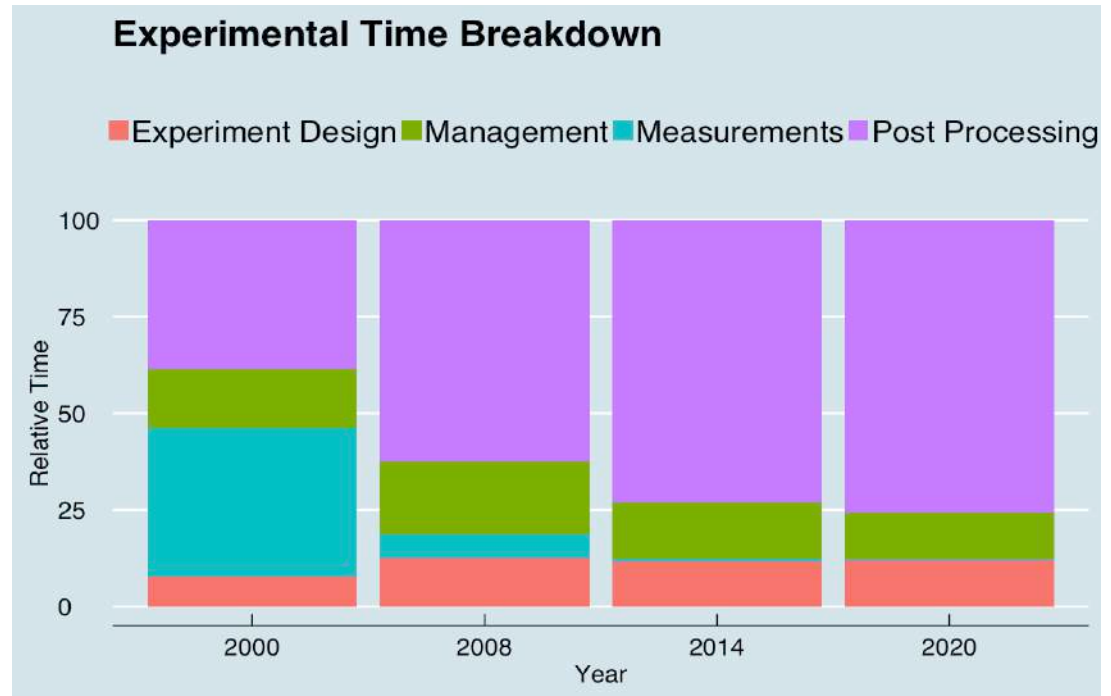
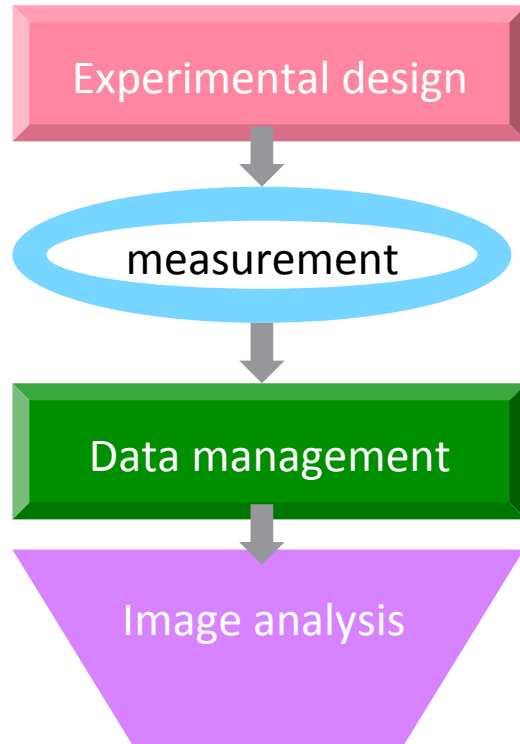


G. Lovric , PhD thesis, Lovric et al. PLOS On2017e

Image-based quantitative information retrieval



High throughput quantitative imaging

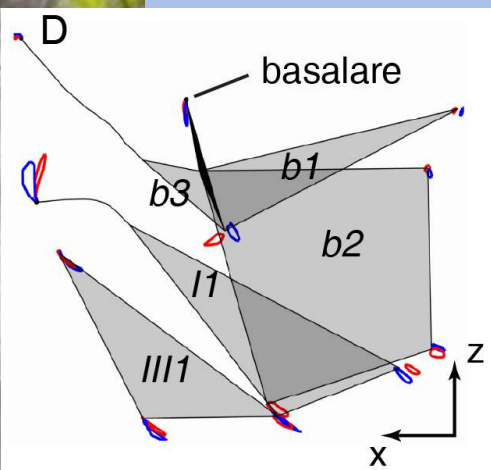
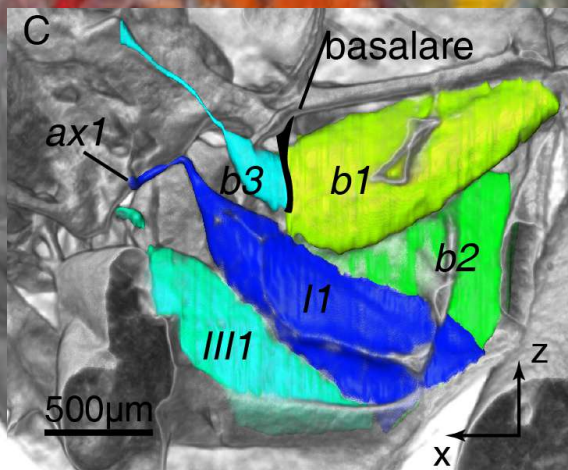
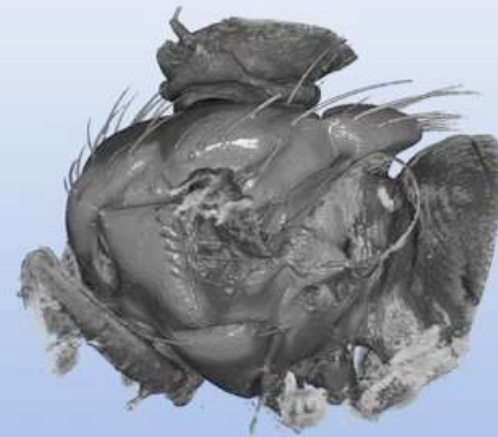


Courtesy of Kevin Mader

1TB / 2 min

Survival ~ 3 s

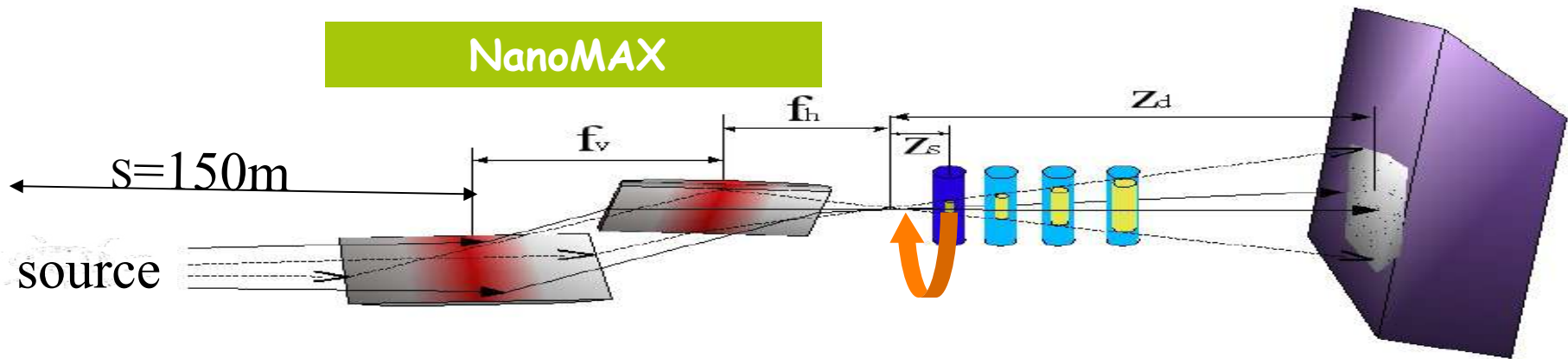
of insect



3 kHz effective temporal
resolution
10-20 μm spatial
resolution

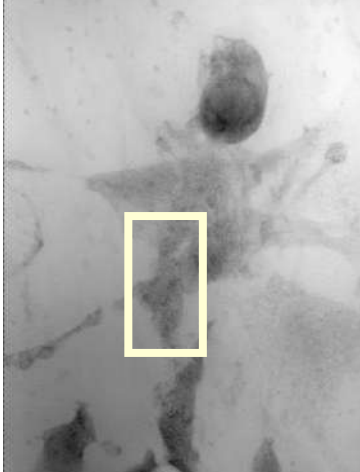
Nanoimaging, Fluorescence spectroscopy

NanoMAX



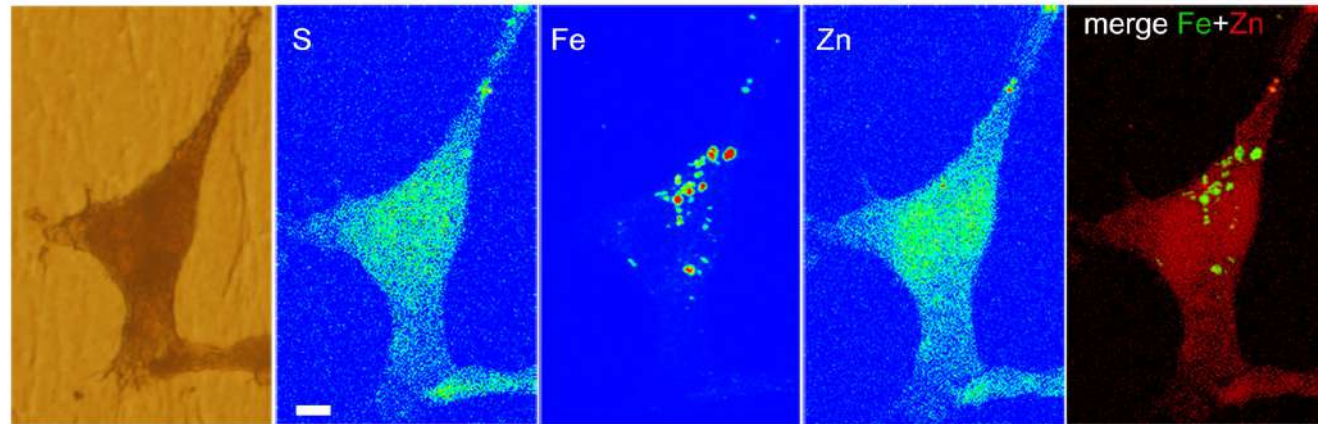
$10\ \mu\text{m}$

A neuron cell phase image



Mokso, Cloetens et al. , APL 2007

Chemical element distribution



Ortega. , Mol. Neurobiol. 2016

Holographic nanotomography of human cerebellum

FULL PAPER

Nanotomography

ADVANCED
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www.advancedscience.cor

Hard X-Ray Nanoholotomography: Large-Scale, Label-Free, 3D Neuroimaging beyond Optical Limit

Anna Khimchenko, Christos Bikis, Alexandra Pacureanu, Simone E. Hieber, Peter Thalmann, Hans Deyhle, Gabriel Schweighauser, Jürgen Hench, Stephan Frank, Magdalena Müller-Gerbl, Georg Schulz, Peter Cloetens, and Bert Müller*

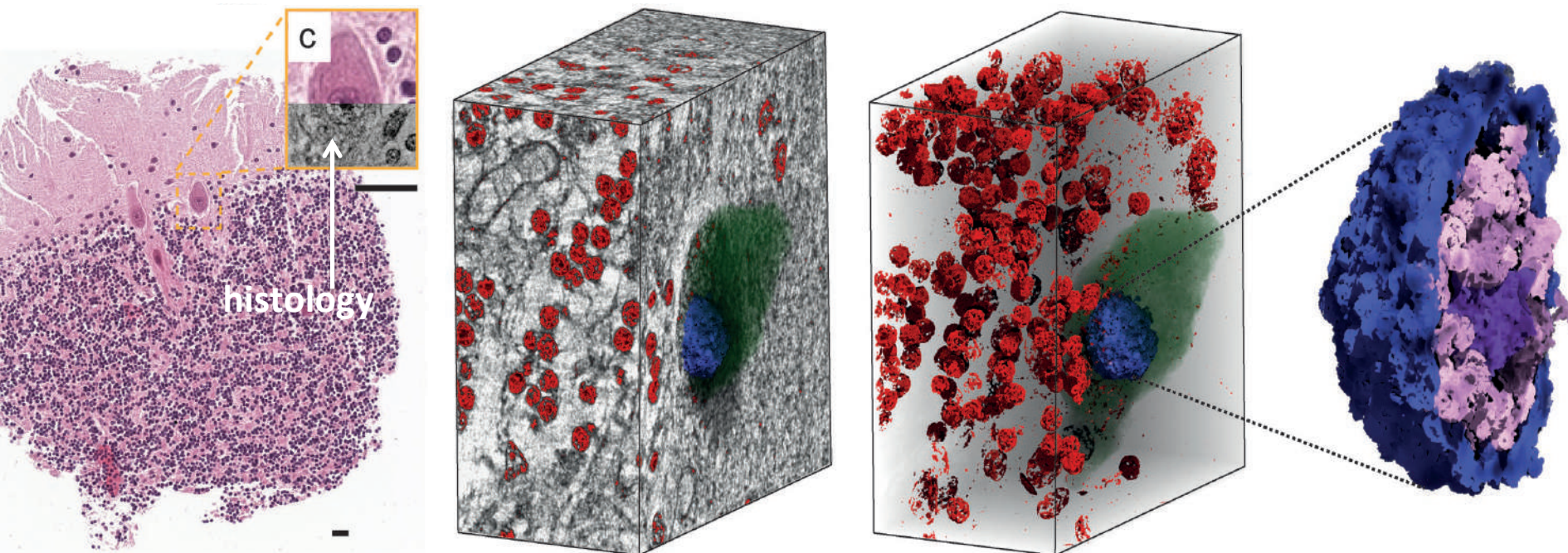
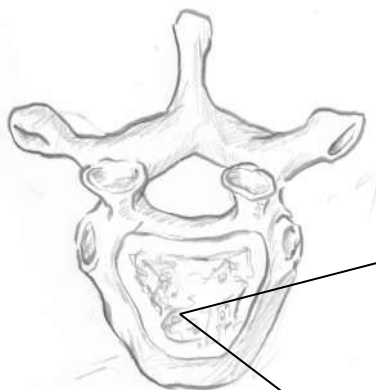


Figure 5. A 3D rendering of (sub)cellular structures within a Purkinje cell, measured with an effective pixel size of 100 nm. Region-growing segmentation allows for the extraction of cell soma (green), nuclear envelope (blue), nuclear content (pink), and nucleolus (violet). Intensity thresholding (red) enables the discrimination of granule cell nuclei. Average diameter of Purkinje cell soma: 54 μm ; average diameter of Purkinje cell nucleolus 3.5 μm .

Bone nanostructure (Small Angle X-ray scattering)

Human vertebra

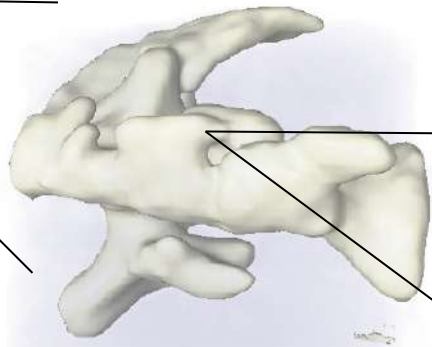


Trabecular bone



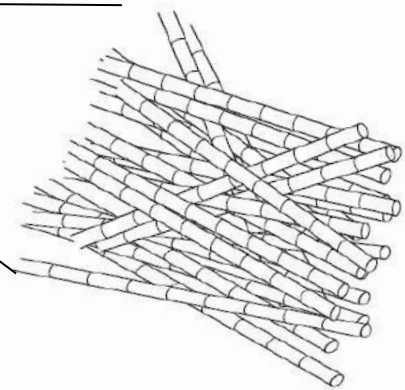
1 mm

Single trabecula
mm scale

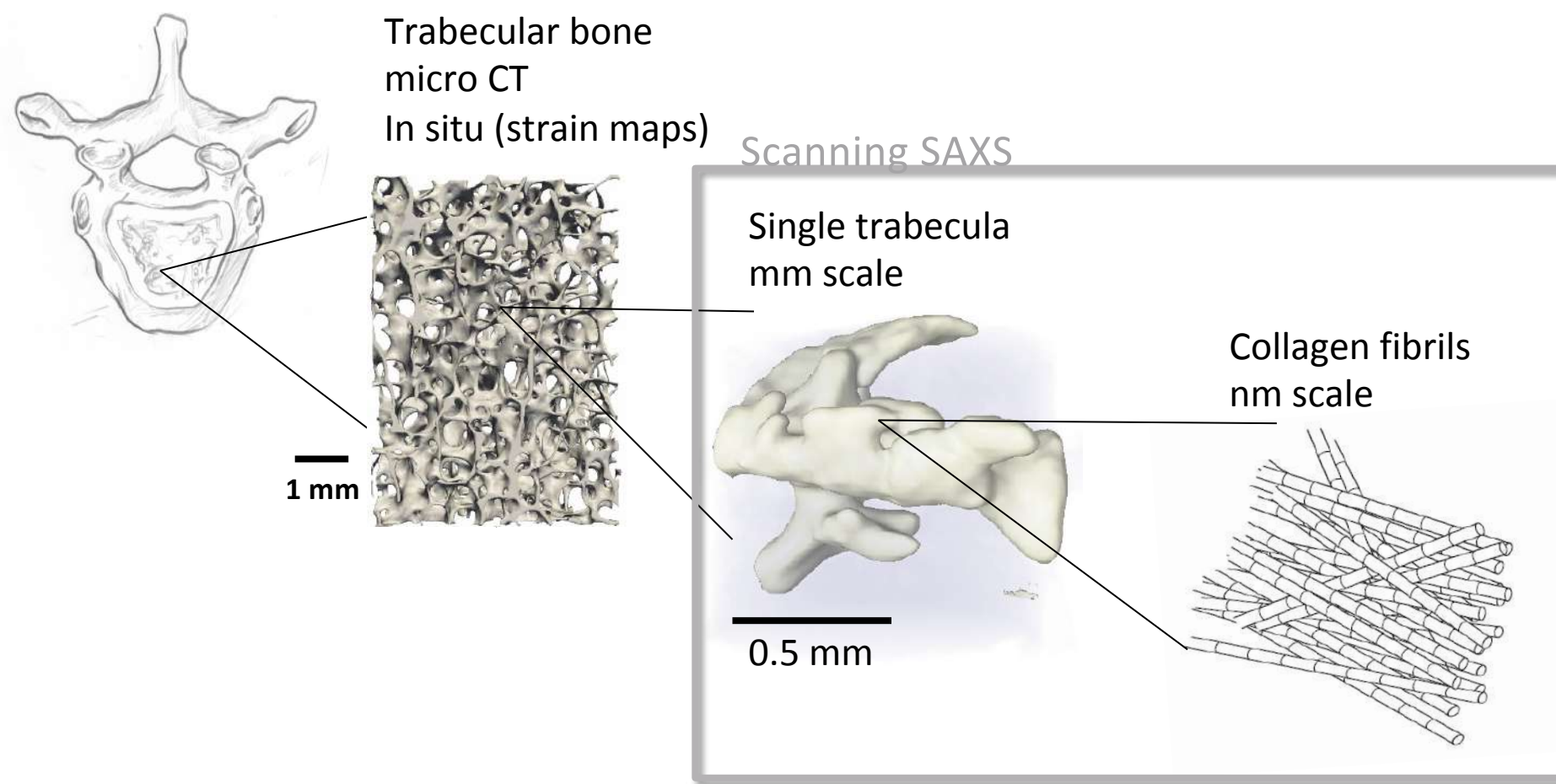


0.5 mm

Collagen fibrils
nm scale



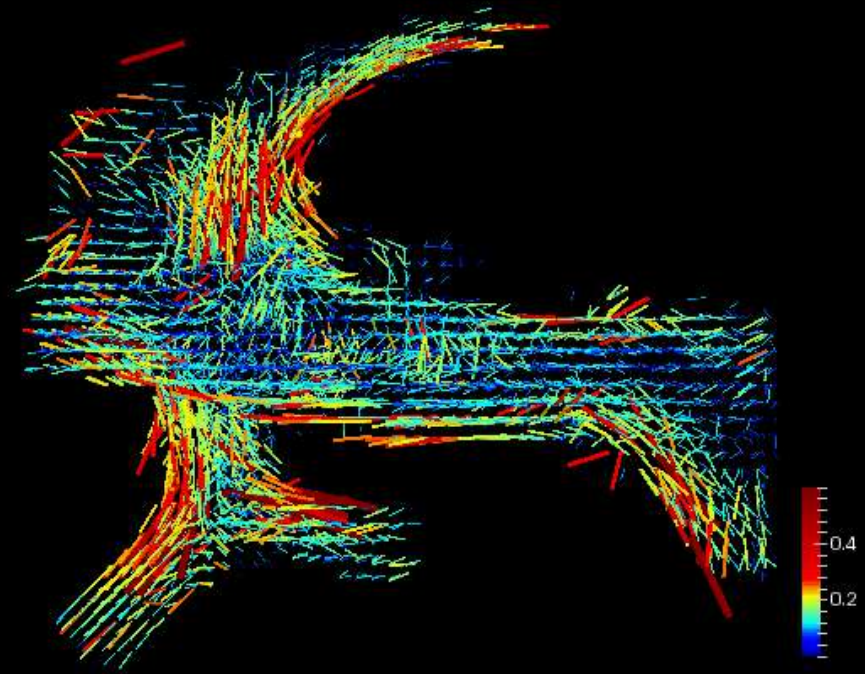
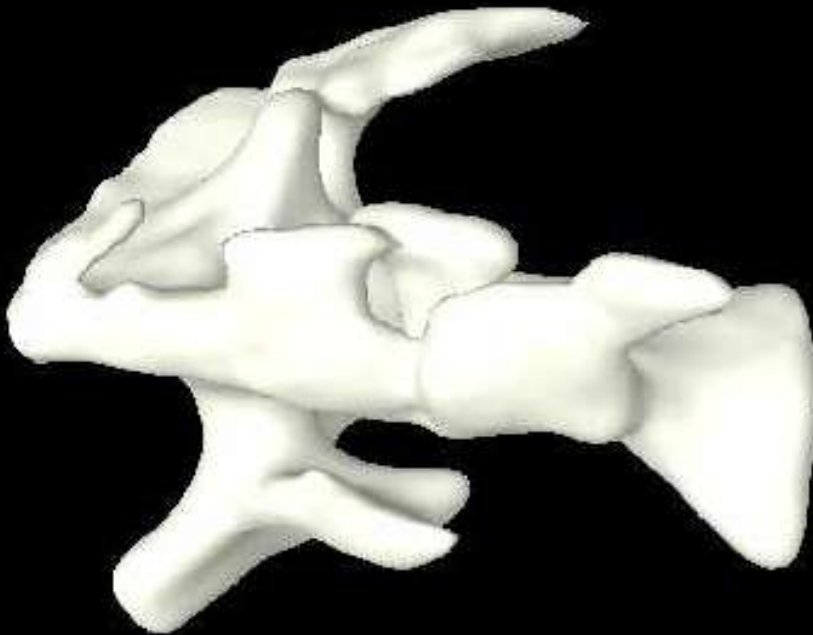
Bone nanostructure (Small Angle X-ray scattering)



Bone nanostructure (SAXS tensor tomography)

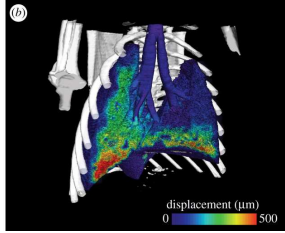


CoSAX : in operation 2019
ForMAX: project phase 2017-2021



Hierarchical imaging of tissues

Medical imaging and therapy
(in vivo, medium resolution $>30\text{ }\mu\text{m}$, low dose, large beam, large FOV)



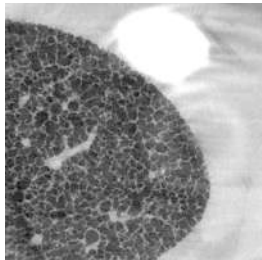
Longitudinal in vivo

The whole organ

2 mm

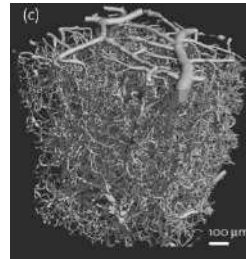


Down to a micrometer resolution
(rarely in vivo, high resolution $\sim 1\text{ }\mu\text{m}$, higher dose, small FOV)



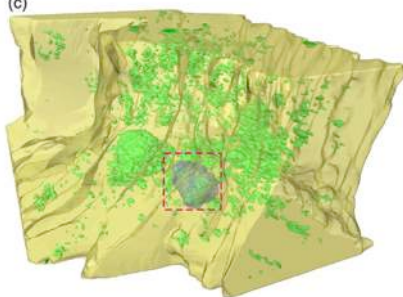
Accute in vivo

Zooming into organs



Nano-scale imaging beamlines
(fixed tissue, resolution $\sim 100\text{ nm}$, high dose, small samples $\sim 100\text{ }\mu\text{m}$)

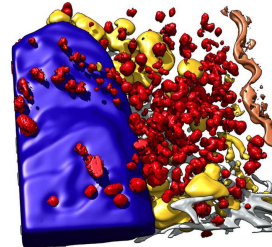
Bronchial wall



(d)

Fixed tissue

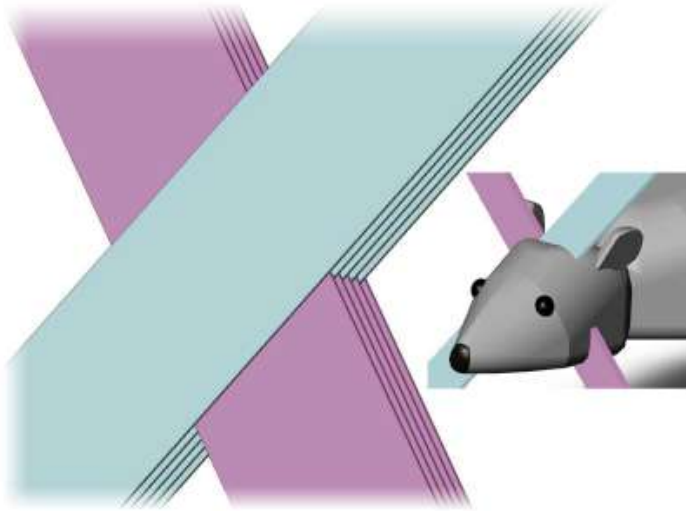
Cell level



3 orders of magnitude in spatial resolution



Microbeam radiation therapy



Microbeam irradiated normal CNS of weaned piglets (1.5cm x 1.5cm ~28 mm-wide beams ~210 mm on center, 625 Gy). The histological sections look normal, except for "stripes" due to the dropout of neuronal/astroglial nuclei. This sharp spatial fractionation is preserved throughout the cerebellum. No tissue necrosis, hemorrhage or demyelination was observed.

E. Bräuer-Krisch, et al. New irradiation geometry for microbeam radiation therapy, *Phys Med Biol* 50 (2005) 3103-3111.

A close-up photograph of numerous pink frosting swirls arranged in a grid-like pattern on a dark, textured surface. The swirls are uniform in size and shape, each with a distinct peak and ridges. A semi-transparent dark gray rectangular box is centered over the image, containing the text "Thank you" in a white, elegant script font.

Thank you